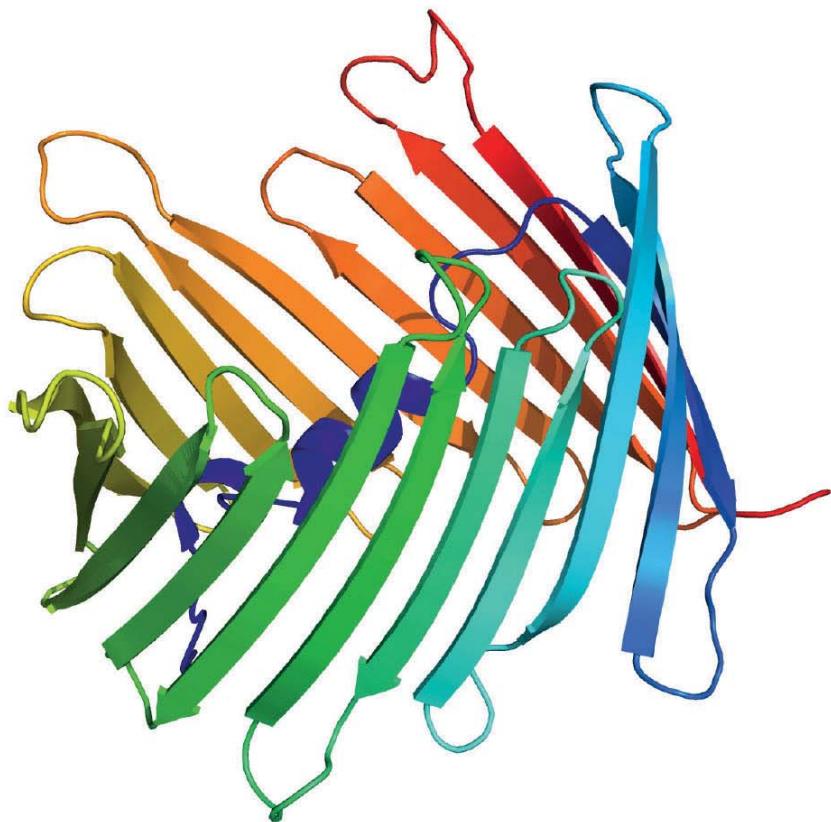


Christian D. Schmidt

# NMR spectroscopic investigations on VDAC



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## NMR spectroscopic investigations on VDAC



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# NMR spectroscopic investigations on VDAC

## Dissertation

zur Erlangung des mathematisch-naturwissenschaftlichen Doktorgrades

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*Für meine Familie*



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# Abstract

This thesis is improving the understanding of the gating process of the voltage dependent anion channel (VDAC). VDAC is the most abundant protein in the outer mitochondrial membrane of eukaryotes and has a plethora of different functions there. It is the main pathway for metabolites that are going in and out the mitochondria, is interacting with many proteins in the OMM and probably also involved in the processes around cell death. Ever since its discovery in the 1970s and especially since the publication of three high resolution structures of VDAC in 2008 the mechanism of the voltage gating of the protein is a scientific question.

In this thesis, the assignment of hVDAC1 in liquid state NMR is further improved and  $C_\beta$ -assignments are determined.

Based on a vastly improved NOE network and previously recorded  $R_1/R_2$  relaxation data, a 3D solution state structure of E73V hVDAC1 was determined. This structure is providing evidence that the N-terminal  $\alpha$ -helix of the protein is tightly connected to the  $\beta$ -barrel and has a kink feature, that has previously been described only by an X-ray 3D structure. The  $\beta$ -barrel shows a significant elliptic deformation that is resembling a previously determined global deformation mode of the pore and is caused by the altered lateral pressure of the surrounding micelles with respect to phospholipid membranes.

Furthermore, a hVDAC1 mutant that is stabilizing a closed state of the protein was developed and is described. This “Quintuple hVDAC1” is exhibiting properties previously determined for N-terminal deletion constructs in solid state NMR and BLM measurements, but can – contrary to the deletion constructs – still be investigated by liquid state NMR measurements. Several lines of evidence point to and suggest a partial movement of the N-terminal  $\alpha$ -helix in this mutant. Based on the results, two possible models for a closed state of hVDAC1 are developed and discussed. The discussion is also covering differences between detergent micelles and lipid bilayer reconstituted VDAC and the respective consequences on the stability of the samples.

# Publication list

This thesis is based on the following publication:

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# Nomenclature

ADP	adenosine 5'-diphosphate
AFM	atomic force microscopy
APS	ammonium persulfate
APSY	automated projection spectroscopy
ATP	adenosine 5'-triphosphate
BLM	black lipid membrane / bilayer lipid membrane
CEST	chemical exchange saturation transfer
CPMG	Carr–Purcell–Meiboom–Gill
CuX <sub>2</sub> Ph	copper phenantroline
DEST	dark-state exchange saturation transfer
DMPC	dimyristoyl-phosphatidylcholine
DPhPC	diphytanoyl-phosphatidylcholine
DTT	dithiothreitol
EDTA	ethylenediaminetetraacetic acid
EM	electron microscopy
GRS	glycine rich sequence
HDL	high-density lipoprotein
HEPES	4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid
HK	hexokinase
HMQC	heteronuclear multiple-quantum correlation spectroscopy
HSQC	heteronuclear single-quantum correlation spectroscopy

## Nomenclature

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hVDAC	human voltage dependent anion channel
HVEM	high-voltage electron microscopy
IMM	inner mitochondrial membrane
IMP	integral membrane protein
IMS	inter membrane space
IPTG	isopropyl $\beta$ -D-1-thiogalactopyranoside
LDAO	lauryldimethylamine- <i>N</i> -oxide
MD	molecular dynamics
MFA	model free analysis
MTSL	<i>S</i> -(1-oxyl-2,2,5,5-tetramethyl-2,5-dihydro-1 <i>H</i> -pyrrol-3-yl)methyl methanesulfonothioate
mVDAC	murine voltage dependent anion channel
ncVDAC	<i>neurospora crassa</i> voltage dependent anion channel
NMR	nuclear magnetic resonance
NOESY	nuclear Overhauser effect spectroscopy
OMM	outer mitochondrial membrane
OPOE	octyl polyoxyethylene
ORF	open reading frame
PB	Poisson-Boltzmann
PCA	principal component analysis
PCR	polymerase chain reaction
PDB	Protein Data Bank
PEG	polyethyleneglycol
PMSF	phenylmethanesulfonyl fluoride
PRE	paramagnetic relaxation enhancement
RDC	residual dipolar coupling
RMSD	root mean square deviation
rVDAC	rat voltage dependent anion channel

S/N	signal-to-noise
scVDAC	<i>saccharomyces cerevisiae</i> voltage dependent anion channel
SD	standard deviation
SDS-PAGE	sodium dodecyl sulfate polyacrylamide gel electrophoresis
SEM	standard error of the mean
TEMED	tetramethylethylenediamine
TROSY	transverse relaxation-optimized spectroscopy
VDAC	voltage dependent anion channel
<i>wt</i>	wild type
zfVDAC	<i>zebra fish</i> voltage dependent anion channel



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# 1 Introduction

## 1.1 Membrane proteins

Membrane proteins are a large subgroup of proteins along with soluble globular proteins, fibrous proteins and disordered proteins. It has been estimated by various methods that about 20 to 30% of all open reading frames (ORFs) over all organisms encode membrane proteins.<sup>[1-3]</sup> Unlike soluble proteins, that are able to fold with high variability, membrane proteins are only known to occur in two topology categories:  $\alpha$ -helical bundles and  $\beta$ -barrels. Membrane proteins give the membranes their characteristical functional properties and allow them to carry out their respective activities. As such, the membrane protein content expressed as mass ratio in a membrane ranges from less than 25% (in the myelin membrane) to up to 75% (in internal membranes of mitochondria and chloroplasts).<sup>[4]</sup>

$\alpha$ -helical membrane proteins can be found in literally all types of biological membranes: besides occurring in the plasma membrane they are also found in the inner membranes (mitochondria, chloroplasts, endoplasmatic reticulum, peroxisomes and in bacteria). Their function comprises transport processes across membranes, metabolism, cell signaling and regulation processes and as such they are heavily addressed targets for drug discovery.<sup>[5]</sup>

In contrast to that,  $\beta$ -barrel membrane proteins are only found in the outer membranes of Gram-negative bacteria, scarcely in the cell walls of Gram-positive bacteria and in the outer membranes of mitochondria and chloroplasts. However, in those membranes  $\beta$ -barrel membrane proteins often-times make up the majority of the integral membran proteins. The importance of membrane proteins becomes evident with the fact, that they are targeted by more than 50% of small molecule drugs used in the treatment of human diseases: they have been and still are an intensively addressed drug target.<sup>[6]</sup>

### 1.1.1 Membrane proteins & NMR

The fact that membrane proteins are such a prominent drug target reasons, why there is also a big interest in the scientific community in determining the 3D structures of membranes proteins. In the Membrane Protein Structures Database of the Stephen White laboratory at University of California Irvine ([blanco.biomol.uci.edu/mpstruc/](http://blanco.biomol.uci.edu/mpstruc/); status of May 2017) 692 unique membrane protein structures are deposited. Of these, only 53 structures were solved with the means of NMR spectroscopy. Until now, X-ray crystallography is still the most commonly applied technique, although structure determination by cryo-EM is becoming more and more the method of choice in recent years.<sup>[7]</sup> However, the fact that of roughly 130'000 structures in the RCSB Protein Data Bank ([www.rcsb.org/pdb](http://www.rcsb.org/pdb); status of April 2017) only about 4130 structures are membrane proteins (which is only 3 %, i.e. far less than 20 to 30 % of all structures, cf. above) shows that all three techniques are experiencing difficulties in structure determination of membrane proteins.

NMR has the unique ability to access not only rigid structures but also dynamics and different states. Furthermore, there is still ongoing advancement in liquid and solid state NMR spectroscopy, e.g. APSY<sup>[8]</sup> or non-uniform sampling<sup>[9]</sup> for liquid state NMR or the development of 0.7 mm rotors that allow spinning frequencies of up to 111 kHz and thus the direct detection of protons for solid state NMR. Further increased field strengths of at the moment up to 1 GHz proton resonance frequency raise the overall sensitivity of the spectrometers. Magnets with 1.2 GHz proton resonance frequency are being in development.

That NMR is still a viable tool to determine 3D structures of membrane proteins can be seen by the fact that in 2017 already two structures of membrane proteins or domains of proteins embedded in membranes solved with NMR have been reported: the structure of the C-terminal transmembrane domain of the HDL receptor, SR-BI,<sup>[10]</sup> and the structure of the epidermal growth factor receptor transmembrane domain dimer.<sup>[11]</sup> These are faced with 20 structures solved by X-ray diffraction or cryo-EM in 2017 (according to [blanco.biomol.uci.edu/mpstruc/](http://blanco.biomol.uci.edu/mpstruc/); status of May 2017).

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## 1.2 Mitochondria

Mitochondria are organelles found in nearly all eukaryotic organisms (monocercomonoides sp. being the only one known exception<sup>[12]</sup>) and are the main source of the cells' energy currency by generating ATP through oxidative phosphorylation.<sup>[13]</sup> They are separated from the cytoplasm of the surrounding cell by two membranes, the inner and the outer mitochondrial membrane (IMM and OMM), which in turn are separated by the inner membrane space (IMS). Following the endosymbiotic hypothesis, the two membranes are leftovers from the mitochondria's origin as prokaryotic cells.<sup>[14]</sup> Besides their key function in the energy balance of the cell, mitochondria also play a role in the regulation of cell metabolism, cell-cycle control, development, antiviral responses and cell death.<sup>[15]</sup>

The IMM contains a series of respiratory enzyme complexes. Electrons passing through these complexes are used to build up a proton gradient across the IMM which is then enabling the ATP synthase to synthesize ATP.<sup>[13]</sup> Furthermore, the IMM contains the ADP/ATP carrier, which is catalyzing the ADP versus ATP exchange over the IMM and is thus critical for the supply of ATP from the mitochondria to the cytosol.<sup>[16,17]</sup>

The OMM is populated by four major integral membrane protein families. The translocase of the outer membrane (TOM complex), the sorting and assembly machinery (SAM complex) and the mitochondrial distribution and morphology (Mdm) complex all consist of a core subunit, which is an integral β-barrel membrane protein, and several smaller proteins that regulate the assembly and dynamics of the complexes.<sup>[18–22]</sup> These three OMM protein families are responsible for the translocation and insertion of nearly all synthesized proteins into the mitochondria.<sup>[23]</sup>

The last family is mediating the flux of all metabolites (including ADP and ATP) and ions across the OMM.<sup>[24]</sup> It is the family of voltage dependent anion channels (VDACs). They are named this way due to their anion selectivity and their voltage-dependent conductance.<sup>[25,26]</sup> In high density regions they cover up to 80 % of the membranes surface<sup>[27]</sup> and thus are the most abundant protein family in the OMM. They are known to form various oligomers<sup>[28]</sup>, which was determined by EM as early as in the 1960s.<sup>[29]</sup> The following chapter will describe this family in more detail.

## 1.3 VDAC – The voltage-dependent anion channel

VDAC, the voltage dependent anion channel, was discovered in two ways independently of each other: by electrophysiological measurements of the outer mitochondrial membrane and by electron microscopic (EM) observations of the same.<sup>[29]</sup>

Electrophysiological measurements of single-channels in membranes were made possible by the stable formation of planar membranes, which was described by Mueller *et al.* in 1962<sup>[30]</sup> and by Montal and Mueller in 1972.<sup>[31]</sup> However, in the first years after the invention of this technique, mainly channel-forming antibiotics that were reconstituted into these membranes were studied, as the patch-clamp technique was not invented before 1976.<sup>[32]</sup> Indeed, VDAC was the first intrinsic membrane channel that was reconstituted and studied at the single-channel level with planar membranes by Schein, Colombini *et al.* in the beginning of 1975, published in 1976.<sup>[25]</sup> Also the designation of the channel as “voltage dependent anion channel” is found in this paper first. Interestingly, Schein *et al.* originally wanted to study a voltage-gated calcium channel, but finally found out, that they reconstituted something completely different into the membrane. Different from what they expected, the reconstituted channels showed a conductance that depended on the applied voltage and favored anions like chloride over calcium. Since at that time the presence of channels in the mitochondrial membrane seemed to be unlikely, it was a big surprise that they were stemming from the mitochondrion. Further studies revealed that the VDAC channels were located in the OMM<sup>[25]</sup> and Marco Colombini concluded that they are the reason for its apparent “leakiness”, that had functionally been described by Werkheiser and Bartley more than 20 years before.<sup>[33,34]</sup>

Studies carried out in parallel by Zalman *et al.* were not published before 1980.<sup>[35]</sup> They described a protein with an apparent molecular weight of 30 kDa obtained from rat liver mitochondria, but found this protein to be a nonspecific diffusion channel for saccharides of up to 8 kDa. This publication is the origin of the termin “mitochondrial porin”. Only later it became clear that the mitochondrial porin and VDAC are the exact same protein.

Already more than ten years earlier in the middle of the 1960s, plant mitochondrial membranes had been studied with the means of negative-stain EM and revealed densely packed stain-filled subunits



with 2–3 nm wide “pits”, in a collaboration of Bonner, Parsons *et al.*<sup>[36]</sup> The identity of these pores was finally clarified by Carmen Mannella in the beginning of the 1980’s using high-voltage electron microscopy (HVEM) and in collaboration with Parsons. Working as a PhD candidate in the lab of Bonner, she had previously found with SDS-PAGE experiments that over 50 % of the protein mass of plant outer mitochondrial membranes belong to a protein with a mass of around 30 kDa, labeled “Band I”.<sup>[37]</sup> The obvious hypothesis was, that this band belonged to the subunits that Parsons *et al.* had observed with EM over ten years ago. The HVEM studies in the beginning of the 1980s finally revealed 2D crystals on the fungal outer mitochondrial membrane – which in turn were identified as being composed of VDAC, closing the circle to the observation in 1965.<sup>[38]</sup>

In the following years a wide variety of functions was described for the VDAC channels and it has been suggested that there is more than one isoform of the protein.<sup>[39]</sup> This hypothesis turned out to be true, as up to three isoforms were found, depending on the respective organism that is considered. In mammals like mice or the human, three isoforms of VDAC have been found<sup>[40,41]</sup> in tissue-specific expression-levels, while for example mitochondria from yeast contain only one or two isoforms.<sup>[42]</sup>

The isoform that was discovered in the 1970s is VDAC1, which is also by far the most abundant (factor 10 over VDAC2 and factor 100 over VDAC3 in HeLa cells) and consequently the best characterized isoform of the protein.<sup>[43]</sup> All three isoforms induce quite similar permeability when reconstituted into liposomes (molecular weight cutoff between 3.4 and 6.8 kDa based on the permeability of polyethyleneglycol). Electrophysiological studies show, that the prototypical gating properties (described in the following chapter) shown by VDAC1 are highly conserved among different species. However, VDAC2 exists in two forms, one of them with a lower conductance. VDAC3 does not insert into membranes as readily as VDAC1 and VDAC2 and generally does not show distinct gating properties.<sup>[44]</sup>

### 1.3.1 Voltage gating of VDAC

As described in the previous chapter, one of the most distinctive features of VDAC, and in particular of VDAC1, is its ability to show a conductance depending on the applied voltage, *i.e.* to switch from a high conducting state to a low conducting state depending on the applied potential. This property

of VDAC1 is highly conserved among the different mammalian VDACS: they all show very similar conductance, selectivity and voltage-gating.<sup>[45]</sup>

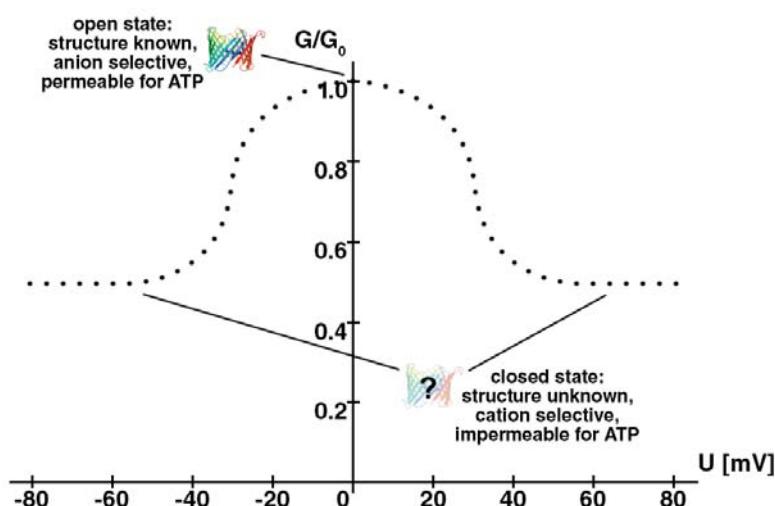
All values presented here have been determined with VDAC reconstituted into planar phospholipid bilayers. The high conducting state of the pore is accessible at low voltages (below  $\pm 20$  mV) with a typical conductance of around 4 to 4.5 nS in 1 M KCl (or around 0.5 nS in 0.1 M KCl).<sup>[45-52]</sup> The conductance of the channels is linear with the KCl activity up to 4 M KCl.<sup>[53]</sup> In this state, VDAC exhibits a slight selectivity for anions ( $P(\text{Cl}^-)/P(\text{K}^+) \approx 2$ )<sup>[45]</sup>, and is permeable to large anions such as ATP.<sup>[54]</sup> Under special conditions (a 5-fold KCl gradient across the membrane) also a cation-selective open state has once been described.<sup>[55]</sup>

At potentials above  $\pm 20$  mV, the probability of the channel to be in the high conducting (“open”) state decreases until it reaches 0 % at  $\pm 40$  to 50 mV, meaning that all channels observed are in lower conducting (“closed”) states.<sup>[45]</sup> The channel reaches different closed states at positive or negative potential, respectively.<sup>[53]</sup> The conductance of these closed states is usually around 50 % compared to the conductance of the open state with KCl as electrolyte in the aqueous phase – around 2 nS in 1 M KCl.<sup>[24]</sup> Deeper and more refined investigation on mVDAC1 revealed that there are two not completely identical closed substates with conductances of 2.6 nS and 1.9 nS – hence the conductance of the closed conformation of the channel is a composite of these states.<sup>[52]</sup> Upon closure, the selectivity of the channels switches to favor small cations over anions<sup>[24,56]</sup> and the pore is furthermore no longer permeable for ADP or ATP.<sup>[54,56]</sup> (See Figure 1.1.)

### 1.3.2 3D structure of VDAC

The 3D structure of VDAC was a long standing question that could finally be solved in 2008 – over 30 years after the protein’s discovery – by three groups independently. (See below.) Before, models with 12<sup>[57-59]</sup>, 13<sup>[60]</sup>, 16<sup>[61-63]</sup>, 18<sup>[64,65]</sup> or 19<sup>[66]</sup>  $\beta$ -strands and an amphipathic N-terminal  $\alpha$ -helix had been discussed. Most of the efforts to develop these models were done by Carmen Mannella, Elizabeth Blachly-Dyson, Michael Forte and Marco Colombini over the course of over 20 years.

The number of  $\beta$ -strands proposed by Blachly-Dyson, Forte and Colombini increased over time from 12 to 13, while Mannella always pointed out that there is a large amount of disagreement over



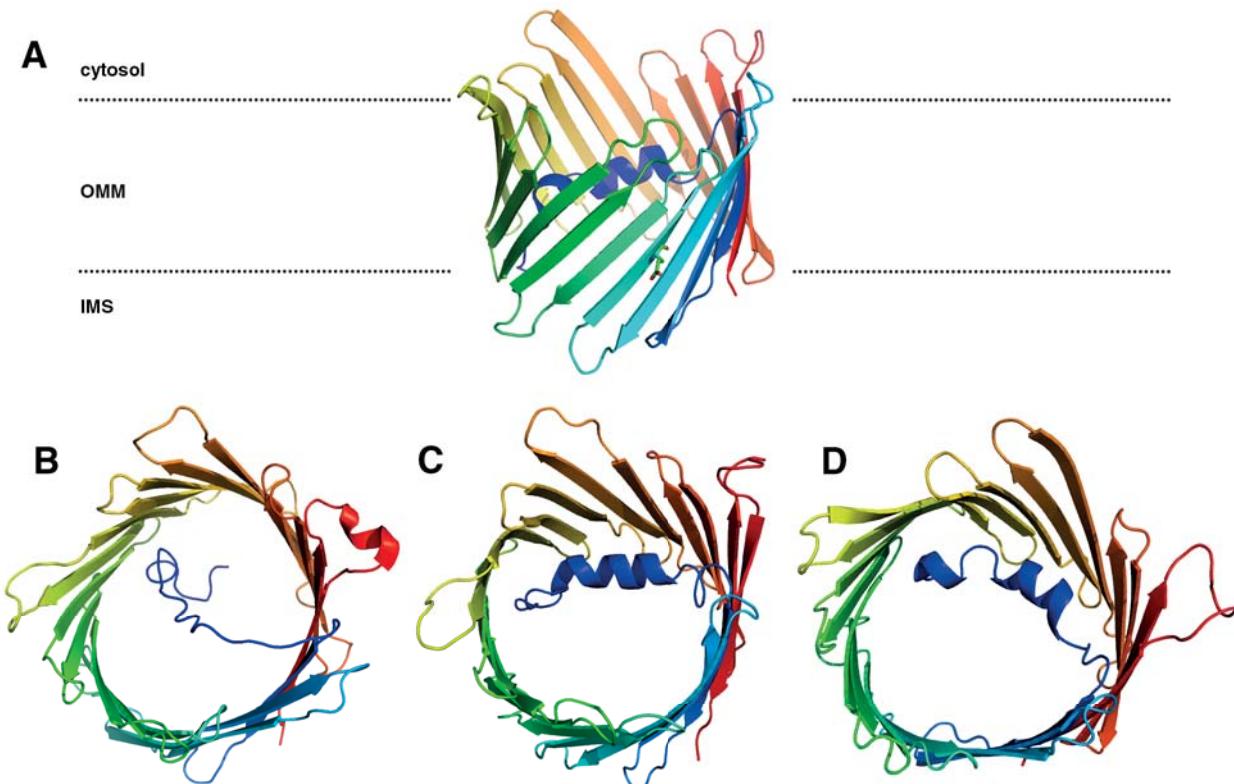
**Figure 1.1: Typical voltage gating profile of VDAC.** Conductance  $G$  of a VDAC channel as function of the applied potential with respect to the zero-voltage conductance  $G_0$ . The profile represents typical values yielded from studies in 1 M KCl as electrolyte. Structure shown is the 3D X-ray structure of mVDAC1 (PDB id: 3EMN).

the number of strands<sup>[67]</sup>, as there are models with 16  $\beta$ -strands published in the 1990s<sup>[61,62]</sup>, that were never really considered by the former. The 12- $\beta$ -strand model was derived in the late 1980s and early 1990s from a simple integration over the hydropathy values<sup>[68]</sup> of the amino acid side chains in groups of 10<sup>[57]</sup> and supported by site-directed mutagenesis that aimed to check for effects on channel selectivity or voltage dependence.<sup>[57,59]</sup> By biotinylation experiments the folding pattern was revised to include an N-terminal  $\alpha$ -helix and 13  $\beta$ -strands in 1998<sup>[60]</sup>, which is still the model Marco Colombini prefers nowadays for the native state of the protein.<sup>[69]</sup>

By neural network-based predictors, 16  $\beta$ -strands were proposed in 2002 again<sup>[63]</sup>, before 18  $\beta$ -strands were predicted by different authors based on computer analyses<sup>[65]</sup> and NMR data<sup>[64]</sup>. The three high resolution structures published in 2008, however, described the channel to be consisting of 19  $\beta$ -strands – in line with a prediction by the Delphi algorithm from as early as 1987.<sup>[66]</sup>

In contrast to a structure from this group, that was based on a combination of X-ray crystallography and solution state NMR data<sup>[70]</sup>, the other two structures were solely based on X-ray crystallography<sup>[46]</sup> or solution state NMR data<sup>[48]</sup>, respectively. All three 3D structures show that the N-terminus is not part of the barrel wall and located in the interior of the pore (unstructured in the NMR structure, as an  $\alpha$ -helix in the NMR/X-ray and X-ray structure). Furthermore, the structures showed that there is an unusual glutamate at position 73 in the sequence, which does not point

to the interior of the pore, but to the surrounding hydrophobic environment. The odd number of  $\beta$ -strands and the fact that the N-terminal helix is not part of the barrel wall was somewhat surprising, as this implies the need of a parallel arrangement of  $\beta$ -strands 1 and 19 and all previously solved  $\beta$ -barrel structures of prokaryotes had an even number of  $\beta$ -strands.<sup>[48]</sup> The three high resolution structures from 2008 are shown in Figure 1.2.



**Figure 1.2: 3D structures of VDAC1 from 2008.** All structures are shown in a cartoon representation and colored from N-terminus to C-terminus from blue to red. Visualization was done with PyMol.<sup>[71]</sup> A: X-ray structure of mVDAC1 (PDB id: 3EMN)<sup>[46]</sup> in a tilted view. The unusual glutamate pointing to the surrounding membrane is shown in stick representation. The dotted lines and the annotations indicate the cytosole, the Outer mitochondrial membrane (OMM) and the inter membrane space (IMS) as suggested by latest studies.<sup>[72,73]</sup> B: Lowest energy NMR structure of hVDAC1 from an ensemble of 20 structures (PDB id: 2K4T).<sup>[48]</sup> C: Combined NMR and X-ray structure of hVDAC1 (PDB id: 2JK4).<sup>[70]</sup> D: X-ray structure of mVDAC1 (PDB id: 3EMN).<sup>[46]</sup>

While the X-ray structure shows that the N-terminal helix should have a short kink from residues 10 to 12, it does not show this feature in the structure based on combined NMR and X-ray data. In the structure based on NMR data alone, the N-terminus is completely unstructured. Also, in the X-ray structure the N-terminal helix is connected tightly to the barrel, while in the structure based on combined data only the first part of the helix is connected to the barrel and in the NMR structure,



the N-terminus is floating freely in the middle of the barrel. Lastly, the N-terminus is rotated by approximately 90° in the structure based on combined data with respect to the X-ray structure. It was discussed in the PhD thesis of Saskia Villinger<sup>[74]</sup>, whether this is due to differences in the resolution of the structures or due to dynamics of the helix. Based on newer results, the former is most likely the case.<sup>[75]</sup>

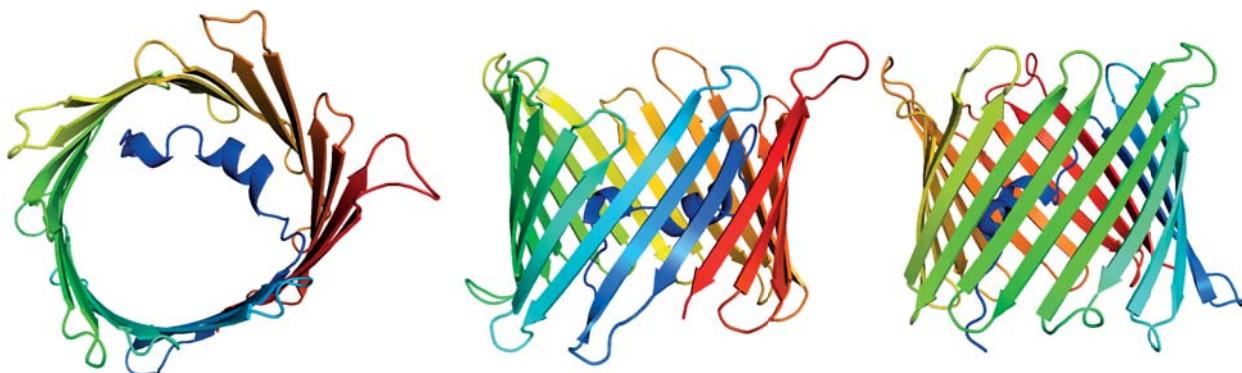
After the publication of these structures the question was raised especially by Marco Colombini, whether these structures represent a native and functional state of the channel or are an artifact of the refolding procedure that was necessary to study the respective samples.<sup>[76]</sup> However, the diameter of the published structures is in excellent agreement with values obtained from EM and AFM experiments<sup>[27,77,78]</sup> and the tilt angle of the β-strands fits well to other resolved structures of β-barrel proteins.<sup>[79]</sup> This would not be the case for the 13-stranded model favored by Marco Colombini.<sup>[69]</sup> Except of him it is thus now universally accepted that the 3D structures published in 2008 indeed represent a native conformation of the channel.

The structures were once again confirmed by further publications in 2014, when the crystal structure of mVDAC1 in presence of ATP was solved. It reveals a low affinity binding site of ATP in the center and bottleneck of the pore: between the N-terminal helix and the C-terminal end of the sequence.<sup>[80]</sup>

Also in 2014 the first 3D high resolution structure of the VDAC2 isoform was reported. The structure of zfVDAC2 was solved by X-ray crystallography and reveals a β-barrel with 19 β-strands and an N-terminal α-helix attached to the barrel wall at the usual position and with a kink consisting of residues 10 to 12 (PDB ID: 4BUM).<sup>[75]</sup> Overall, the X-ray structure of zfVDAC2 is strikingly similar to the X-ray structure of mVDAC1 with an RMSD of 0.98 Å. The only big difference between the structures is the loop from β-strand 1 to 2 with a displacement of 10 Å. The X-ray structure of zfVDAC2 is shown in Figure 1.3.

### 1.3.3 Gating models & evolution of the models

The development of the gating models of VDAC happened in close connection to the amount of structural information available about the channel, as with more structural information also the mode



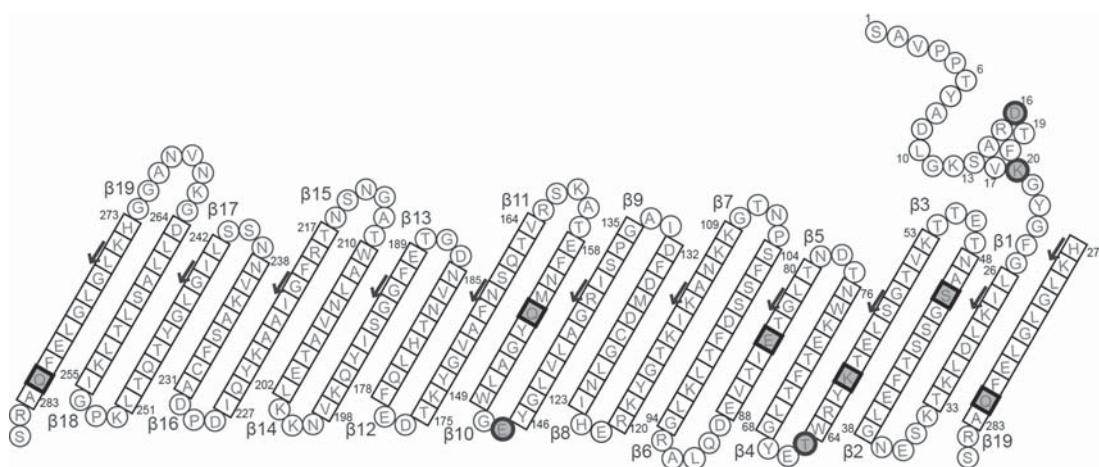
**Figure 1.3: 3D structure of zfVDAC2 from 2014 (PDB ID: 4BUM).** All views are shown in a cartoon representation and colored from N-terminus to C-terminus from blue to red. Visualization was done with PyMol.<sup>[71]</sup> zfVDAC2 from above shows the N-terminal helix with the characteristic kink (left) and the usual barrel fold known from the previous 3D high resolution structures (middle and right).

of gating could be proposed with more accuracy and detail. However, even with several high resolution structures available, the mode of gating is still not fully understood.

One of the earliest and most basic insights into the gating of VDAC was, that the inaccessible volume for polyethyleneglycol (PEG, a hydrophilic polymer) of the channel during closure is decreased by roughly 50 % of its initial volume in the open state. This decrease is in the order of  $2 \cdot 10^4 \text{ \AA}^3$ .<sup>[81]</sup>

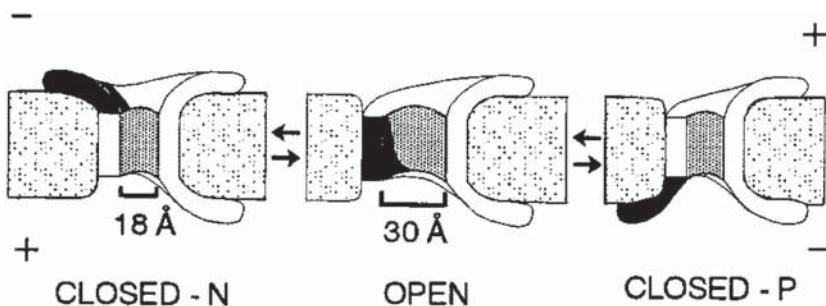
Furthermore, it became evident that there is a positively charged moiety, also called “voltage sensor” region in the protein, which moves during gating and that there are actually two distinct gating processes – one at negative and one at positive potential. These gating processes can be influenced e.g. by altering the voltage sensor with the means of site-directed mutagenesis (to make the voltage dependence asymmetric, stronger or to remove it)<sup>[59,82]</sup>, titration of the voltage sensor (to weaken the gating at higher pH values)<sup>[83]</sup>, phosphorylation (also resulting in asymmetric gating)<sup>[84]</sup> or the composition of the membrane in which VDAC is embedded<sup>[85]</sup>. The voltage sensor was determined using a VDAC model with 12  $\beta$ -strands, but based on the available 3D structures, the voltage sensor would comprise the N-terminal  $\alpha$ -helix and parts of  $\beta$ -strands 2–5, 9–10 and 19.<sup>[59]</sup> (See Figure 1.4.) The residues probed in  $\beta$ -strands 9 and 10 (Glutamate-145 and -152 in scVDAC1, analogous to Glutamate-147 and Glutamine-154 in hVDAC1) were shown to influence only one of the two gating processes.<sup>[82]</sup> If all of the determined voltage sensor regions would move upon closure of the channel, this would entail a large scale rearrangement of the protein domains.<sup>[86]</sup>

It was proposed that the voltage sensor of the protein moves in a way, that exposes it to the membrane surface during the gating process. This was probed with site-directed mutagenesis: single



**Figure 1.4: Voltage sensor in hVDAC1.** Residues that were identified to move during the gating process and thus been proposed to belong to the voltage of the protein are marked. The residues were determined using a model with 12  $\beta$ -strands. Glutamate-145 and -152 were shown to only influence one of the gating processes and are located in the loop from  $\beta$ -strand 9 to 10 and in  $\beta$ -strand 10.

residues were mutated to cysteine, biotinylated and tested for accessibility for streptavidin. It turned out, that it is the previously determined voltage sensor region that is accessible for streptavidin and upon binding the channels appeared to be locked in the closed state.<sup>[87]</sup> This was confirmed by N-terminal antibody studies on native OMM.<sup>[88]</sup> Based on these results the gating model shown in Figure 1.5 was developed.



**Figure 1.5: Gating processes as proposed in 1998.** The voltage sensor of the protein is moving to the surface, depending on the applied potential to reach two different closed states achieved at positive and negative potential. At the same time, during gating the channel diameter decreases. Reprinted from Song *et al.* with kind permission from Elsevier.

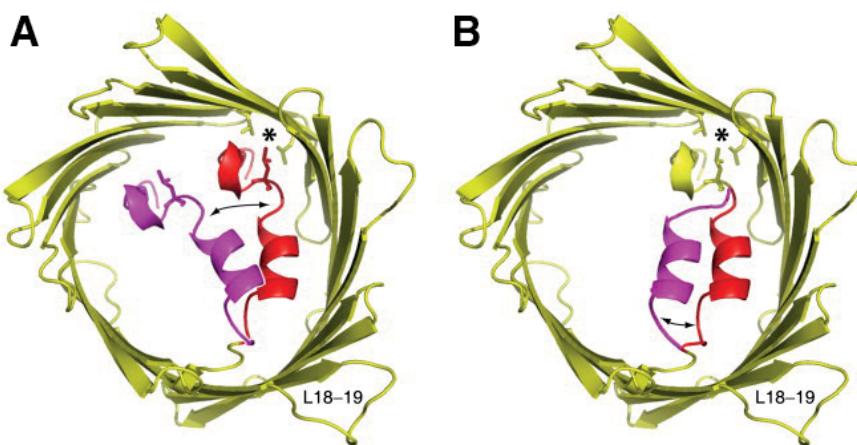
The importance of the N-terminal  $\alpha$ -helix for the channel conductance, stability and the gating process was tested with a vast variety of deletion mutants. Interestingly, results from these experiments seem to be somewhat contradicting. As early as 1996, BLM measurements showed that  $\Delta(2-12)$ -ncVDAC1 and  $\Delta(3-20)$ -ncVDAC1 form unstable, noisy channels with conductances mostly lower

than the usual open state, but were still described to show gating properties, however significantly weakened.<sup>[50]</sup> Partly contrary to that,  $\Delta(1\text{--}8)\text{-scVDAC1}$  also shows instable, flickering channels with lower conductance than the usual open state, but has completely lost its voltage dependence.<sup>[89]</sup> In 2008,  $\Delta(1\text{--}7)\text{-hVDAC1}$  and  $\Delta(1\text{--}19)\text{-hVDAC1}$  were construed. The former shows a broad distribution of conductance values with a maximum around 2.4 nS. The latter has a predominant conductance of around 1 nS, shows only some channels with around 2.4 nS and has furthermore lost its gating properties (no information was given about that on the former construct).<sup>[90]</sup> A very similar  $\Delta(1\text{--}20)\text{-hVDAC1}$  construct however shows conductances broadly distributed between 0.5 and 3 nS (i.e. no predominant conductance of 1 nS) without voltage dependence.<sup>[91]</sup> Finally,  $\Delta(1\text{--}26)\text{-mVDAC1}$  was described to have lost its voltage dependence, but “exhibited high conductance” (no exact values were given).<sup>[47]</sup>

After it was discovered that VDAC is a 19-stranded  $\beta$ -barrel, these results could be rationalized, as the N-terminal  $\alpha$ -helix obviously stabilizes the  $\beta$ -barrel and thus its complete or partial removal causes destabilization of the channel, resulting in (partial) collapse of the barrel structure and thus noisy and more closed state-like BLM measurements. Subsequent gating models were developed based on this combined knowledge and the available 3D structures.

The authors of the X-ray structure of mVDAC1 and two other authors had proposed detaching and movement of the N-terminal  $\alpha$ -helix into the middle of the pore as mechanism for the gating of the channel.<sup>[46,92]</sup> This model was refined by Hiller and Wagner specifying that only the second part of the helix moves and the hydrophobic contact between Leucine-10 and the  $\beta$ -barrel is retained.<sup>[93]</sup> (See Figure 1.6.)

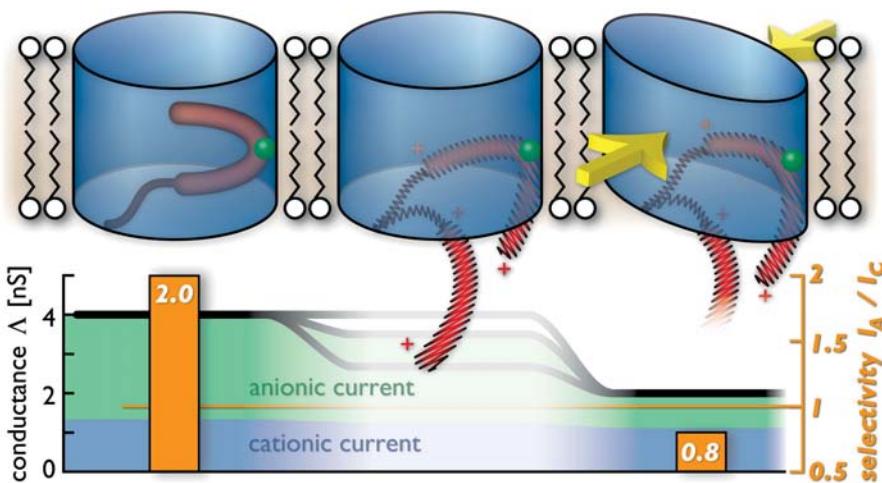
These models, however, are contradictory to the finding that the pore volume decreases upon closure.<sup>[81]</sup> Furthermore they could be ruled out also theoretically by Poisson-Boltzmann (PB) electrostatic calculations, as the helix motion proposed in these models does not show voltage dependence at all. Even a complete removal of the N-terminal  $\alpha$ -helix out of the barrel can explain only roughly half of the voltage dependence observed in experiments and the barrel would still be anion-selective according to these calculations. (It was, however, concluded that this motion of the helix could be part of the overall gating motion.) In the same publication it was shown with PB elec-



**Figure 1.6: Gating as proposed in 2009.** A: The N-terminal  $\alpha$ -helix as the voltage sensor moves to the middle of the pore during gating. B: Only the second part of the helix moves, retaining the hydrophobic contact between Leucine-10, Valine-143 and Leucine-150. Reprinted from Hiller and Wagner with kind permission from Elsevier.<sup>[93]</sup>

trostatic calculations that the three high resolution 3D structures from 2008 are selective for anions and thus should be identical to the open state of the protein.<sup>[94]</sup> This was confirmed by computational electrophysiology simulations<sup>[95]</sup>, from which it was likewise concluded that the published 3D structures are anion selective and have a conductance of  $4.2 \pm 0.2$  nS (SEM) in 1 M KCl.<sup>[91]</sup> According to these simulations, a cation selective state of the pore with a conductance of approximately 2 nS can be reached by removing the N-terminal  $\alpha$ -helix out of the barrel and collapsing the barrel to an ellipticity of around 0.5.<sup>[91]</sup> The calculated volume decrease due to this collapse would be around  $1 \cdot 10^4 \text{ \AA}^3$ , which is in the same order of magnitude compared to previous experimental values, but not completely coinciding to these. (See above, a loss of  $2 \cdot 10^4 \text{ \AA}^3$  was determined with PEG.<sup>[81]</sup> To reach this volume decrease with an elliptic deformation, an even higher ellipticity of approximately 0.75 would be necessary.)

Further experimental evidence against the models presented in Figure 1.6 is provided by the finding, that an L10N hVDAC1 mutant (i.e. a mutant in which the hydrophobic contact between Leucine-10, Valine-143 and Leucine-150 is destroyed) is already closed at very low voltages. This indicates that increasing the mobility of the N-terminal  $\alpha$ -helix by destroying the hydrophobic contact forces the channel into a closed state.<sup>[91]</sup>



**Figure 1.7: Gating as proposed in 2012.** The gating process consists of the N-terminal  $\alpha$ -helix detaching from the hydrophobic contact point and moving out of the barrel together with a partial collapse of the barrel, which is calculated to end up in a low-conducting, cation selective state. Reprinted from Zachariae *et al.* with kind permission from Elsevier.<sup>[91]</sup>

One of the most recent gating models, which seeks to comprise all available information was published in 2012 by this group and is shown in Figure 1.7.<sup>[91]</sup> The N-terminal  $\alpha$ -helix as positively charged voltage sensor moves out of the barrel, in line with respective experimental results.<sup>[87,88]</sup> Upon this, the barrel partly collapses, explaining its lower diameter and volume in the closed state. The calculated loss of volume would be in the order of  $10^4 \text{ \AA}^3$ . As mentioned above, this would, however, not completely explain the volume loss determined experimentally.<sup>[81]</sup> With these combined movements the barrel ends up in a state that is calculated to be slightly cation selective and to have a conductance of around 2 nS.

### 1.3.3.1 Physiological relevance of VDAC gating

The physiological relevance of the voltage gating of VDAC is still not fully understood. Because of the large permeability of the VDAC channel in both its open and closed state, it is often assumed that there cannot be a membrane potential large enough to induce closure of the channel under physiological conditions by voltage alone, which would render this special property of the channel biologically insignificant.<sup>[96]</sup> However, an electrochemical equilibrium of small ions does not necessarily mean that there is no potential across the OMM, as there are lots of charged proteins in the cytosol and the IMS, for which VDAC is not permeable. Additionally, measurements of the pH difference between



the IMS and the cytosol showed that there could indeed be a transmembrane potential: two groups independently measured pH differences, that would – based on the Nernst equation – translate to potentials of  $\approx 40 \text{ mV}^{[97]}$  or 20 to 30 mV<sup>[98]</sup>, respectively. These values are exactly in the range in which VDAC switches from the open to the closed state. Thus a biological relevance of the voltage gating process of VDAC is well conceivable.

Even if the voltage gating process of VDAC is of no physiological relevance, it is still a fact that VDAC in its open state is permeable for ADP and ATP, while VDAC in its closed state is not.<sup>[99]</sup> Furthermore, it is known that VDAC interacts with many of the proteins and compounds available in the OMM and is also involved in many processes therein.<sup>[100]</sup> For example, tubulin is known to block VDAC channels by inserting its negatively charged tail into the channel in a way that is also seen during voltage gating (lower conductance, inversion of selectivity, impermeability for ATP).<sup>[101,102]</sup> Tubulin polymerizes under cellular activity, lowering the number of tubulin dimers and thus lowering the amount of VDAC blockage, which verifies the functional role of the VDAC-tubulin interaction in regulating mitochondrial respiration.<sup>[96,99]</sup> Additionally, VDAC interacts with HK-I, which induces closure of the channel<sup>[103]</sup> and many pro- and anti-apoptotic proteins, making VDAC a key protein in apoptosis.<sup>[100]</sup>

## 1.4 Aims and Outline

Despite the efforts of over 40 years of research, a multitude of phenomenological results regarding the gating process of VDAC, and many gating models, the exact nature of the closed states (or even a single one of them) of the protein is still unknown. The aim of this thesis is to improve the existing models of the closed state of VDAC and to reach a level of information which might open the path to determining a high resolution 3D structure of the channel in its closed state.

The first part of the thesis deals with a further improvement of the assignment of the channel by the means of additionally recorded experiments such as TROSY-HNCACB and by further improving the sample preparation. This is connected with a new expression vector used for the preparation of the hVDAC1 samples. Also, the assignment of already existing and previously recorded experiments was improved dramatically, leading to an overall improvement of available information on the channel based on chemical shifts.

The second part of the thesis is based on the improved assignment of the first part. In collaboration with Mariusz Jaremko and Łukasz Jaremko, the improved assignment of a HMQC-NOESY spectrum was utilized to calculate a high resolution 3D structure of E73V hVDAC1.

The third and last part focusses on the development of a model for the closed state of VDAC. This is based on the construction of a Quintuple hVDAC1 mutant with unique gating properties. The spectral properties of this mutant are determined by NMR spectroscopy and BLM measurements are performed. A new position of the N-terminal helix was proven and probed by the means of liquid and solid state NMR spectroscopy.

## 2 Materials & Methods

### 2.1 Materials

#### 2.1.1 Chemical reagents

Lauryldimethylamine-*N*-oxide (LDAO) was purchased from Sigma-Aldrich and *S*-(1-oxyl-2,2,5,5-tetramethyl-2,5-dihydro-1H-pyrrol-3-yl)methyl methanesulfonothioate (MTSL) from Toronto Research Chemicals.  $^{13}\text{C}$ -D-glucose (U- $^{13}\text{C}$ 6, 99 %) and  $^{15}\text{NH}_4\text{Cl}$  ( $^{15}\text{N}$ , 99 %) were purchased from Cambridge Isotope Laboratories, Sigma-Aldrich or Cortecnet. Other chemicals were purchased from the usual suppliers and used without further purification. BenchMark<sup>TM</sup> Protein Ladder was purchased from Invitrogen (now Thermo Fisher Scientific).

#### 2.1.2 Bacterial strains

- *Escherichia coli* strain M15 [pRep4] (Qiagen, Hilden; phenotype: *E. coli* K12 NaI<sup>S</sup> Str<sup>S</sup> Rif<sup>S</sup> Thi<sup>-</sup> Lac<sup>-</sup> Ara<sup>+</sup> Gal<sup>+</sup> Mtl<sup>-</sup> F<sup>-</sup> RecA<sup>+</sup> Uvr<sup>+</sup> Lon<sup>+</sup>; carries pREP4 (*lacI* encoding repressor plasmid)) for pQE60 constructs.
- *Escherichia coli* strain BL21 [DE3] (Novagen, Merck Biosciences, Darmstadt, phenotype: *E. coli* E. coli str. B F<sup>-</sup> ompT gal dcm lon hsdSB(r<sub>B</sub><sup>-</sup>m<sub>B</sub><sup>-</sup>) λ(DE3 [lacI lacUV5-T7p07 ind1 sam7 nin5]) [malB<sup>+</sup>]K-12(λ<sup>S</sup>)) for pET28a constructs.

## 2 Materials & Methods

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### 2.1.3 Growth media

**Table 2.1:** M9 minimal medium (standard/deuterated).

medium	substance	amount
M9 minimal medium	M9 salt (5x)*	200 mL
	Trace elements <sup>+</sup>	10 mL
	Thiamine HCl (5 mg/mL)	6 mL
	CaCl <sub>2</sub> (2 M)	0.05 mL
	MgSO <sub>4</sub> (1 M)	2 mL
	D-glucose or <sup>13</sup> C-D-glucose (20 % w/v)	20 mL
	<sup>15</sup> NH <sub>4</sub> Cl (0.25 g/mL)	4 mL
	D <sub>2</sub> O	758 mL
<hr/>		
*) M9 salt (5x)	Na <sub>2</sub> HPO <sub>4</sub>	33.9 g
	KH <sub>2</sub> PO <sub>4</sub>	15 g
	NaCl	2.5 g
	D <sub>2</sub> O	ad 1.0 L
<hr/>		
+) Trace elements	FeSO <sub>4</sub> · 7H <sub>2</sub> O	0.6 g
	MnCl <sub>2</sub> · 4H <sub>2</sub> O	0.115 g
	CoCl <sub>2</sub> · 6H <sub>2</sub> O	0.08 g
	ZnSO <sub>4</sub> · 7H <sub>2</sub> O	0.07 g
	CuCl <sub>2</sub> · 2H <sub>2</sub> O	0.03 g
	H <sub>3</sub> BO <sub>3</sub>	0.002 g
	(NH <sub>4</sub> ) <sub>6</sub> Mo <sub>7</sub> O <sub>24</sub> · 4H <sub>2</sub> O	0.025 g
	EDTA	0.5 g
	D <sub>2</sub> O	ad 100 mL



## 2.1.4 SDS page gels

**Table 2.2:** Resolving gel and stacking gel for SDS page.

gel	substance	amount
resolving gel 12 %	30 % Acrylamide	2.0 mL
	TrisHCl pH 8.8 (1 M)	1.88 mL
	H <sub>2</sub> O (bidest.)	1.03 mL
	10 % SDS	50 µL
	10 % APS	50 µL
	TEMED	2.5 µL
stacking gel 3 %	30 % Acrylamide	250 µL
	TrisHCl pH 6.8 (1 M)	313 µL
	H <sub>2</sub> O (bidest.)	1.88 mL
	10 % SDS	25 µL
	10 % APS	25 µL
	TEMED	2 µL

## 2.2 Methods

### 2.2.1 Sample preparation of hVDAC1

Cloning, expression and purification of *wt* and mutant hVDAC1 with a C-terminal His<sub>6</sub>-tag were performed as previously described in the doctoral theses of Thomas Meins<sup>[104]</sup>, Monika Bayrhuber<sup>[64]</sup> and Saskia Villinger<sup>[74]</sup> by Stefan Becker and Karin Giller in the Department of NMR-based Structural Biology, Max Planck Institute for Biophysical Chemistry, Göttingen. Due to identical procedures, parts of the following chapters are identical to their theses and are marked accordingly. The preparation of hVDAC1 was done mostly as described previously.<sup>[49]</sup>

#### 2.2.1.1 Cloning of hVDAC1<sup>1</sup>

The plasmid containing *wt* hVDAC1, cloned into pDS56/RBSII-6xHis vector via the *Bam*H1 and *Bgl*II restriction sites as described<sup>[49]</sup>, was previously kindly provided by Thomas Meins (Max Planck Institute for Biochemistry, Martinsried). The PCR construct of *wt* hVDAC1 was amplified from this vector and ligated with the *Nco*I and *Bgl*II restriction sites in the pQE60 vector (Qiagen, Hilden). Cloning the pET28a construct of *wt* hVDAC1 was done by amplification from the pQE60 vector. For the following ligation the *Nco*I and *Xho*I restriction sites were used.

#### 2.2.1.2 Site directed mutagenesis<sup>1</sup>

Point mutations into the cloned hVDAC1 coding sequence were introduced with the QuikChange Site-Directed Mutagenesis Kit (Agilent Technologies, Santa Clara), which was used essentially as recommended by Agilent. hVDAC1 mutants, vectors and primers used in this thesis are listed in Table 2.3. Successful mutagenesis was verified by DNA sequence analysis.

**Table 2.3:** Oligonucleotide primers for the mutagenesis of hVDAC1-His<sub>6</sub>.

Mutation	Host	Vector	Primers (forward and reversed)
C127S	<i>wt</i>	b)	5' -CACATTAACCTGGGCTCCGACATGGATTTCGAC- 3' 3' -GTCGAATCCATGCGAGCCCAGTTAATGTG- 5'
C127S/C232S	C127S	b)	5' -GACCCTGACGCCCTCTCTCGGCTAAAGTG- 3' 3' -CACTTAGCCGAGAAGGAGGCCTCAGGGTC- 5'

<sup>1</sup>Due to identical procedures, parts of this section are identical to the thesis of Saskia Villinger<sup>[74]</sup>

Mutation	Host	Vector	Primers (forward and reversed)
E73V	wt	b)	5' -CGGCCTGACGTTTACAGTGAATGGAATACCGAC-3' 3' -GTCGGTATTCATTTCACTGTAAACGTCAGGCCG-5'
C127A/C232S	C127S/C232S	a)	5' -CACATTAACCTGGGCCGACATGGATTTCGAC-3' 3' -GTCGAATCCATGTCGGCGCCAGGTTAATGTG-5'
C127A/C232A	C127A/C232S	a)	5' -GACCTGACGCCCTCTCGCTAAAGTG-3' 3' -CACTTAGCCGAGAACGGCGCTCAGGGTC-5'
A14C/C127A/C232A	C127A/C232A	a)	5' -GCGCATCTGGCAAATCTGCAGGGATGTCTCACCAAG-3' 3' -CTTGGTGAAGACATCCCTGCAAGATTGCAAGATCGGC-5'
A14C/C127A/S193C/ C232A	A14C/C127A/C232A	a)	5' -GGACAGAGTTGGCGCTGCATTACAGAAAGTGAAC-3' 3' -GTTCACTTCCTGTTAATGCAGCCGCAAACCTGTCCC-5'
E73V/C127A/C232S	C127A/C232S	c)	5' -CGGCCTGACGTTTACAGTGAATGGAATACCGAC-3' 3' -GTCGGTATTCATTTCACTGTAAACGTCAGGCCG-5'
G21V	wt	c)	5' -GCCAGGGATGCTTCCAAGGTCTATGGATTGGCTTAATAAG-3' 3' -CTTATTAAGCCAATCCATAGACCTGGTGAAGACATCCCTGGC-5'
G21V/G23V	G21V	c)	5' -GATGCTTCCAAGGGCTATGTATTGGCTTAATAAGCTTG-3' 3' -CAAGCTTATAAGCCAATACATAGCCCTGGTGAAGACATC-5'
G21V/E73V/C127A/ C232S	E73V/C127A/C232S	c)	5' -GCCAGGGATGCTTCCAAGGTCTATGGATTGGCTTAATAAG-3' 3' -CTTATTAAGCCAATCCATAGACCTGGTGAAGACATCCCTGGC-5'
G21V/G23V/E73V/ C127A/C232S	G21V/E73V/C127A/ C232S	c)	5' -GATGCTTCCAAGGGCTATGTATTGGCTTAATAAGCTTG-3' 3' -CAAGCTTATAAGCCAATACATAGCCCTGGTGAAGACATC-5'
A2C/G21V/G23V/ E73V/C127A/C232S	G21V/G23V/E73V/ C127A/C232S	c)	5' -GAAGGAGATACCATGTGTGCCACCCACGTATGCCG-3' 3' -CGGCATACGTGGTGGCACACATGGTATATCCTTC-5'
A2C/E73V/C127A/ C232S	E73V/C127A/C232S	c)	5' -GAAGGAGATACCATGTGTGCCACCCACGTATGCCG-3' 3' -CGGCATACGTGGTGGCACACATGGTATATCCTTC-5'

Vectors: a) pDS56/RBSII, b) pQE60, c) pET28a

### 2.2.1.3 Expression of $^2\text{H}$ , $^{13}\text{C}$ and $^{15}\text{N}$ labelled hVDAC1<sup>1</sup>

Following transformation into *Escherichia coli* M15 [pRep4] (for pQE60 constructs) or *Escherichia coli* BL21 [DE3] (for pET28a constructs), respectively, cells were grown at 37 °C over night in M9 minimal medium containing 100 µg/mL ampicillin and 25 µg/mL kanamycin. According to the desired labelling ( $^2\text{H}$ (75 %)/ $^{13}\text{C}$ / $^{15}\text{N}$ ,  $^2\text{H}$ (99 %)/ $^{13}\text{C}$ / $^{15}\text{N}$  or  $^2\text{H}$ (75 %)/ $^{15}\text{N}$ ), glucose and  $\text{NH}_4\text{Cl}$  in the M9 minimal medium (Table 2.1) were replaced with protonated or deuterated  $^{13}\text{C}$ -D-glucose and/or  $^{15}\text{NH}_4\text{Cl}$ . In order to adapt the culture to  $\text{D}_2\text{O}$  based media the cells were consecutively inoculated into M9 medium with a  $\text{D}_2\text{O}$  content of 33, 65 and 100 % and incubated at 37 °C for 24 h at each step. The fully adapted cell culture was used for inoculation of 1.5 L expression cultures with an initial  $\text{OD}_{600}$  of 0.1 AU. The culture was allowed to grow at 37 °C, until an  $\text{OD}_{600}$  of 0.8 AU was reached. Subsequently, hVDAC1-His<sub>6</sub> expression into inclusion bodies was induced with 1 mM IPTG. Cells were harvested 8 to 14 h (depending on the mutant) after induction by centrifugation at 5,000 g for 30 min, shock frozen in liquid N<sub>2</sub> and stored at -70 °C.

<sup>1</sup>Due to identical procedures, parts of this section are identical to the thesis of Saskia Villinger<sup>[74]</sup>

#### 2.2.1.4 Refolding and purification of hVDAC1<sup>1</sup>

Frozen cells were thawed, resuspended in buffer 1 (100 mM Tris/HCl pH 7.5; 1 mM EDTA; 5 mM DTT, 10 mM NaCl, 0.2 mM PMSF), and incubated with 0.1 mg/mL lysozyme for 30 min. Following lysozyme treatment, 1 mM MgCl<sub>2</sub>, 0.1 mM MnCl<sub>2</sub>, and 0.05 mg/mL DNase I were added and the cells were lysed by the French press passes. Inclusion bodies were harvested by centrifugation at 5,000 g for 30 min at 4 °C at 20,000 psi. The pellet was resuspended in buffer 1 containing 3 % (w/v) OPOE, stirred for 3 h at room temperature and pelleted by centrifugation at 5,000 g for 30 min at 4 °C. Finally, the pellet was washed with buffer 1 to remove detergent contaminations. Inclusion bodies were solubilized in buffer 2 (100 mM Tris/HCl pH 7.5; 1 mM EDTAm 5 mM DTT, 6 M guanidinium chloride) and insoluble material was removed by centrifugation at 100,000 g for 45 min. The denatured hVDAC1-His<sub>6</sub> protein containing supernatant was adjusted with buffer 2 to 15 mg/mL and stored at -70 °C until use.

Refolding was performed at 4 °C by dropwise dilution of solubilized protein in buffer 3 (100 mM Tris/HCl pH 8.0; 1 mM EDTA; 5 mM DTT, 1 % (w/v) LDAO) until a final concentration of 0.6 M guanidinium chloride was reached. The resulting protein solution was stirred over night at 4 °C, centrifuged at 100,000 g for 45 min and finally 5-fold diluted with buffer 4 (100 mM phosphate buffer ph 7.5).

Refolded hVDAC1-His<sub>6</sub> was purified by immobilized metal ion affinity chromatography (IMAC). For this purpose, hVDAC1 was bound to a 5 mL Ni<sub>2+</sub>-Sepharose HP Column (GE Healthcare, Chalfont St Giles), washed with 100 mL buffer 5 (20 mM phosphate buffer ph 7.5; 20 mM imidazole; 0.2 % LDAO), and afterwards eluted by 50 mL buffer 6 (20 mM phosphate buffer pH 7.5, 300 mM imidazole; 0.2 % LDAO). hVDAC1 containing fractions (as judged by SDS-PAGE), were pooled. Improved sample purification was reached by an additional gel filtration step that was applied to all new mutants described in this thesis. For this purpose, the pooled hVDAC1 fractions obtained from Ni<sub>2+</sub> affinity chromatography were concentrated and applied to a Superdex 200 16/60 column (GE Healthcare, Chalfont St Giles) equilibrated with buffer 7 (25 mM BisTris pH 6.8, 150 mM NaCl, 0.2 % LDAO). Eluted hVDAC1 fractions were pooled and the protein concentration was

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<sup>1</sup>Due to identical procedures, parts of this section are identical to the thesis of Saskia Villinger<sup>[74]</sup>



measured by UV absorption at 280 nm wavelength using the theoretical extinction coefficient of  $38,390 \text{ M}^{-1} \text{ cm}^{-1}$ . LDAO was added to the pooled fractions in order to adjust the final concentration of a 0.6 mM hVDAC1 sample to a theoretical LDAO content of 6 % (w/v), assuming that no detergent is lost during further purification steps. After LDAO addition, the sample was dialysed against NMR buffer (25 mM BisTris/HCl pH 6.8, 0.2 % LDAO) for 4 h and concentrated with a centrifugal filter device (Amicon Ultra-30 k, Millipore) to a protein concentration of 0.4-1 mM (for pQE60 samples) or 0.15-0.4 mM (for pET28a samples), respectively. By comparison with a sample of known LDAO concentration  $^1\text{H}$  NMR spectroscopy revealed typical LDAO concentrations of 2-4 % (for pQE60 samples) or 1-2 % (for pET28a samples), respectively, in the final samples.

### 2.2.2 Spin labelling<sup>1</sup>

A2C/E73V/C127A/C232S hVDAC1 and A2C/G21V/G23V/E73V/C127A/C232S hVDAC1 were modified with the thiol-specific nitroxide spin label reagent MTSL. 100  $\mu\text{g}/\mu\text{L}$  MTSL in cold acetone were added to the protein solution in a three- to five-fold molar excess prior to the refolding step and the solution was incubated for 1.5 h at room temperature. Afterwards, refolding and purification were performed as described in Chapter 2.2.1.4.

### 2.2.3 NMR spectroscopy

NMR spectra were recorded on  $^2\text{H}(75\%)/^{15}\text{N}$  or  $^2\text{H}(75\%)/^{13}\text{C}/^{15}\text{N}$  labeled samples containing 0.15–0.6 mM hVDAC1 (or the respective hVDAC1 mutant), 25 mM BisTris pH 6.8, 1–2 % (45–90 mM) LDAO and usually 10 %  $\text{D}_2\text{O}$ . Unless stated otherwise, all spectra were measured at 37 °C on Bruker 700, 800, 900 or 950 MHz Avance I, Avance III or Avance III HD spectrometers equipped with cryogenic probes. Recorded spectra were processed using NMRPipe<sup>[105]</sup> and analyzed using TopSpin<sup>[106]</sup>, SPARKY<sup>[107]</sup> or CcpNmr Analysis<sup>[108]</sup>.

<sup>1</sup>Due to identical procedures, parts of this section are identical to the thesis of Saskia Villinger<sup>[74]</sup>

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### 2.2.3.1 Assignment improvement and assignment of hVDAC1 mutants

For the improvement of the backbone resonance assignment reported earlier<sup>[64,74]</sup>, a TROSY-HNCACB experiment<sup>[109,110]</sup> was recorded on E73V/C127A/C232S hVDAC1 (for the pulse program see Appendix A.1). A mixed-time parallel evolution HMQC-NOESY spectrum<sup>[111]</sup> on E73V hVDAC1 was used to further improve the inter-strand and sequential H<sub>N</sub>-H<sub>N</sub> NOEs. To assign the various hVDAC1 mutants described in this thesis, <sup>1</sup>H-<sup>15</sup>N-TROSY spectra<sup>[112]</sup>, TROSY-HNCACB spectra and HMQC-NOESY spectra were used. Furthermore, as previously, existing N<sub>H</sub> and C<sub>α</sub> shifts from solid state NMR experiments were taken into account.<sup>[113]</sup> All 3D spectra used in the thesis are listed in table 2.4.

**Table 2.4:** Experiments and parameters for hVDAC1 assignment improvement and assignment of hVDAC1 mutants.

Sample	Spectrum	Parameters	Comments
<sup>2</sup> H(75 %)/ <sup>13</sup> C/ <sup>15</sup> N-E73V hVDAC1-pDS56/RBSII	TROSY-HNCA*	35×26×512 pts; ns = 72; 900 MHz	linear prediction in <sup>13</sup> C and <sup>15</sup> N dimensions
	<sup>15</sup> N-NOESY-HMQC*	85×25×512 pts; ns = 26; 800 MHz mt = 240 ms	
<sup>2</sup> H(75 %)/ <sup>13</sup> C/ <sup>15</sup> N-A14C/ C127A/S193C/C232A hVDAC1-pQE60	TROSY-HNCA	35×26×512 pts; ns = 92; 900 MHz	linear prediction in <sup>13</sup> C and <sup>15</sup> N dimensions
<sup>2</sup> H(75 %)/ <sup>13</sup> C/ <sup>15</sup> N-E73V/ C127A/C232S hVDAC1-pET28a	TROSY-HNCACB	35×26×512 pts; ns = 52; 800 MHz	
<sup>2</sup> H(75 %)/ <sup>13</sup> C/ <sup>15</sup> N-G21V/ G23V/E73V/C127A/C232S hVDAC1-pET28a	TROSY-HNCACB	35×22×512 pts; ns = 112; 950 MHz	
	<sup>15</sup> N-NOESY-HMQC	85×25×512 pts; ns = 34; 800 MHz mt = 240 ms	
	TROSY-HNCO	29×26×512 pts; ns = 76 700 MHz	



Sample	Spectrum	Parameters	Comments
<sup>2</sup> H(75 %)/ <sup>13</sup> C/ <sup>15</sup> N-A2C/ E73V/C127A/C232S hVDAC1-pET28a + MTS	TROSY-HNCACB	50×34×512 pts; ns = 72; 800 MHz	

\* recorded by Saskia Villinger  
 Spectroscopic parameters are given as number of complex points in F1, F2 and F3, number of scans, field strength for <sup>1</sup>H and mixing time

### 2.2.3.2 Estimation of $T_2$ and the global correlation time $\tau_c$

To estimate the overall transverse relaxation time of the protein, 1D <sup>1</sup>H (1–1) echo experiments were recorded.<sup>[114]</sup> Relaxation delays of 110, 1000, 2000 and 3000 μs were used for G21V/G23V hVDAC1. Transverse relaxation times  $T_2$  were estimated from two values following the formula:

$$T_2 = \frac{2(\Delta_1 - \Delta_2)}{\ln(I_1/I_2)} \quad (2.1)$$

With  $\Delta_1$  and  $\Delta_2$  being the relaxation delays of the two points and  $I_1$  and  $I_2$  being the maximum signal intensities at a certain chemical shift at these two relaxation delays.

The overall rotational correlation time  $\tau_c$  was then estimated as:

$$\tau_c \approx \frac{1}{5 \cdot \frac{T_2}{[ms]}} \cdot [\mu s] \quad (2.2)$$

### 2.2.3.3 Determination of chemical shift differences

To assess the effects of mutations on hVDAC1, <sup>1</sup>H-<sup>15</sup>N-TROSY spectra were recorded in order to follow <sup>1</sup>H and <sup>15</sup>N chemical shift changes. If resonances of residues in spatial proximity to mutation sites could only be ambiguously assigned, they were excluded from the analysis. Normalized weighted average chemical shift differences (CSD),  $\Delta_{HN}$ , were calculated as:

$$\Delta_{HN} = \sqrt{\frac{(\Delta\delta_N/5)^2 + (\Delta\delta_H)^2}{2}} \quad (2.3)$$

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where  $\Delta\delta_N$  and  $\Delta\delta_H$  are the chemical shift differences for the  $^{15}\text{N}$  and  $^1\text{H}$  dimensions, respectively. Errors in CSD were estimated as described in the PhD thesis of Saskia Villinger<sup>[74]</sup> and are given as average values: 0.014 ppm for  $^{15}\text{N}$  CSD and 0.0028 ppm for the average CSD.

If CSD values were mapped to a 3D structure of VDAC1, they were mapped to the crystal structure of mVDAC1 (PDB ID: 3EMN)<sup>[46]</sup> or the new E73V hVDAC1 solution structure (PDB ID: 5JDP)<sup>[115]</sup> and visualized with PyMOL<sup>[71]</sup>.

### 2.2.3.4 Determination of intensity changes

To follow the effects of MTSL labeling experiments, intensities were determined as absolute peak heights in the recorded  $^1\text{H}$ - $^{15}\text{N}$ -TROSY spectra. If resonances of residues in spatial proximity to mutation sites or the MTSL label, respectively, could only be ambiguously assigned, they were excluded from the analysis. Intensity ratios  $I_{\text{labeled}}/I_{\text{ref}}$  were scaled according to resonances that were unaffected from the labeling and possible CSD effects.

Errors in resonance intensity ratios were calculated by error propagation using the root mean square noise of the respective spectra.

If intensity ratios were mapped to a 3D structure of VDAC1, they were mapped to the crystal structure of mVDAC1 (PDB ID: 3EMN)<sup>[46]</sup> or the new E73V hVDAC1 solution structure (PDB ID: 5JDP)<sup>[115]</sup> and visualized with PyMOL<sup>[71]</sup>.

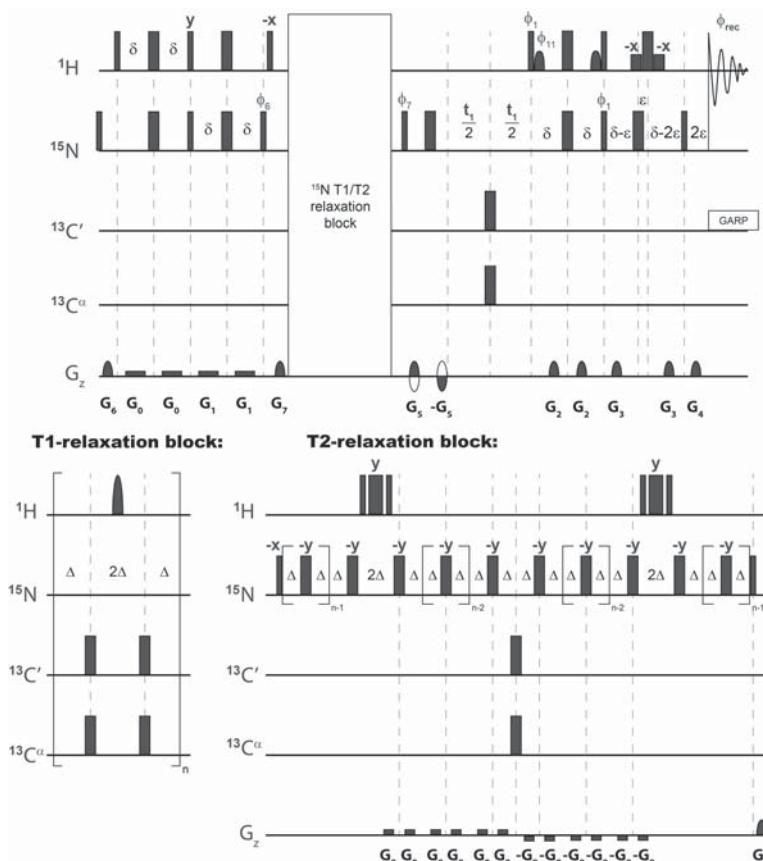
### 2.2.3.5 Relaxation data experiments

The recording and analysis of  $^{15}\text{N}$  relaxation data is part of recent joint publication<sup>[115]</sup> and is therefore in big parts reproduced from the Supporting Information of that publication.

The TROSY-based pulse sequence for measurement of  $^{15}\text{N}$ - $R_2$  relaxation rates was adapted from the original  $R_{1\rho}$  pulse program<sup>[116]</sup> with an appropriate CPMG block introduced. (See Figure 2.1.) The sequence was tested on  $^{15}\text{N}$ -labelled ubiquitin and yielded within the experimental accuracy the same  $^{15}\text{N}$ - $R_2$  rates as determined with HSQC-based schemes (data not shown).<sup>[117,118]</sup>  $^{15}\text{N}$ - $R_1$  rates were measured using the published TROSY-based pulse sequence (see Figure 2.1).<sup>[116]</sup>  $^{15}\text{N}$ - $R_1$  and  $^{15}\text{N}$ - $R_2$  relaxation experiments were recorded on 600 and 700 MHz Bruker Avance III and



900 MHz Bruker Avance I spectrometers equipped with cryogenic probes. For all measurements the temperature was carefully adjusted to 37 °C using a standard sample or ethylene glycol. To achieve nearly complete signal recovery between experiments, the recycle delay was set to 5 s.



**Figure 2.1: TROSY-based NMR pulse sequences for measurement of  $^{15}\text{N}$ - $R_1$  and  $^{15}\text{N}$ - $R_2$  relaxation rates in high-molecular weight proteins.** For each experiment the appropriate temperature compensation block (T-comp) was used (not shown in the scheme for clarity). Narrow and wide rectangles correspond to 90° and 180° flip angle pulses, respectively. The lower amplitude pulses marked -x are 90° pulses with low power, shaped pulses with low amplitude are shaped pulses, all of these pulses are serving as water flip-back pulses.<sup>[119]</sup> Delay durations are  $\delta = 2.65$  ms and  $\epsilon = G_4$ . Gradients are  $G_0$  (2.65 ms, 2.1 G/cm),  $G_1$  (2.65 ms, 1.4 G/cm),  $G_2$  (300  $\mu$ s, 7 G/cm),  $G_3$  (1 ms, 35 G/cm) and  $G_9$  ( $\Delta$ , 0.3 G/cm) with rectangular shape and  $G_4$  (60.8  $\mu$ s, 23 G/cm),  $G_5$  (300  $\mu$ , -23 G/cm),  $G_6$  (1 ms, 19.6 G/cm),  $G_7$  (200  $\mu$ s, -35 G/cm) and  $G_8$  (200  $\mu$ s, 28 G/cm) with sine-bell shape. Phase cycling is done following the scheme  $\phi_6 = 4(y), 4(-y)$ ;  $\phi_7 = y, x, -y, -x$ ;  $\phi_1 = y$ ;  $\phi_{\text{rec}} = y, -x, -y, x, -y, x, -x$ . Quadrature detection was implemented by using the Rance-Kay echo/anti-echo scheme<sup>[120]</sup> with the polarity of the gradients  $G_5$  and  $-G_5$  inverted, and phase  $\phi_7 = y, -x, -y, x$  and  $\phi_1 = -y$  for the second FID generated for each quadrature pair. (As described by Lakomek *et al.*<sup>[116]</sup>) The 180° pulse on  $^1\text{H}$  in the middle of the  $T_1$ -relaxation block is a IBURP2 type pulse with a duration of 2 ms at 600 MHz.  $\Delta$  in the  $T_2$ -relaxation block was set to 1.0 ms.



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### 2.2.3.6 Relaxation data analysis

The Relaxation rates were obtained by 2-parameter non-linear mono-exponential fits of the obtained signal intensities over the applied relaxation delays using the MacDonald algorithm.<sup>[121]</sup> Errors were determined from the variance covariance matrices. Only non-overlapping signals from the 2D <sup>1</sup>H-<sup>15</sup>N TROSY experiments were selected for further analysis resulting in 179  $R_2/R_1$  pairs at each magnetic field.

In the formulation of model-free approaches (MFA), spectral density functions describe the overall tumbling as isotropic with a correlation time  $\tau_R$ . Because even small degrees of motional anisotropy influence relaxation rates, reorientational anisotropy has to be taken into account to avoid determination of false parameters of internal motion. To this end, model-free approach spectral density functions are combined with a spectral density function, which describes anisotropic molecular tumbling. The latter comprises five terms and is given as:<sup>[122]</sup>

$$J(\omega) = \sum_{i=1}^5 A_i \frac{\tau_i}{1 + (\omega\tau_i)^2} \quad (2.4)$$

The correlation times  $\tau_i$  are expressed by the principal components,  $D_k$ , of the rotational diffusion tensor according to  $\tau_1 = (4D_1 + D_2 + D_3)^{-1}$ ,  $\tau_2 = (D_1 + 4D_2 + D_3)^{-1}$ ,  $\tau_3 = (D_1 + D_2 + 4D_3)^{-1}$ ,  $\tau_4 = 6[D + (D^2 - L^2)^{1/2}]^{-1}$  and  $\tau_5 = 6[D + (D^2 - L^2)^{1/2}]^{-1}$  where  $D = (D_1 + D_2 + D_3)/3$  and  $L^2 = (D_1D_2 + D_2D_3 + D_3D_1)/3$ . A useful way of representing the diffusion tensor is to calculate the overall rotational correlation time  $\tau_c$ , the anisotropy  $A$  and the rhombicity  $\eta$  according to:

$$\begin{aligned}\tau_c &= 1/[2(D_x + D_y + D_z)] \\ A &= 2D_z/(D_x + D_y) \\ \eta &= 1.5(D_y - D_x)[D_z - 0.5(D_y + D_x)]\end{aligned}$$

with  $D_x \leq D_y \leq D_z$ . Directional factors  $A_i$  describe the orientation of the relaxation vector in the molecule-fixed coordinate system in terms of the directional cosines  $l$ ,  $m$  and  $n$ :  $A_1 = 3m^2n^2$ ,  $A_2 = 3l^2n^2$ ,  $A_3 = 3m^2l^2$ ,  $A_4 = (d - e)/2$  and  $A_5 = (d + e)/2$  where  $d = 0.5[3(l^4 + m^4 + n^4) - 1]$ ,

$e = [\delta_1(3l^4 + 6m^2n^2 - 1) + \delta_2(3m^4 + 6l^2n^2 - 1) + \delta_3(3n^4 + 6l^2m^2 - 1)]/6$  and  $\delta_i = (D_i - D)/(D^2 - L^2)^{1/2}$ .

$A_i$  is normalized according to  $A_1 + A_2 + A_3 + A_4 + A_5 = 1$ . The model-free spectral density function of an anisotropically tumbling molecule according to the formalism by Lipari and Szabo<sup>[123,124]</sup> can thus be rewritten as

$$J^{MFA}(\omega) = \frac{2}{5} \sum_{i=1}^5 A_i \left[ \frac{S^2 \tau_i}{1 + (\omega \tau_i)^2} + \frac{(1 - S^2) \tau_{int,i}}{1 + (\omega \tau_{int,i})^2} \right] \quad (2.5)$$

where  $1/\tau_{int,i} = 1/\tau_i + 1/\tau_{int}$ .

The spectral density function for the extended model-free approach (EMFA) of an anisotropically tumbling molecule is then (assuming the existence of an additional slow motion, with the order parameter  $S_s^2$  and its internal time  $\tau_{int,s}$  being outside the extreme narrowing limit and slower than  $S_f^2$  but faster than molecular tumbling):

$$J^{EMFA}(\omega) = \frac{2}{5} \sum_{i=1}^5 A_i \left[ \frac{S_f^2 S_s^2 \tau_i}{1 + (\omega \tau_i)^2} + \frac{(1 - S_{f,i}^2) \tau_{f,i}}{1 + (\omega \tau_{f,i})^2} + \frac{S_f^2 (1 - S_{s,i}^2) \tau_{s,i}}{1 + (\omega \tau_{s,i})^2} \right] \quad (2.6)$$

where  $1/\tau_{k,i} = 1/\tau_i + 1/\tau_k$ . The indices  $f$  and  $s$  correspond to fast and slow internal motions, respectively.

The experimental  $^{15}\text{N}-R_1$  and  $^{15}\text{N}-R_2$  rates of E73V hVDAC1 from three magnetic fields (proton frequency of 600, 700 and 900 MHz) were fitted according to a 2-parameter MFA with the order parameter  $S^2$  and the time scale for internal motions  $\tau_{int}$  as a model of internal motion. To this end, a grid search was performed, searching for the minimum in the global target function  $\chi$ , which is given by the weighted sum of squared residuals:

$$\chi = \sum_{i=1}^N \Gamma_i = \sum_{i=1}^N \sum_{j=1}^M [(P_{ij,exp} - P_{ij,calc})^2 / \sigma_{ij}^2] \quad (2.7)$$

where the sum is over  $M$  relaxation data for each of the  $N$  residues of E73V hVDAC1, and  $P_{ij,calc}$  are the appropriate relaxation parameters calculated from the assumed model of internal motion.  $\sigma_{ij}$  are the standard deviations of the experimentally derived  $P_{ij,exp}$ . The global target function consists

of the sum of partial target function  $\Gamma_i$ , which characterize the fit quality for the i-th residue.<sup>[125]</sup> The simple 2-parameter MFA analysis fully reproduced the experimental input. The finding that the 2-parameter model is sufficient for description of the ps–ns time scale dynamics of E73V hVDAC1 is in agreement with the magnetic field dependence of the  $^{15}\text{N}$ - $R_2$  rates for each individual residue.

#### 2.2.3.7 Structure calculation and refinement

Structure determination of E73V hVDAC1 was performed in two steps using the Xplor-NIH software platform<sup>[126]</sup>. First, an initial set of conformations was obtained by folding the extended polypeptide chain against backbone torsion angles and side-chain anglex  $\chi_1$  restraints, which were estimated by TALOS-N<sup>[127]</sup> on the basis of the experimental chemical shifts, together with distance restraints from a 3D  $^{15}\text{N}$ -edited HMQC-NOESY experiment, as well as hydrogen bond restraints that were verified by H/D exchange experiments. Subsequently the lowest-energy structures were taken for further refinement.

In this second step, the diffusion tensor parameters  $\tau_c$ ,  $A$  and  $\eta$  were used as input in Xplor-NIH for structure refinement against the experimental  $^{15}\text{N}$ - $R_2/R_1$  rates.<sup>[126,128–131]</sup> We determined initial values for  $\tau_c$ ,  $A$  and  $\eta$  through the model-free approach described above. This requires the coordinates of the protein. We used the X-ray structure of mVDAC1 (PDB ID: 3EMN)<sup>[46]</sup> for that purpose. The resulting tensor parameters were used for refinement of the 3D structure of E73V hVDAC1 yielded in the first step against 124  $R_2/R_1$  ratios in Xplor-NIH. During this refinement step also multidimensional torsion angle data base potentials (torsionDB<sup>[130]</sup>) and backbone H-bond data base potentials (HBDB<sup>[131]</sup>) were used. The calculated ensemble of lowest energy conformations was well converged. Next, the lowest energy conformer was selected, the directional cosines were recalculated, and the experimental relaxation rates were again fitted with the 2-parameter MFA model. This resulted in similar tensor parameters. The refined set of tensor parameters was again used for structure calculation and yielded virtually the same coordinates.



## 2.2.4 BLM measurements

Bilayer measurements on various hVDAC1 mutants were performed by Roland Benz (Department of Life Sciences and Chemistry, Jacobs-University gGmbH, Bremen) based on a long-standing collaboration. The measurements in the planar phospholipid bilayer membranes were generally performed as described.<sup>[24]</sup> 5 µL of a solution of 1 % DiPhPC in n-decane were painted over the hole (area ≈ 0.4 mm<sup>2</sup>) of a Teflon cuvette, creating lipid bilayers of the Mueller-Rudin type.<sup>[30]</sup> A solution of 1 M KCl, 10 mM HEPES, pH 6.0 was used as electrolyte. hVDAC1 mutants in LDAO were diluted with 1 % (w/v) Genapol to a final concentration of approximately 5 µM and mixed 1:1 with cholesterol powder suspended in 1 % (w/v) Genapol. Aliquots of VDAC (typically 1–5 µL) were added to both cuvette chambers at a standard membrane potential of 20 mV. The insertion of single channels was monitored by the increase of the current measured with the Ag/AgCl electrodes and an in-house amplifier.

Measurements with variable potentials were performed by equilibrating the system for 1–2 hours until no significant increase in current (tantamount to no further channel insertions) was observed anymore. The steady-state voltage-dependent closure of the channels was measured by increasing the potential in steps of ± 10 mV and calculating the ratio of the final (i.e. after closure) and initial (i.e. before closure) conductance at each applied voltage step.

## 2.2.5 Calculation of PRE effect differences

Due to the noise influenced results yielded from the PRE experiments of Quintuple hVDAC1 and A2C-Quintuple hVDAC1, respectively, the intensity changes induced due to the MTSI labeling were enhanced the following way:

For each mutant pair (A2C-Quintuple & Quintuple hVDAC1 and A2C/E73V/C127A/C232S & E73V/C127A/C232S hVDAC1), the intensity ratios  $I(\text{Quint})$  and  $I(\text{wt})$  were calculated for each residue. Based on residues which didn't show broadening due to the attached MTSI label, these ratios were normalized to 1. The difference in the intensity ratios was then calculated for each residue as  $\Delta(I)_i = I(\text{Quint})_i - I(\text{wt})_i$ .



## 2 Materials & Methods

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The differences of the intensity ratios were pooled in groups of five residues and normalized:

$$\Delta(I, \text{grouped})_j = \frac{\sum_{i=5j-4}^{5j} \Delta(I)_i}{5} \quad (2.8)$$

To further reduce the effects of the noise in the data, a binomial filter was applied to the floating average of three of these grouped intensity ratio differences:

$$\Delta(I, \text{filtered})_n = \frac{\Delta(I, \text{grouped})_{n-1} + 2 \cdot \Delta(I, \text{grouped})_n + \Delta(I, \text{grouped})_{n+1}}{4} \quad (2.9)$$

After all, these  $\Delta(I, \text{filtered})$  values were plotted as function of the n-th five residue group. (See Chapter 3.4.4.)

# 3 Results

## 3.1 Improving the assignment of hVDAC1

VDAC backbone resonance assignment has been a challenge over the years. In 2007, 70/90 % (overall/barrel) of the backbone amide resonances of *wt* hVDAC1 were reported to be assigned in micellar solution by Monika Bayrhuber.<sup>[64,70]</sup> With the help of further experiments and due to improved sample quality, this could be increased to 85/93 % and 88/97 % (overall/barrel) for *wt* hVDAC1 and E73V hVDAC1, respectively, by Saskia Villinger.<sup>[74,132]</sup> This improvement was also made possible by chemical shift information obtained from solid-state NMR data, which delivered the necessary information to assign three residues in the N-terminal  $\alpha$ -helix (Lysine-12, Glutamate-16 and Valine-17).<sup>[113]</sup>

In this work, the existing assignments were reviewed completely and – supported by additional recorded spectra – improved, leading to  $C_\beta$  shifts, improved backbone assignment (see Chapter 3.1.2), and a multitude of newly assigned  $H_N$ - $H_N$  NOE contacts (see Chapter 3.1.3) that were used in the calculation of a 3D structure of E73V hVDAC1 (see Chapter 3.2).

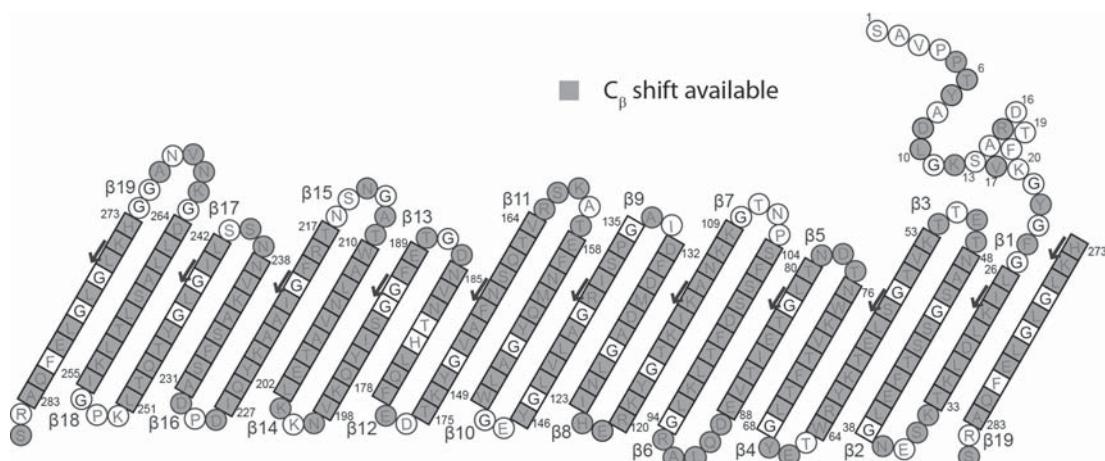
### 3.1.1 Additional $C_\beta$ shifts

In previous studies in this working group, even in fully  $^{13}C$  labelled samples  $C_\beta$  resonance assignments for hVDAC1 in LDAO micelles were missing and have never been reported. Thus only the  $C_\alpha$  resonances were available to obtain sequential information to assign the backbone resonances of the protein.<sup>[64,74]</sup> At the same time,  $C_\beta$  assignments were reported in the literature for samples in liquid state (hVDAC1 reconstituted in LDAO micelles)<sup>[48]</sup> as well as in solid state (hVDAC1 in

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DMPC liposomes).<sup>[113]</sup> Hence, it is not the protein itself which makes it generally impossible to obtain C<sub>B</sub> shifts. It could only possibly be the complete system consisting of buffer, micelle and protein which for some reason prevents the detection of C<sub>B</sub> resonances. If this is not the case, with the right technical approach the determination of C<sub>B</sub> shifts should be possible.

Indeed it was found, that using a E73V/C127A/C232S hVDAC1 mutant and a slightly modified version of the standard Bruker pulse program (i.a. a shorter  $^1\text{H}$ - $^{15}\text{N}$ -INEPT delay, 1.75 ms instead of 2.3 ms (which corresponds to  $1/(4J_{NH})$ )), recording a 3D TROSY-HNCACB<sup>[109,110]</sup> was possible in 2 days and 14 hours with excellent results. The pulse program used can be found in Appendix A.1.



**Figure 3.1: Map of the existing C<sub>β</sub> assignments in E73V/C127A/C232S hVDAC1.** Residues with available C<sub>β</sub> shift are colored in gray, Glycine residues are lettered in black. Residues in β-strands are boxed, residues in the loops and the N-terminal α-helix are circled. The topology map was adopted and modified from previous works [46,74]

The newly acquired HNCACB data helped to verify the existing backbone assignments. Indeed, the known assignments could be confirmed nearly completely. (See Chapter 3.1.2 for details) The current state of the assignment comprises 87.4 % of the existing  $C_\beta$  in the complete protein and 98.3 % in the barrel, respectively (values given for E73V/C127A/C232S hVDAC1). (See Figure 3.1) In the barrel, only Histidine-181, Threonine-182 and Phenylalanine-281 are unassigned and missing.

In the case of Histidine-181, the missing assignment is caused by the low signal strength of the corresponding HNCACB peaks that made it impossible to find the correct resonances. Threonine-182 is



missing completely in all spectra and the existing assignment of Phenylalanine-281 for *wt* hVDAC1 actually has a possible equivalent in the E73V/C127A/C232S hVDAC1 spectra, but the corresponding HNCACB peaks do not fit to the previously reported values.<sup>[74]</sup>

In comparison to the assignment statistics reported by Hiller *et al.*<sup>[48]</sup>, 221 instead of 194 existing C<sub>β</sub> are assigned. In particular, the assignment in the second part of the N-terminal helix and the assignments around Glutamate-73 or Valine-73, respectively, were drastically improved. Also, in most of the loops more residues are assigned now than reported by them.

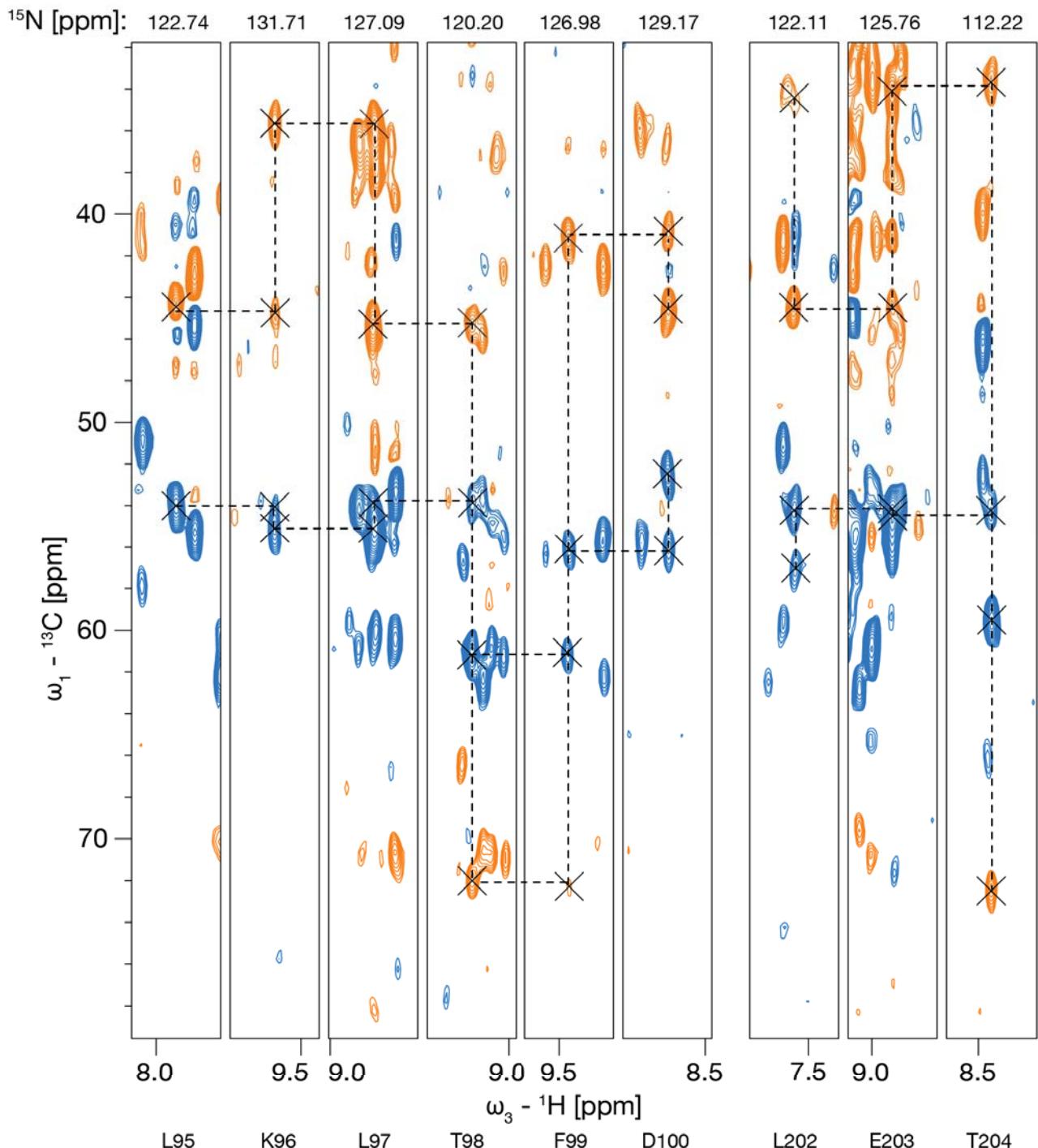
### 3.1.2 Improved backbone assignment

In previous works in this group, great effort had already been put into the creation and improvement of the backbone assignment of hVDAC1. β-barrel membrane proteins can be characterized relatively well by NMR and it is the helical membrane proteins that require more effort<sup>[133]</sup>. However, still there are regions of hVDAC1 that proved to be notoriously difficult to assign. Most prominent are the region around Glutamate-73 and the N-terminal helix. In the former, in the case of *wt* hVDAC1 severe peak broadening is caused by chemical exchange<sup>[132]</sup>, which can be “solved” by the E73V mutation, making seven more residues in the β-strands 4 and 5 detectable (Threonine-72 to Tryptophan-75 and Glycine-82 to Glutamate-85). In the latter, the backbone assignment was improved with chemical shift information from solid-state NMR data.<sup>[74]</sup> Furthermore, also the region around Threonine-182 as well as all loops between the β-strands exhibit broadened peaks. For the loops this is obviously due to their increased conformational flexibility. In particular, loops with a Proline (loop from β-strand 6 to β-strand 7, β-strand 15 to β-strand 16 and β-strand 17 to β-strand 18) are problematic in terms of assignment, as Prolines are missing the amide proton and thus always induce breaks in the sequential connectivity.

In detail, for assignment of hVDAC1 several triple-resonance experiments were recorded (TROSY-HNCA, TROSY-HN(CO)CA, TROSY-HNCO), labeling with MTSL at various places to yield PRE data and amino acid selective labeling were done.<sup>[64]</sup> The assignment was improved with the help of triple-resonance experiments on E73V hVDAC1 (TROSY-HNCA and <sup>15</sup>N-NOESY-HMQC).<sup>[74]</sup> Since these developments took place over the course of 10 years it is not anticipated that the assign-

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ment will be improved drastically in the future. Nevertheless, especially with the help of the newly recorded TROSY-HNCACB spectral data further improvement of the assignment was achieved in this work.



**Figure 3.2: Strips of TROSY-HNCACB on E73VC127AC232S hVDAC1.** Generic backbone walk shown through the example of Leucine-95 to Aspartate-100 (left). Backbone walk that lead to the assignment of Leucine-202 (right). Positive contours are shown in blue, negative contours are shown in orange.

With the help of the additional  $C_\beta$  shift information, Leucine-202 could now be assigned. (See Figure 3.2) Interestingly, because of the very similar  $C_\alpha$  shifts of Leucine-202 and Glutamate-203, the resonance in question had previously been assigned to Glutamate-203. This assignment had already been revised prior to the beginning of this PhD thesis, but shows that it was known that this resonance should be located in that region of the beginning of  $\beta$ -strand 14.

The loop from  $\beta$ -strand 2 to  $\beta$ -strand 3 had been tentatively assigned before. As it contains three Threonine residues (49, 51 and 52), the additionally recorded TROSY-HNCACB proved to be very helpful, since Threonine has a unique pattern with the  $C_\beta$  resonating at lower field than the  $C_\alpha$ . This way, the previously tentative resonance assignments could now be replaced by safe assignments, at the same time confirming the NOE connectivities (see Figure 3.3).

A similar case are Leucine-91 and Alanine-92. Here, it is the unique  $C_\beta$  resonance position of Alanine-92, which helped to identify the resonances of these residues with more certainty. By this very same strategy the sequence of the three residues Threonine-250, Leucine-251, and Lysine-252 could additionally be assigned safely. The last additional residue found is Glutamate-177.

Regarding the N-terminal helix, the TROSY-HNCACB helped to confirm the assignments of all previously assigned residues except Alanine-8 and Alanine-14. In these two cases the characteristic  $C_\beta$  resonance is missing. This, however, does not prove the assignments to be wrong, as in the spectrum the Alanine- $C_\beta$  resonances are always quite weak. Taken together with the overall low signal strength of the residues in the N-terminal helix it is not unlikely, that these two resonances in question are simply too weak to detect.

After all, one residue had to be deassigned because it never behaved as expected. Lysine-20 had no connections to surrounding residues in the NOESY-spectrum and furthermore does not react at all to mutations in its close proximity (G21V/G23V/E73V/C127A/C232S hVDAC1 and derivatives of this mutant). As it had been assigned only ambiguously, it was decided to remove this assignment from the list.

Overall, the backbone assignment in hVDAC1 was increased to 88.1 %/94.8 % and 92.1 %/98.4 % (overall/barrel) of the residues for *wt* hVDAC1 and E73VC127AC232S hVDAC1, respectively. (See also Figure 3.3)

The largest part of the unassigned residues is now located in the last part of the N-terminal helix and the beginning of the linker from the helix to the barrel (Phenylalanine-18 to Glycine-21 and Glycine-23). Also, some loops (from  $\beta$ -strands 1 to 2, 3 to 4, 6 to 7, 13 to 14, 14 to 15 and 17 to 18) and residues in the C-terminal end of the protein (Phenylalanine-281, Glutamate-282, Leucine-284) are still unassigned. For all of these residues even the additional information from the TROSY-HNCACB spectrum did not help finding the assignments. (On the contrary, Lysine-20 was even deassigned.) It is thus doubtful whether they can be assigned with reasonable effort or assigned at all.

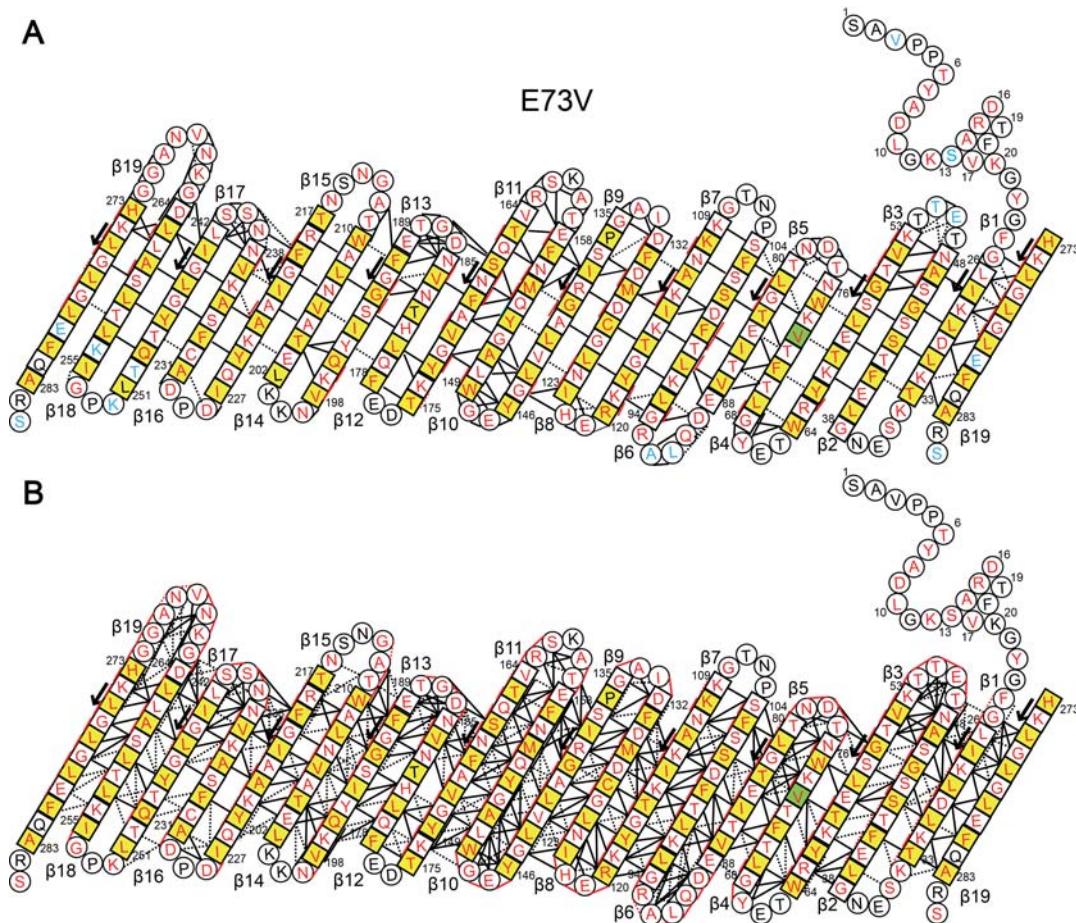
### 3.1.3 Improved backbone NOE network

When revising the data and spectra received from Saskia Villinger, it turned out that the 3D  $^{15}\text{N}$ -NOESY-HMQC of the E73V hVDAC1 mutant was not completely assigned at all.

As can be seen in Figure 3.3, a significant amount of the NOE contacts in the spectrum was previously unassigned. Taken together, there were 534 NH-NH contacts assigned by Saskia Villinger (counting each resonance as a single contact, meaning that a  $\text{H}_\text{N}$ - $\text{H}_\text{N}$  NOE that is visible in both directions is counted as two contacts).

By thorough analysis of the spectrum and by purposeful search for contacts that would be expected to be visible, 900 new  $\text{H}_\text{N}$ - $\text{H}_\text{N}$  contacts could be found, raising the number of the overall observed contacts to 1431 (a small number of contacts was deleted) and thus increasing the number of assigned contacts nearly threefold.

As depicted in Figure 3.3, the newly assigned NOE contacts are well spread over the whole barrel. Not only are there numerous new sequential short range contacts found in all  $\beta$ -strands, but also a big amount of newly assigned mid and long range inter-strand contacts. Where previously often only the strong  $\text{H}_\text{N}$ - $\text{H}_\text{N}$  contacts indicating hydrogen bonds were assigned, now basically every residue has additionally assigned inter-strand contacts. The highest density of  $\text{H}_\text{N}$ - $\text{H}_\text{N}$  NOEs can be found in  $\beta$ -strands 5 to 11, where roughly 600 contacts are located alone – more than previously had been assigned in the whole protein. These  $\beta$ -strands are comprising the region where the N-terminal  $\alpha$ -helix is attached to the barrel wall (hydrophobic contact patch of Valine-143 in  $\beta$ -strand 9 and



**Figure 3.3: NOEs of E73V hVDAC1.** Overview of the  $\text{H}_\text{N}$ - $\text{H}_\text{N}$  NOEs obtained from a NOESY-HMQC spectrum of E73V hVDAC1 recorded by Saskia Villinger. Assigned residues are lettered in red, ambiguously assigned residues in blue and unassigned residues are lettered in black. Residues in  $\beta$ -strands are boxed, residues in the loops and the N-terminal  $\alpha$ -helix are circled. Yellow shaded residues are pointing to the micelle, the mutation site E73V is highlighted in green. Solid black (solid red) lines indicate inter-strand (sequential)  $\text{H}_\text{N}$ - $\text{H}_\text{N}$  NOEs observed in both directions, dotted black (dotted red) lines indicate inter-strand (sequential) NOEs observed only in one direction and overlapping NOEs. A: Assignment and NOE network as assigned and reported by Saskia Villinger. (This part of the figure was adapted from the PhD thesis of Saskia Villinger.<sup>[74]</sup>) B: Latest and most up to date assignment and NOE network.

Leucine-150 in  $\beta$ -strand 10 to Leucine-10 in the helix) and the  $\beta$ -strands between said region and the Valine-73 mutation site. In the  $\beta$ -strands 15 to 17 however, the network of NOEs is visibly least dense. This is in line with previous results, that showed that the average peak intensity for residues located in the N-terminal  $\beta$ -strands is significantly lower than those of the central  $\beta$ -strands where E73V mutation and the N-terminal  $\alpha$ -helix are stabilizing the barrel.<sup>[132]</sup>

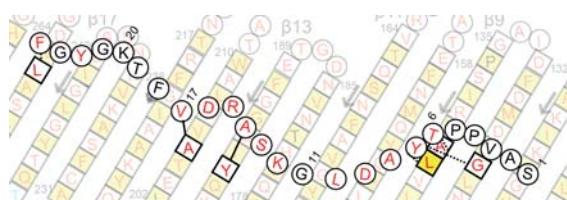
Interestingly, also the large number of newly assigned contacts did not result in a possible assignment for Threonine-182. Following the pattern of strong  $\text{H}_\text{N}$ - $\text{H}_\text{N}$  contacts indicating hydrogen bonds

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there should be a strong NOE from Valine-171 to Threonine-182 and *vice versa*. However, no such NOE could be found, leaving the assignment of Threonine-182 an open question.

There are also some new contacts between residues in the N-terminal  $\alpha$ -helix found. The number of internal contacts in the helix was increased from 21 to 32 inter residue contacts plus 9 intra residue contacts. For example, there are four new contacts from and to Lysine-12 alone, which had no contacts before at all. (Data not shown here.)



**Figure 3.4: NOEs of E73V hVDAC1 between N-terminal  $\alpha$ -helix and  $\beta$ -barrel.** Assigned residues are lettered in red and unassigned residues are lettered in black. Residues in  $\beta$ -strands are boxed, residues in the loops and the N-terminal  $\alpha$ -helix are circled. Yellow shaded residues are pointing to the micelle. Solid black lines indicate  $H_N$ - $H_N$  NOEs observed in both directions, dotted black lines indicate NOEs observed only in one direction. Residues in the barrel without contact to helix residues are greyed out for clarity.

Finally, the contacts between N-terminal  $\alpha$ -helix and  $\beta$ -barrel were completely revised and improved. Although there are still only 7 contacts between  $\alpha$ -helix and  $\beta$ -barrel, these contacts can be translated into constraints that are not violated when calculating a 3D structure based on the NOE data (see Chapter 3.2). From the previously existing three contacts between  $\alpha$ -helix and  $\beta$ -barrel, two had to be removed (Threonine-6 to Methionine-155 and *vice versa*) because they were violating the structure and the contact between Alanine-14 and Isoleucine-194 was altered to be a better fitting contact between Alanine-14 and Tyrosine-195 that is now assigned in both directions. An overview of the contacts between  $\alpha$ -helix and  $\beta$ -barrel is shown in Figure 3.4.



## 3.2 A new NMR structure of E73V hVDAC1

As mentioned above in Chapters 3.1.2 and 3.1.3, not only the assignment of hVDAC1 could be improved to a previously unprecedented level, but also the NOE network in E73V hVDAC1 could be improved drastically. In connection with  $^{15}\text{N}$  spin-relaxation data from E73V hVDAC1 and by once again revising the E73V hVDAC1 H/D exchange data, this was used to determine a 3D structure of E73V hVDAC1 based solely on NMR data that by far exceeds the accuracy and quality of the previously published NMR structure of hVDAC1<sup>[48]</sup>.

The following chapter describes how this was done and achieved. This chapter of this PhD thesis is part of a recent collaborative publication<sup>[115]</sup> and thus parts of this chapter are reproduced from this publication and contain some identical text passages. The calculations described in the following were performed by Mariusz Jaremko, Łukasz Jaremko and Markus Zweckstetter.

### 3.2.1 A 3D structure of E73V hVDAC1 based on the improved NOE network as starting point<sup>1</sup>

In previous works, to compensate for the sparsity of long-range structural information based on NOE-based interproton restraints (due to the fact that hVDAC1 in micelles is a system so large it requires deuteration<sup>[134–136]</sup>), it was tried to record residual dipolar couplings (RDCs), as they often prove useful in that purpose.<sup>[137,138]</sup> However, it turned out to be impossible to align hVDAC1 in LDAO micelles in liquid crystalline alignment media. In charged polyacrylamide gels the partial alignment of hVDAC1 in LDAO micelles was possible, but overall the resulting spectra and data were of poor quality.<sup>[74]</sup>

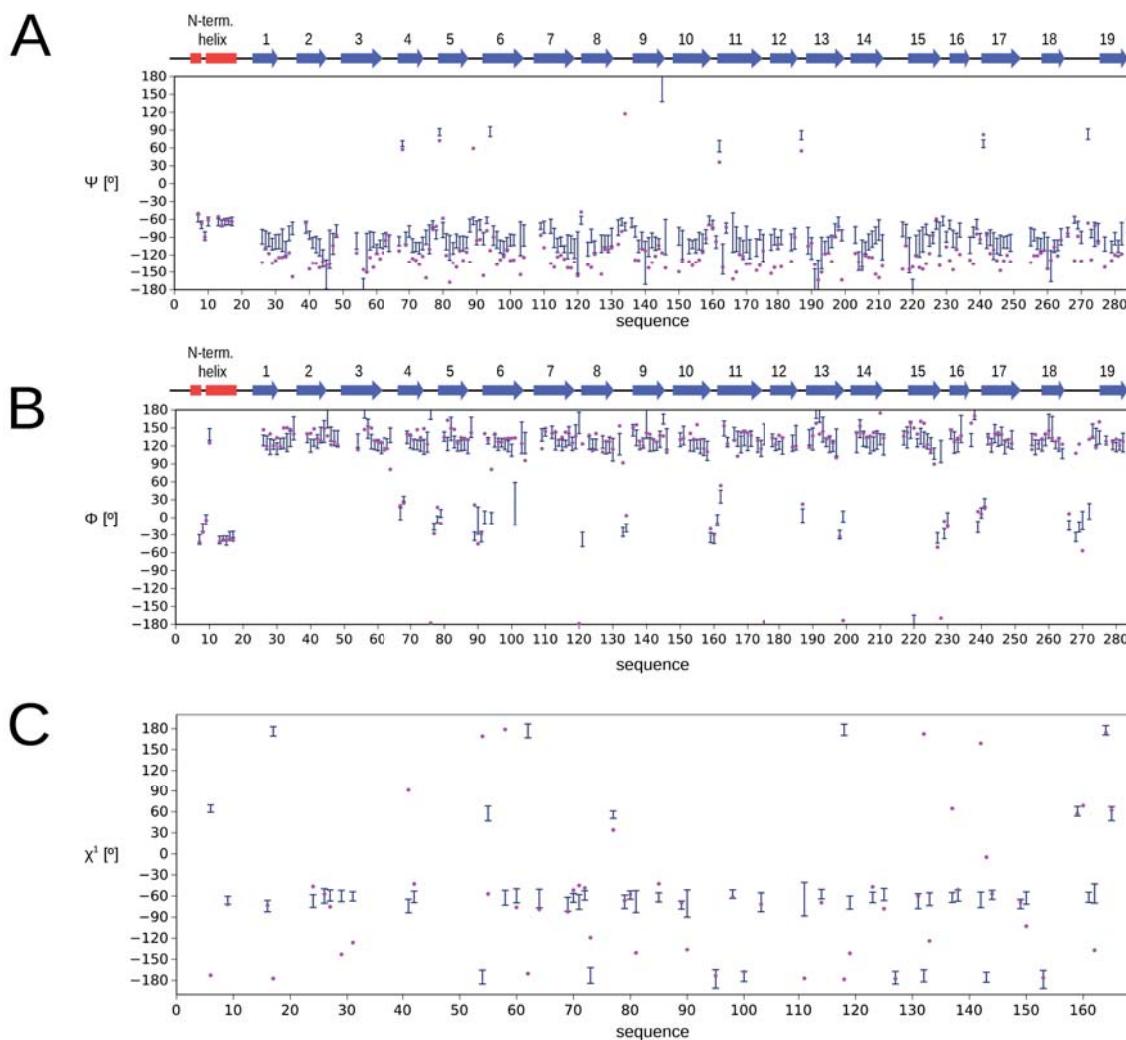
These limitations were addressed by exploiting the long-range order that is encoded in the nuclear magnetic spin relaxation of macromolecules like hVDAC1 in LDAO micelles due to their anisotropic rotational diffusion.<sup>[129,139]</sup> (This approach was previously never applied to membrane proteins.) A combination of these information from heteronuclear relaxation rates together with NOE data and the

<sup>1</sup>This section is based on a recent joint publication<sup>[115]</sup> and contains akin text passages.

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application of empiric potentials to hydrogen bonds determined from the aforementioned E73V hVDAC1 H/D exchange data<sup>[131]</sup> lead to a structure of E73V hVDAC1 in unprecedented resolution.

Based on the improved assignment of 92.1 % of the backbone H<sup>N</sup> and N resonances and the corresponding C<sub>α</sub> and C' shifts, the backbone torsion angles  $\psi$ ,  $\phi$  and the side-chain angle  $\chi^1$  were estimated using TALOS-N.<sup>[127]</sup> (See Figure 3.5.)



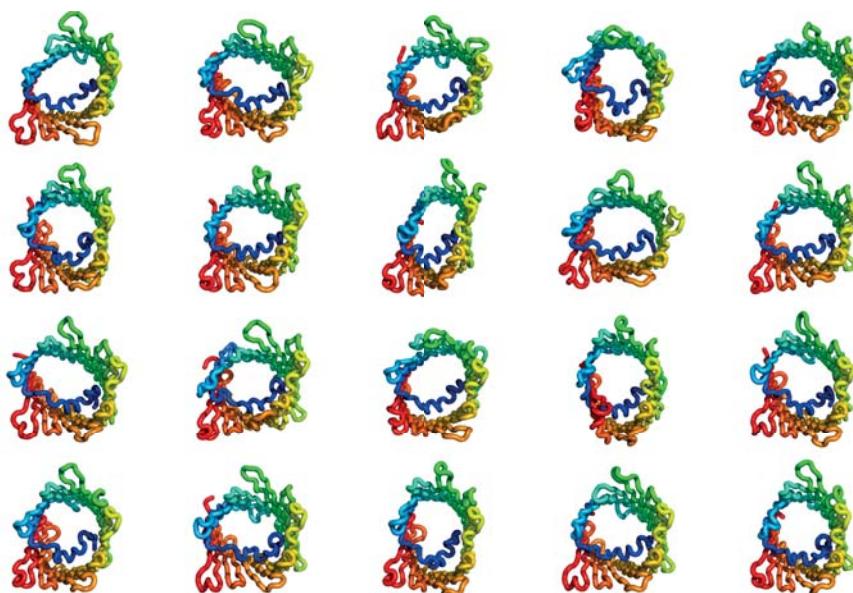
**Figure 3.5: Torsion angle information from experimental chemical shifts.** Angles on the basis of the experimental backbone H<sup>N</sup>, N, C<sub>α</sub>, C' chemical shifts in E73V hVDAC1. Pink dots show the values observed in the crystal structure of mVDAC1. (PDB ID: 3EMN)<sup>[46]</sup> A: Backbone dihedral angle  $\psi$ . B: Backbone dihedral angle  $\phi$ . C: Backbone torsion angle  $\chi^1$ . Figure was adapted from our joint publication<sup>[115]</sup> under the terms of the Creative Commons License CC BY-NC-ND.

After refinement, the aforementioned NOE data (see Figures 3.3 and 3.4) provided 656 sequential, medium- and long-range proton-proton distance restraints. 128 hydrogen bond restraints from the E73V hVDAC1 H/D exchange data were also present. (Originally from our group and published in



Bayrhuber *et al.*<sup>[70]</sup>, refined data not shown.) The combined data (NOE restraints, hydrogen bond restraints and backbone angle restraints) were used to fold the protein from the extended polypeptide with a simulated annealing protocol using the platform Xplor-NIH.<sup>[126]</sup>

The resulting ensemble of the 20 lowest energy structures had a backbone RMSD of 2.5 Å and like in other previously published NOE-based structures<sup>[48,70]</sup> the definition of the barrel shape and the conformation and position of the N-terminal helix could only be determined with modest accuracy. Nevertheless, in the majority of these 20 structures the N-terminal helix shows the kink between Leucine-10 and Lysine-12, a structural feature that previously had not been described based on NMR data. (See Figure 3.6.)



**Figure 3.6: 20 lowest energy structures.** E73V hVDAC1 calculated exclusively from the NOE- and hydrogen bond-based distance and chemical shift-derived angular restraints. The structures are colored from the N- to the C-terminus from blue to red. Figure was adapted from our joint publication<sup>[115]</sup> under the terms of the Creative Commons License CC BY-NC-ND.

### 3.2.2 Improving the structure with $^{15}\text{N}$ spin-relaxation data<sup>1</sup>

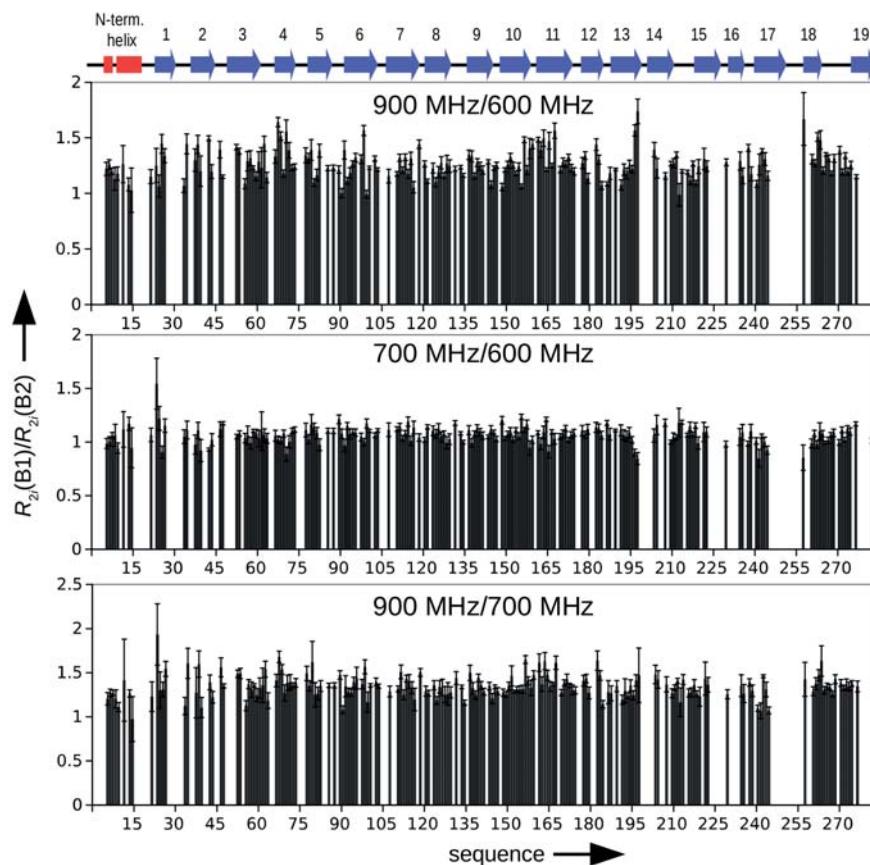
The accuracy and precision of the E73V hVDAC1 could now be further improved by  $^{15}\text{N}$  spin-relaxation data derived from measurements at magnetic fields corresponding to 60 to 90 MHz nitrogen frequency. (The measurements were performed by Saskia Villinger, the analysis of the data was

<sup>1</sup>This section is based on a recent joint publication<sup>[115]</sup> and contains akin text passages.

### 3 Results

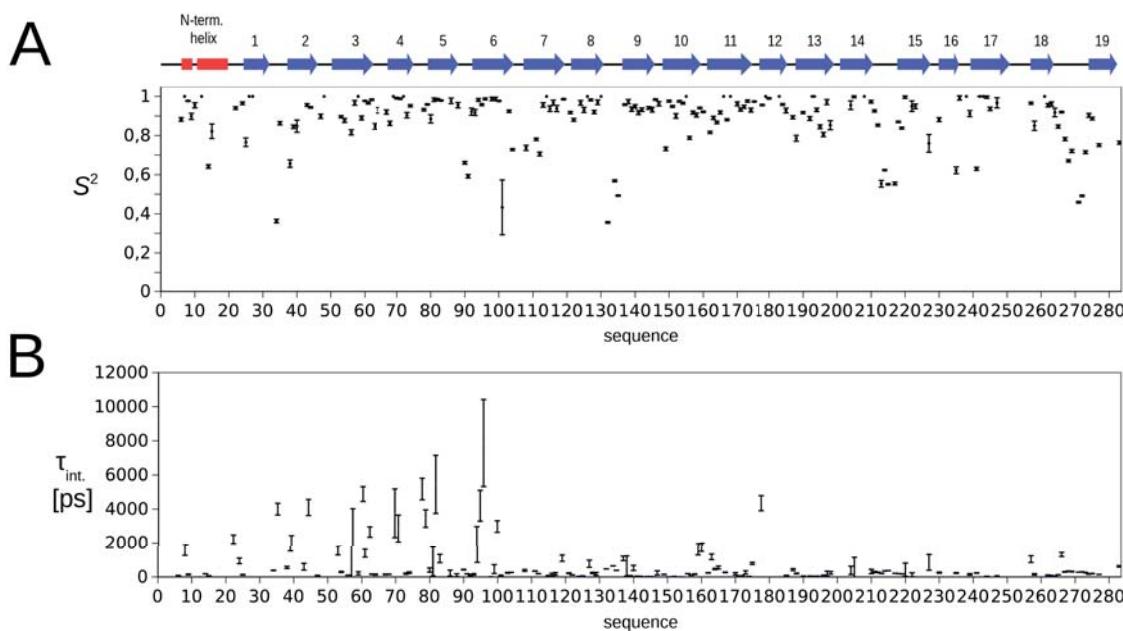
performed by Mariusz Jaremkó, Łukasz Jaremkó and the author of this thesis. The pulse programs used can be found in Appendix A.2 and A.3.) The measurements were performed using the TROSY approach.<sup>[116]</sup> This way, 537 pairs of  $R_1$  and  $R_2$  relaxation rates of E73V hVDAC1 were obtained with average errors of 2.7 % at 60 MHz, 3.1 % at 70 MHz and 4.4 % at 90 MHz.

By measuring at several magnetic fields possible contributions of  $R_{ex}$  (chemical exchange motions slower than the overall tumbling but fast enough to average the chemical shifts, *i.e.* motions in the range from  $\mu\text{s}$  to  $\text{ms}$ ) to the transverse relaxation rate  $R_2$  can be assessed. The contributions from  $R_{ex}$  would enhance the  $R_2$  rates and the enhancement would scale quadratically with the magnetic field.<sup>[140]</sup> Thus, plotting the ratio of  $R_2$  rates at two magnetic fields,  $R_{2(B_1)}/R_{2(B_2)}$ , as a function of the residue number can provide evidence for  $R_{ex}$ . As can be seen in Figure 3.7 there is no evidence of significant contributions from  $R_{ex}$  in E73V hVDAC1.



**Figure 3.7: Dynamic information encoded in  $^{15}\text{N}$  spin-relaxation rates.** Residue-specific ratios of  $^{15}\text{N}$ - $R_2$  relaxation rates [ $R_{2i(B_1)}/R_{2i(B_2)}$ ] from three magnetic fields. Error bars show one SD. Figure was adapted from our joint publication<sup>[115]</sup> under the terms of the Creative Commons License CC BY-NC-ND.

Additionally, with measuring at three magnetic fields the overall quality of the data can better be evaluated and ps–ns motions can be determined with more accuracy.<sup>[141]</sup> By applying MFA analysis<sup>[142]</sup> on the relaxation rates it was demonstrated that the majority of the residues in the 19  $\beta$ -strands of E73V hVDAC1 are rigid on the ps–ns time scale and only a few residues in the N-terminal  $\beta$ -strands had elevated values for fast internal motion. The loops nevertheless showed higher flexibility, in particular the long loop between  $\beta$ -strands 18 and 19. (With rigid residues being defined by a Lipari–Szabo order parameter  $S^2 > 0.85$  and a correlation time for internal motions  $\tau_i < 500$  ps. See Figure 3.8.)

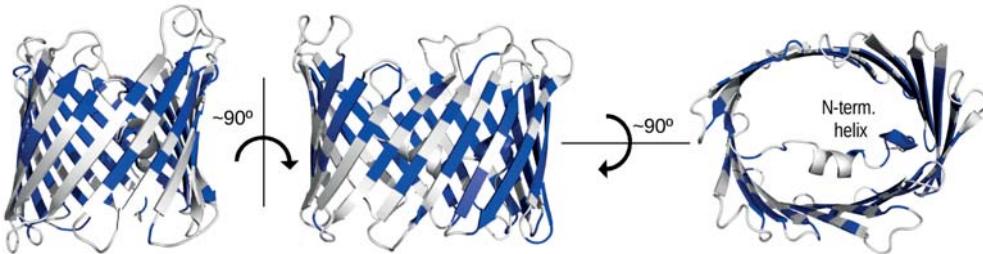


**Figure 3.8: Model-free analysis of the  $^{15}\text{N}$  relaxation data of E73V hVDAC1 (2-parameter MFA model with  $S_f^2$  and  $\tau_{int,f}$  and fully anisotropic reorientation.** A: Residue-specific  $S_f^2$  order parameters. B: Internal total correlation time ( $\tau_{int,f}$ ) describing ps–ns time scale motions of individual residues. Errors were estimated by 200 Monte Carlo simulations and represent one SD. Figure was adapted from our joint publication<sup>[115]</sup> under the terms of the Creative Commons License CC BY-NC-ND.

It was now made use of the fact, that the dependence of the transverse and longitudinal relaxation rates on the rotational diffusion anisotropy of the protein provides information not only on internal motion but also on long-range structure.<sup>[129]</sup> It had already been shown by MFA analysis that E73V hVDAC1 is very rigid in comparison to *wt* hVDAC1.<sup>[132]</sup> Thus, it was expected that information on the long-range order which could be extracted with higher accuracy in E73V hVDAC1. As shown in the last paragraphs, exactly this assumption holds true. The residues with order parameters

### 3 Results

$S^2 > 0.85$ , *i.e.* the most rigid ones, were now selected to be analyzed for that purpose. 124 residues mainly found in the  $\beta$ -strands were selected. This corresponds to 124  $R_1$  and  $R_2$  values at 3 fields, respectively, so all in all 372  $R_1$  and 372  $R_2$  rates or 744 experimental values in total. The respective residues are shown in Figure 3.9.



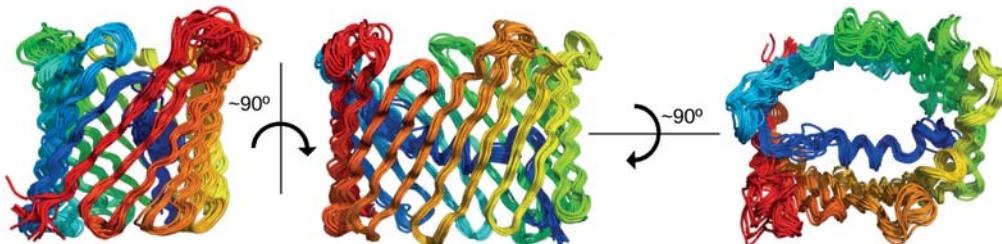
**Figure 3.9:  $^{15}\text{N}$ - $R_2/R_1$  rates used for structure refinement.** Residues for which  $R_2/R_1$  rates were used during structure refinement are shown in blue. Data is mapped on the lowest-energy conformation of E73V hVDAC1 after refinement. Figure was adapted from our joint publication<sup>[115]</sup> under the terms of the Creative Commons License CC BY-NC-ND.

To extract the structural information encoded in the relaxation rates, the rotation correlation time  $\tau_c$ , the anisotropy  $A$  and the rhombicity  $\eta$  of the diffusion tensor have to be known *a priori*. These parameters were estimated by applying MFA analysis<sup>[142]</sup> to the lowest-energy NOE-based structure of E73V hVDAC1 obtained previously (cf. Figure 3.6). This structure of E73V hVDAC1 was then refined against the experimental  $^{15}\text{N}$ - $R_2/R_1$  rates of the 124 previously selected residues. This was done with a specially designed Xplor-NIH protocol that makes use of the knowledge-based potentials torsionDB and HBDB<sup>[130,131]</sup> to optimize the geometry of the torsion angles and hydrogen bonds. The potentials were used to narrow the conformational space in which the protein can move during refinement to the physically most justified conformations as they are found among the highest-quality X-ray structures. Additionally, they prevent overfitting of the orientational restraints, in particular in the case of outliers.<sup>[129]</sup>

To check the robustness and stability of this approach the same calculations were performed with diffusion tensor parameters obtained by MFA analysis of the previously published VDAC1 structures with the PDB IDs 3EMN<sup>[46]</sup> and 2JK4<sup>[70]</sup>. Also, the tensor parameters  $A$  and  $\eta$ , which were derived from the  $R_2/R_1$  refined E73V hVDAC1 structure were taken, altered by  $\pm 0.2$  units and  $-50\%$  and  $+100\%$ , respectively, and the calculation was repeated using these values. As with previous



results with this approach from globular proteins and their complexes<sup>[139,143]</sup>, all these calculations converged to highly similar 3D structures, proving the high robustness and stability of the protocol. The 20 lowest-energy structures of E73V hVDAC1 obtained by refinement as described above are shown in Figure 3.10, the lowest energy structure can be seen in Figure 3.9.



**Figure 3.10: 20 lowest-energy structures of E73V hVDAC1 after refinement against  $R_2/R_1$  rates.** Structures are colored from the N- to the C-terminus from blue to red. Figure was adapted from our joint publication<sup>[115]</sup> under the terms of the Creative Commons License CC BY-NC-ND.

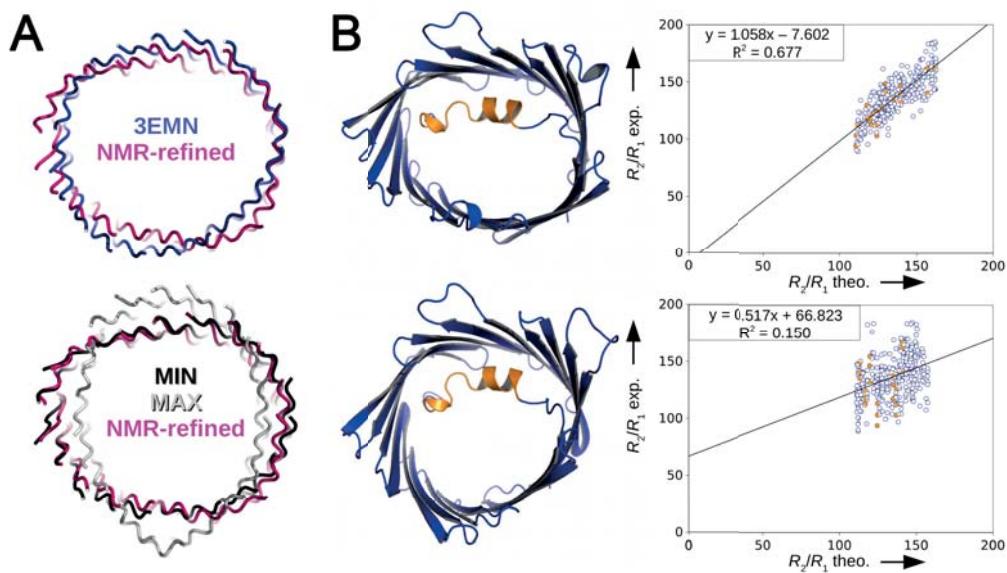
The ensemble of the 20 lowest-energy structures tightly clusters around a single conformation and exhibits a backbone RMSD of 1.1 Å for all 283 residues. Each of the 19 β-strands is well defined and the N-terminal α-helix is rigidly attached to the barrel wall with no sign of flexibility. The most flexible part of the structure is the linker, which connects the N-terminal helix to β-strand 1.

### 3.2.3 Validating the high-resolution structure of E73V hVDAC1<sup>1</sup>

When comparing the high-resolution structure of E73V hVDAC1 described here to the previously reported crystal structure of mVDAC1<sup>[46]</sup>, it becomes obvious that the former is more anisotropic. It can be shown that this is a direct consequence of the experimental  $R_2/R_1$  rates, as the  $R_2/R_1$  rates fit better to the solution structure of E73V hVDAC1 than to the crystal structure of mVDAC1. (See Figure 3.11.)

The largest deviations between the experimental rates and the values back-calculated from the crystal structure of mVDAC1 are found in β-strands 9 and 10. Interestingly, as can also be seen in Figure 3.11, the ellipsoidal shape of the solution structure of E73V hVDAC1 as it is encoded in the  $R_2/R_1$  rates is more similar to an anisotropic barrel shape found during a 100 ns MD trajectory of membrane-embedded mVDAC1<sup>[132]</sup> than to the starting X-ray structure of mVDAC1<sup>[46]</sup>. ( $C_\alpha$

<sup>1</sup>This section is based on a recent joint publication<sup>[115]</sup> and contains akin text passages.



**Figure 3.11: Validation of the high-resolution structure of E73V hVDAC1.** A: Comparisons of the lowest-energy structure of E73V hVDAC1 to previous structures. The new structure (pink) is more anisotropic than the previously published crystal structure of mVDAC1.<sup>[46]</sup> (top) The barrel shape of E73V hVDAC1 (pink) is similar to one of the two maximum excursions (black and grey) observed in a 100 ns MD trajectory of mVDAC1 embedded in a phospholipid bilayer.<sup>[132]</sup> (bottom) B: Left side: Solution structure of E73V hVDAC1 (PDB ID: 5JDP, top) and crystal structure of mVDAC1 (PDB ID: 3EMN, bottom). Right Side: Correlation of experimental  $R_2/R_1$  rates from three magnetic fields (scaled according to the theoretical field dependence) with values back-calculated from the respective structure to the left. The residues of the N-terminal helix together with their determined  $R_2/R_1$  rates on the correlation plots are shown in orange. Figure was adapted from our joint publication<sup>[115]</sup> under the terms of the Creative Commons License CC BY-NC-ND.

RMSD of residues 26–283 of 2.1 Å to the MD structure versus 3.3 Å to the crystal structure.) This suggests that the E73V mutation and/or the micellar environment stabilize a conformation that the VDAC1 pore can adopt in a phospholipid bilayer according to molecular dynamics simulations. Nevertheless, implications of this finding for possible gating pathways or movements of the channel remain unclear.

The NMR ensemble of E73V hVDAC1 refined against the  $R_2/R_1$  rates was deposited together with the experimental data in the Protein Data Bank (PDB ID 5JDP and 30065).

### 3.3 Incorporating the protein's closed state in NMR – Developing a Quintuple hVDAC1 mutant

#### 3.3.1 General strategy

As shown and discussed above in Chapter 1.3.3, the mode of gating and the nature and structure of the closed state (or its variety of several quite alike closed states) of VDAC1 are a long-standing question in the scientific community. One of the main goals of this thesis was to tackle this question by the means of NMR spectroscopy.

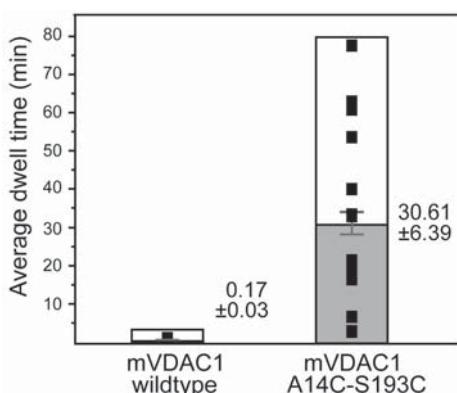
For quite obvious reasons it is not possible to apply a voltage to a liquid state NMR sample like it can be applied to a BLM, because the complexes of the protein and the detergent molecules are tumbling freely in the samples and there are no defined compartments as there are in e.g. BLM measurements. These compartments make it possible to study proteins embedded in a membrane under various parameters such as different ionic strength, pH or medium composition on both sides of the membrane.<sup>[144]</sup> Solid state NMR samples exhibit the same drawback, because also they lack defined compartments.

It is thus necessary to find other approaches to study the closed state of VDAC1 with NMR. Since it is known that the N-terminal  $\alpha$ -helix plays a key role in the gating of the pore<sup>[100]</sup>, many studies focused also on mutations of the helix or other parts of the protein, to gain more insight about the behavior of the helix in the gating process (cf. Chapter 1.3.3). In the course of these studies some groups found VDAC1 mutants that showed unusual or altered gating properties, respectively.

The approach of this thesis is thus, to use hVDAC1 mutants which stabilize the closed state or alter the gating process in a fashion that it can be expected to see the closed state enriched in NMR samples, *i.e.* also at zero voltage. The first of these mutants affixes the N-terminal helix to the  $\beta$ -barrel<sup>[52]</sup> (see Chapter 3.3.2), the second of these mutants features a stiffened linker between  $\alpha$ -helix and  $\beta$ -barrel which significantly changes the gating properties of the mutant<sup>[145]</sup> (see Chapter 3.3.3).

### 3.3.2 Affixing the N-terminal helix to the $\beta$ -barrel

Studies on A14C/C127A/S193C/C232A hVDAC1 were performed based on a publication by Mertins *et al.* from 2012<sup>[52]</sup>, where this mutant, albeit based on mVDAC1, had been described. The two native Cysteine residues in this mutant had been replaced by Alanine. Based on the crystal structure of mVDAC1 (PDB ID: 3EMN)<sup>[46]</sup> two new artificial Cysteines had been added by mutating Alanine-14 and Serine-193. This should allow the formation of a disulfide bond between the N-terminal  $\alpha$ -helix and  $\beta$ -strand 13 of the  $\beta$ -barrel. Indeed Mertins *et al.* could observe the formation of the desired disulfide bond, attesting the 3D structures published in 2008. In the oxidized state, *i.e.* with the disulfide bond formed, this mutant had been described to have special gating properties that seemed to be useful for NMR studies. In general, the mutant has a “usual” open state with conductance of  $3.74 \pm 0.09$  nS (oxidized mutant) and  $3.75 \pm 0.12$  nS (reduced mutant), respectively, and closed states with conductances of  $1.85 \pm 0.07$  nS (oxidized mutant) and  $2.55 \pm 0.12$  nS as well as  $1.67 \pm 0.09$  nS (reduced mutant, all values measured in 1 M KCl, 5 mM CaCl<sub>2</sub>), respectively.



**Figure 3.12: Dwelling times of *wt* mVDAC1 and A14C/C127A/S193C/C232A mVDAC1.** Reprinted from Mertins et al.<sup>[52]</sup> under the terms of Creative Commons Attribution License.

Oxidation and reduction of the mutant had been performed by treatment with CuX<sub>2</sub>Ph and DTT, respectively. The special gating properties of the oxidized mutant are as follows: when a potential of 40 mV is applied, instead of switching into a closed state after seconds like *wt* VDAC1 is doing, the mutant dwells in the open state for over 30 minutes on average. (See Figure 3.12) Switching into the closed state additionally occurred only in 8 % of the traces, that were recorded. The authors of the study hypothesized, that the closure of the cross-linked channel is mediated by mechanical

strain effects due to the covalent bond between  $\alpha$ -helix and  $\beta$ -barrel and that closure is caused by the barrel collapsing, the N-terminal helix irreversibly unfolding or a combination of these two.

The fact that this particular mutant in the oxidized state is failing to reopen once it closed, raised our interest. In principle, also at zero voltage and also when the closed state itself is attained with strongly reduced probability, as the authors conclude<sup>[52]</sup>, there should – based on the argument of Boltzmann distribution – still be a very small but certain amounts of channels, that are in the closed state. That the mutant is not able to reopen, should cause the closed state of the protein to be enriched in the sample over time. This might make it observable at zero voltage in NMR samples.

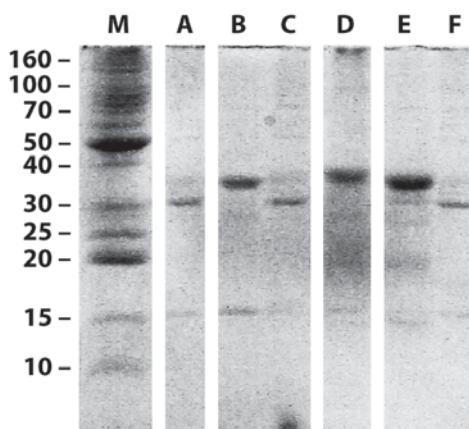
### 3.3.2.1 A14C/C127A/S193C/C232A hVDAC1 samples are oxidized by air

A14C/C127A/S193C/C232A hVDAC1 was expressed, refolded in LDAO and purified following the procedures described in Materials & Methods. To make sure the mutant is in the oxidized state (*i.e.* to make sure the disulfide bond is formed), SDS-PAGE experiments were performed, following a similar procedure as in the publication. As shown in Figure 3.13, hVDAC1 exhibits the expected behavior like described by Mertins *et al.* In all cases that are described in the following, the hVDAC1 samples were taken directly out of the NMR buffer.

First, conditions were chosen such that it should be possible to reach reference states of the mutant (completely oxidized and completely reduced). Line A shows the mutant under the influence of oxidant CuX<sub>2</sub>Ph, line B shows the mutant treated with the reductants DTT and  $\beta$ -mercaptoethanol. The increased electrophoretic mobility in line A indicates the formation of the disulfide cross-link, whereas the disulfide bridge is reduced in line B, being completely in line with the results of Mertins *et al.* and showing that the creation of the desired reference states was successful.

Next, the behavior of the mutant under varying conditions was investigated. Line C shows the mutant without oxidizing or reducing treatment and stored at 37 °C for 6 hours before loading, while line F shows the mutant without treatment and stored at room temperature before loading. In these two cases the mutant behaves like the one that was oxidized by CuX<sub>2</sub>Ph, indicating that air oxygen is sufficient to oxidize the protein and form the disulfide bond independently from the temperature. This points to that the protein in the NMR buffer and consequently also in the NMR

### 3 Results



**Figure 3.13: SDS-PAGE, stained with Coomassie blue.** M: Molecular weight marker with molecular weights marked on the left, A: A14C/C127A/S193C/C232A hVDAC1 treated with CuX<sub>2</sub>Ph, B: A14C/C127A/S193C/C232A hVDAC1 treated with DTT and β-mercaptoethanol, C: A14C/C127A/S193C/C232A hVDAC1 stored at 37 °C, D: V75C hVDAC1, E: C127S/C232S hVDAC1, F: A14C/C127A/S193C/C232A hVDAC1 stored at room temperature.

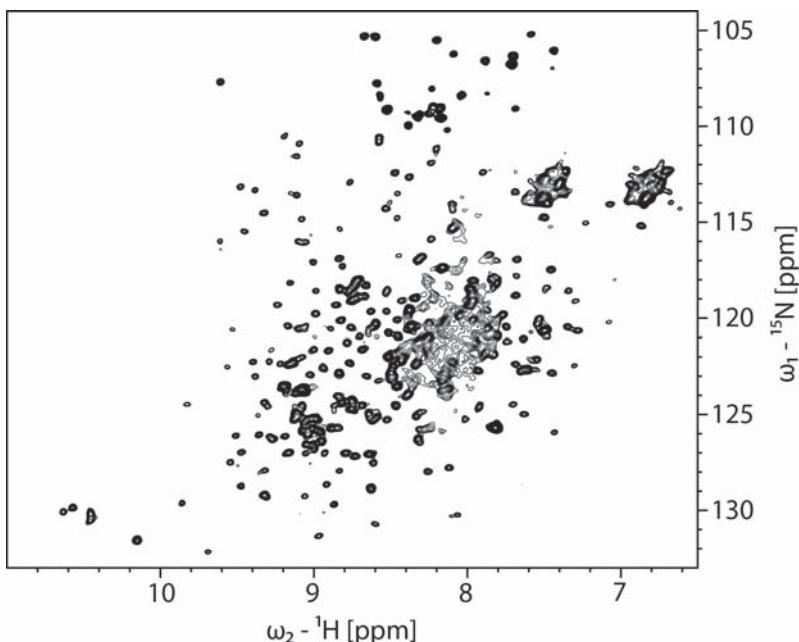
samples is in the state that showed the special gating properties and of which it was hypothesized that it should allow the enriching of the closed state of the protein over time.

To cross-check this hypothesis, line D was loaded with V75C hVDAC1 and line E was loaded with Cysteine-free hVDAC1 (C127S/C232S hVDAC1). In these two cases the protein behaves like in line B – giving further indication that in this case there are indeed no disulfide bridges formed (which is in the case of Cysteine-free hVDAC1 obviously not possible at all). Furthermore, in none of the lines there is a sign of significant oligomerization or intermolecular disulfide bridges formed, as there is no spot with higher molecular weight than approximately 30 kDa.

#### 3.3.2.2 A14C/C127A/S193C/C232A hVDAC1 in liquid state NMR

A14C/C127A/S193C/C232A hVDAC1 delivers well dispersed spectra (see Figure 3.14), that allowed the assignment of 85.6 % of the assignable backbone amide groups. This is only slightly worse than in the *wt* hVDAC1, where at the current state 88.1 % of the backbone amide resonances are assigned (for further information about the current assignment statistics and the improved assignment of hVDAC1 see Chapter 3.1).

The assignment was performed with the help of the already existing assignments of the *wt* hVDAC1 and – which is especially true for the shifted residues – was supported and completely

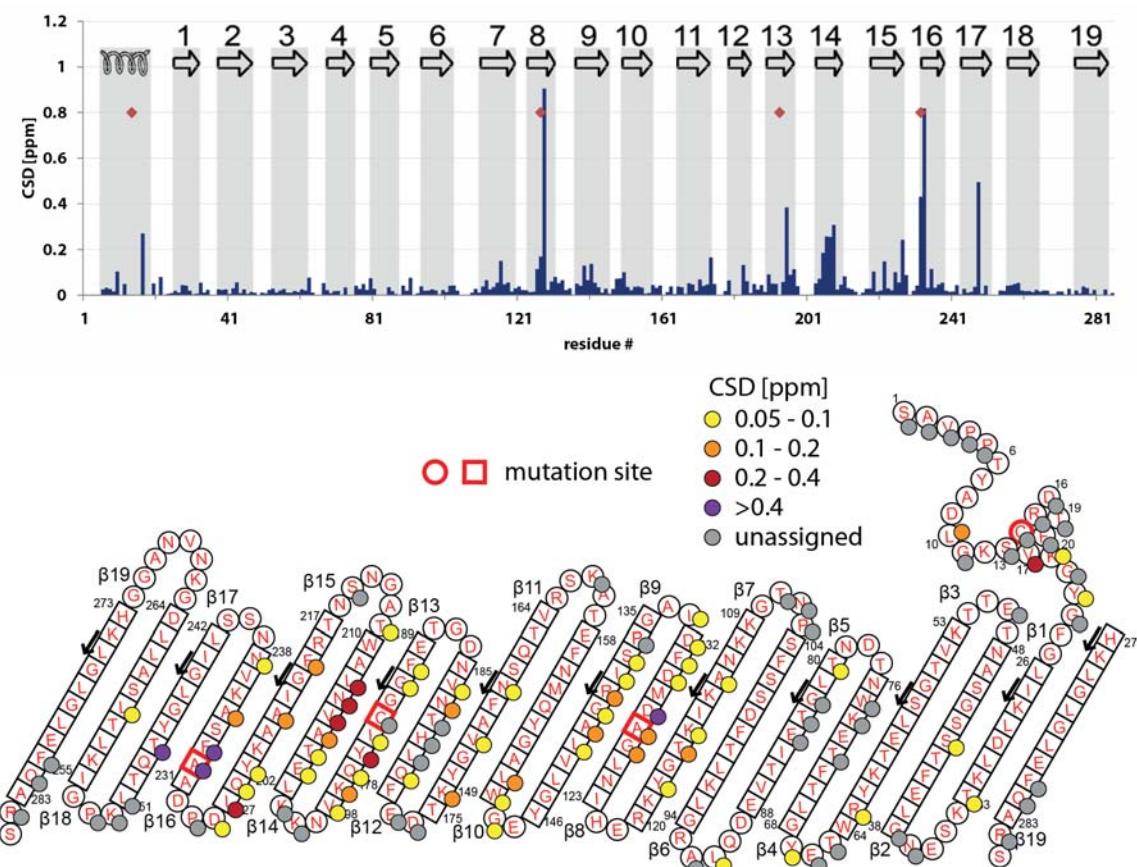


**Figure 3.14:**  $^1\text{H}, ^{15}\text{N}$  TROSY spectrum of A14C/C127A/S193C/C232A hVDAC1.

reviewed via three-dimensional spectra for sequential assignment (TROSY-HNCA and TROSY-HNCO). This way, 81.8 % of the  $\text{C}_\alpha$  resonances and 72.2 % of the C' resonances could be assigned. All assignments are well spread over the whole protein, including residues in the N-terminal helix as well as residues in the  $\beta$  strands and loops.

To probe for structural and conformational differences between *wt* hVDAC1 and A14C/C127A/S193C/C232A hVDAC1, the chemical shift differences (CSD) of combined  $^1\text{H}$  and  $^{15}\text{N}$  chemical shift between the two proteins were determined. The threshold for unshifted residues was set to 0.05 ppm.

As can be seen in Figure 3.15, the largest CSDs are present in spatial proximity around the four mutation sites. For Alanine-14, shifts are visible in the N-terminal helix and the first half of the linker between  $\alpha$ -helix and  $\beta$ -barrel. For the mutations in the  $\beta$ -barrel, the residues with CSD over 0.05 ppm are mostly located in the neighbouring  $\beta$ -strands 7 to 9 (Alanine-127) and  $\beta$  strands 15 to 17 (Alanine-232), respectively. The newly introduced Cysteine-193 causes CSDs of over 0.05 ppm in a slightly bigger range, spanning from  $\beta$ -strand 12 to 14 and including scattered residues in strands 11 and 15, which can be explained by the fact that this residue is attached by the likewise newly introduced disulfide bond to the N-terminal helix and Cysteine-14, respectively. Thus, this mutation

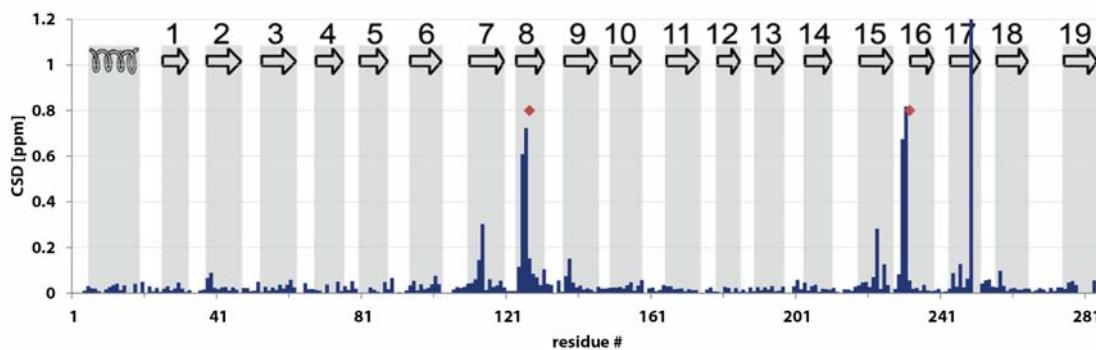


**Figure 3.15: CSD of A14C/C127A/S193C/C232A hVDAC1 in comparison to *wt* hVDAC1.** Chemical shift differences induced by the A14C/C127A/S193C/C232A quadruple mutations in dependence of the residue number (top) and mapped to the topology map of hVDAC1 (bottom). In the upper plot, the mutation sites are marked with red diamonds and the secondary structure of the protein is indicated.

is causing larger conformational disturbances in the direct neighbourhood because the distance between these two residues is roughly, but not perfectly the same as the length of the S-S bond. Based on the X-ray structure of mVDAC1, mutagenesis with PyMol shows that the newly introduced Sulphides are 1.7 Å apart from each other, while a disulfide bridge is approximately 2.05 Å long.

Additionally, two residues in  $\beta$  strand 12 (Leucine-180 and Histidine-181) that were previously assigned in the wild type spectra as well as the mutated Cysteine-193 itself could not be assigned. In general, the residues which show significant CSD in the  $\beta$  strands are shifted by considerable amounts of up to nearly 1 ppm. However, in the barrel and helix regions that are distant to the mutation sites, the CSDs are significantly lower and are nearly completely falling under the threshold of 0.05 ppm.

These findings can be compared with the spectral behaviour of C127S/C232S hVDAC1 in comparison to *wt* hVDAC1. As can be seen in the according plot, the CSD profile looks very similar to the one obtained by comparing A14C/C127A/S193C/C232A hVDAC1 with *wt* hVDAC1. (See Figure 3.16) Only difference are the high CSD values around the two mutation sites Alanine-14 and Serine-193 missing or being there additionally, respectively. It is known that C127S/C232S hVDAC1 is behaving like *wt* hVDAC1 in BLM measurements and is in the open state in liquid state NMR samples.<sup>[146]</sup> Taken together with the fact that the CSD profiles caused by the mutations A14C and S193C do not differ significantly from the ones observed for C127S and C232S in extent and strength, it is concluded that the CSDs visible in A14C/C127A/S193C/C232A hVDAC1 are solely caused by the mutations being present and are no sign for conformational changes in the mutant or the mutant being in the closed state of the protein.



**Figure 3.16: CSD of C127SC232S hVDAC1 in comparison to *wt* hVDAC1.** Chemical shift differences induced by the C127S/C232S mutations in dependence of the residue number. The mutation sites are marked with red diamonds and the secondary structure of the protein is indicated.

### 3.3.3 Stiffening the linker between $\alpha$ -helix and $\beta$ -barrel

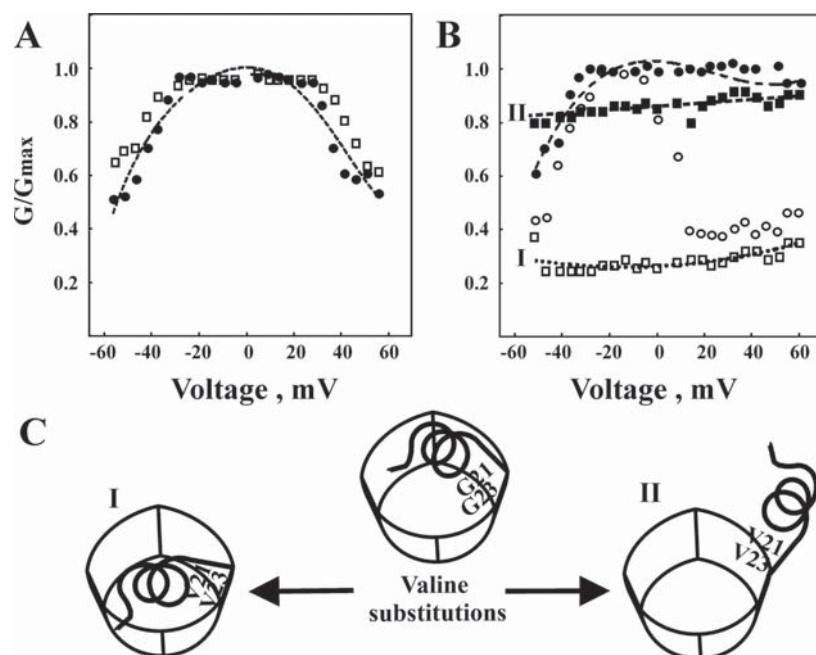
The approach presented here is based on a publication by Geula *et al.* from 2012.<sup>[145]</sup> In that paper the authors describe a G21V/G23V rVDAC1 mutant with interesting properties.

The linker between  $\alpha$ -helix and  $\beta$ -barrel in rVDAC1 consists of a triple-Glycine motif G<sup>21</sup>YGF<sup>25</sup> that is highly conserved among mammals.<sup>[147]</sup> (For example, the respective five residue sequence is identically present also in mVDAC1 and hVDAC1.) As there is a plethora of evidence that the N-terminal helix plays an important role and is moving during the gating process of the channel<sup>[100]</sup>, mutations in the linker might have a crucial influence on the gating. Glycine is the

### 3 Results

only non-chiral proteinogenic amino acid and as such has a larger conformational freedom than all other proteinogenic amino acids, being at the same time able to behave as a quasi L and quasi D conformer.<sup>[148]</sup> Because of this, Glycine rich sequences (GRS) are often found in hinge regions of proteins, allowing flexibility in these regions.

In the described mutant two of the three Glycines in the linker region are replaced by Valine. This obviously makes the linker more stiff and at the same time restricts the conformational freedom of linker and N-terminal helix. As such, an impact of this mutation on the gating properties of the channel can be expected – and this was indeed described by Geula *et al.* (see Figure 3.17).



**Figure 3.17: G21V/G23V rVDAC1 showing various gating profiles.** A: Gating profile of the mutant in which the mutant (empty squares) is behaving like the *wt* rVDAC1 (black circles). B: Representative gating profiles of the same channel of the mutant in which the channel is closed in all voltages (empty squares, type I), open in all voltages (black squares, type II) or gating in totally unusual manner (empty circles, black circles). C: Interpretation of the gating types as provided by Geula *et al.* Type I (fixed in closed state) has the helix fixed inside the pore, type II (fixed in open state) has the helix pointing out of the channel. The figure is reproduced from Geula *et al.*<sup>[145]</sup>

While the bilayer-reconstituted mutant is able to show a gating behavior that is similar to the one of the *wt* rVDAC1 (and such also as hVDAC1 and mVDAC1) in approximately 50 % of the recorded traces, it also often fails to do so. In the other 50 % of the recorded traces the authors saw unusual behavior, reaching from cases where the mutant is locked in the closed state independently from the applied voltage, over cases where the mutant is locked in the open state independently

from the applied voltage, to cases where the mutant is switching between open and closed state at unusual voltages or only in negative or positive potentials. The authors conclude, that these results demonstrate the function of the GRS in conferring flexibility to the N-terminal helix.

There are two interesting points in their findings and theory, respectively. First, they give a number of 50 % for the share of channels that are gating normally (and thus another 50 % share of channels that are gating abnormally). Usually the number of channels where a gating event during a single voltage scan can be observed is significantly lower and not unusually just around 10 %. (E.g. the previously studied mutant described by Mertins *et al.*<sup>[52]</sup> shows gating in 8 % of the recorded traces, according to them.) The mutant thus shows gating behavior of any kind with much higher probability than usual. Second, the gating model they are proposing (see Figure 3.17) is directly contradictory to the nowadays widest accepted models and calculations, where the helix is moving out of the channel during closure and is staying inside of the pore in the open state.<sup>[91]</sup>

Nevertheless, the findings of Geula *et al.* are also very interesting in the light of NMR spectroscopy. The described type-I-gating features pores that are in the closed state when there is no potential applied – exactly the same situation as one has it in the NMR tube. It is thus expectable to see the closed state of the protein at least to some extent when expressing and refolding it to micelles and studying it with the means of NMR spectroscopy.

The following chapters are describing the results with the G21V/G23V hVDAC1 mutant expressed for NMR spectroscopic investigations, the development of a Quintuple hVDAC1 mutant and the results yielded with this construct as well as the development of a model for the closed state based on modifications of Quintuple hVDAC1.

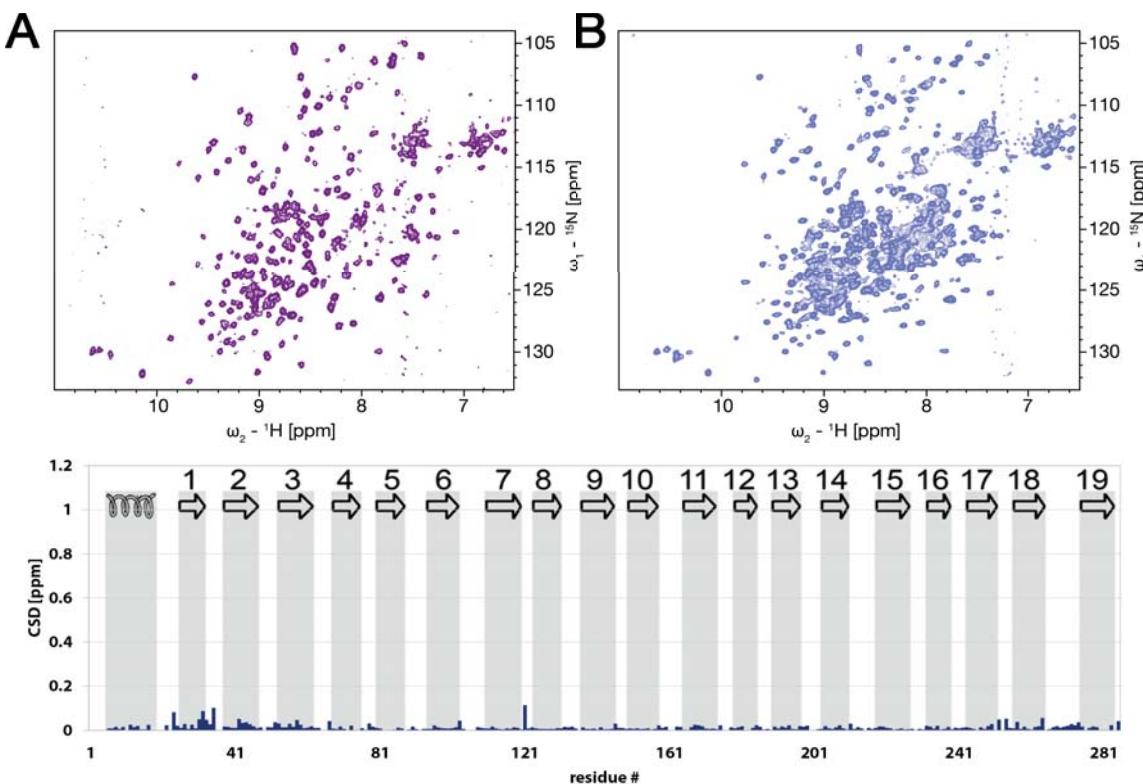
### 3.3.3.1 Changing the expression system to the pET28a expression vector

Because all previous studies in our group were performed on hVDAC1 and the crucial part of the mutant described by Geula *et al.*, namely the GRS in the linker between  $\alpha$ -helix and  $\beta$ -barrel is conserved between rVDAC1 and hVDAC1, it was decided to express a hVDAC1 mutant instead of a rVDAC1 mutant with the same mutations as described – G21V/G23V hVDAC1.

### 3 Results

It became then evident that the G21V/G23V hVDAC1 mutant does not express in the previously used pQE60 vector which allows the expression of hVDAC1 with the correct N-terminus lacking RGS after the starting Methionine. After scanning possible expression vectors it was possible to express the G21V/G23V hVDAC1 mutant with the pET28a expression vector. To validate that hVDAC1 samples expressed in this new system are folded correctly, a *wt* hVDAC1 sample as reference sample was produced by using the pET28a expression vector.

This *wt* hVDAC1 sample shows nicely dispersed  $^{15}\text{N}$ -TROSY spectra that allowed the assignment of basically all for the wild type known residues without the need to record any further experiments just by comparing the  $^{15}\text{N}$ -TROSY spectra of the *wt* hVDAC1 samples expressed in the different expression vectors. (See Figure 3.18)



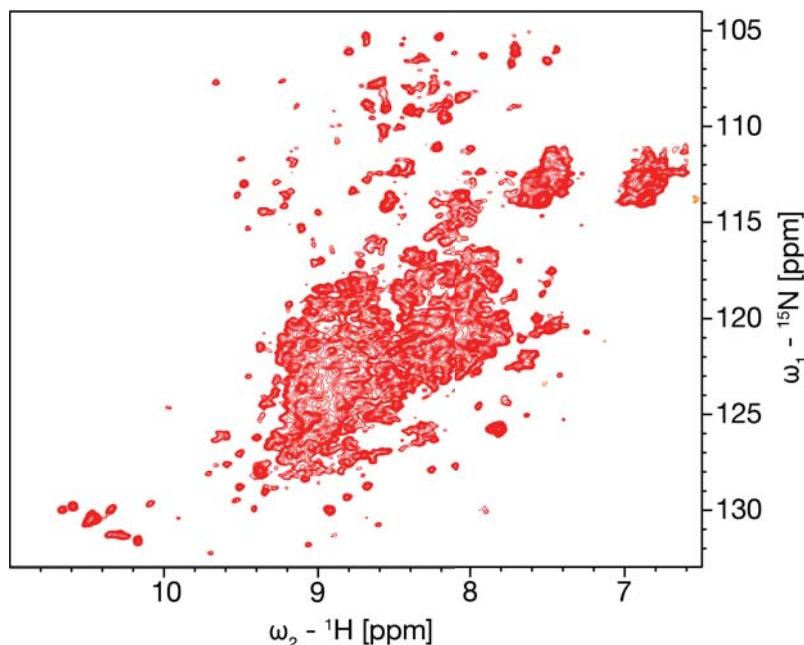
**Figure 3.18: Comparison of *wt* hVDAC1 samples expressed in different expression vectors.** The  $^{15}\text{N}$ -TROSY spectra of *wt* hVDAC1 expressed in the pQE60 vector (A) and the pET28a vector (B) are nearly identical. This is represented in the CSD graph (C).

Although the *wt* hVDAC1 spectrum expressed with the new pET28a vector shows higher amounts of miss-folded protein (regions at proton shift of 8 ppm and 9 ppm with broadened signals), nearly all defined resonances are at identical positions as in the spectrum of *wt* hVDAC1 expressed with the old

pQE60 vector. The only exceptions are a very few residues in loops (e.g. Glutamate-121 being the most prominent one) and residues in the spatial proximity of the C-terminal end, where for technical reasons the sequence does not end with RS but with LE, which could be interpreted as double-mutation that is causing slight resonance shifts. It was thus concluded that the pET28a expression vector is suitable to express hVDAC1 samples and does not have any influence on the folding or properties of the final protein after the expression, opening the possibility to study G21V/G23V hVDAC1 with NMR spectroscopic methods.

### 3.3.3.2 Experiments on G21V/G23V hVDAC1

After G21V/G23V hVDAC1 could finally be expressed, a  $^1\text{H}$ - $^{15}\text{N}$ -TROSY spectrum was recorded to evaluate the sample quality and to start the assignment procedure. Unlike in the most previous cases, the spectral behavior of this mutant turned out to be quite bad. Although the protein was obviously folded correctly, as the signals were dispersed in the usual range (from circa 7.0 to 10.0 ppm in the proton dimension) at 37 °C, the linewidths of the signals were increased significantly. (See Figure 3.19.)

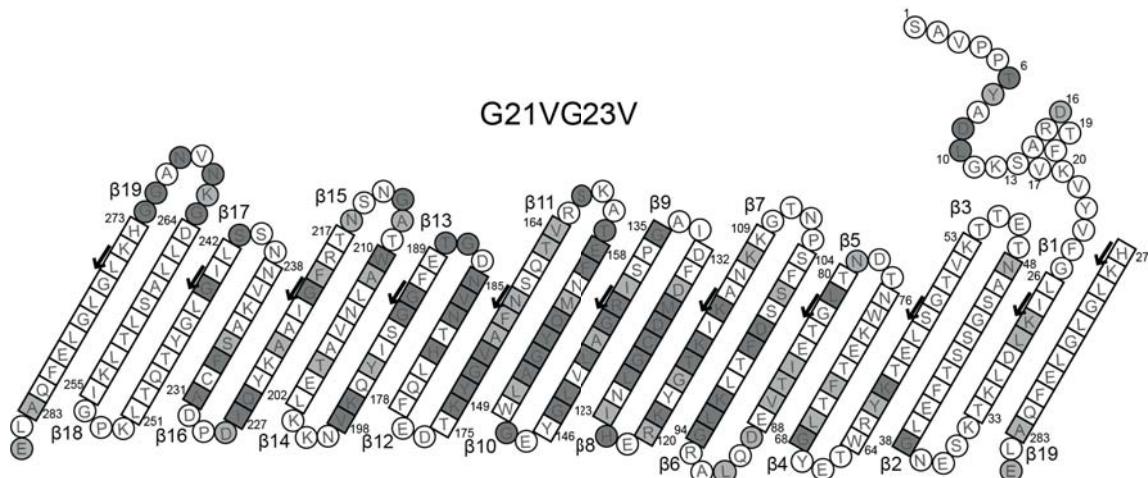


**Figure 3.19:**  $^1\text{H}$ - $^{15}\text{N}$ -TROSY spectrum of G21V/G23V hVDAC1 at 37 °C. The signals are dispersed in the usual range, but are severely broadened, preventing an assignment via 3D spectra.

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Despite the severe broadening of the observed resonances it was tried to perform a tentative assignment of the resonances that could at least be identified as single resonance, even if broadened. This tentative assignment was based on simple comparison of the existing 2D  $^1\text{H}$ - $^{15}\text{N}$ -TROSY spectra, as 3D spectra for sequential assignment were not available for G21V/G23V hVDAC1 and recording them was, given the severe broadening of the lines, scrapped as it would not have any prospect of success.

Based on this approach there were some resonances that could be assigned with relatively high certainty. Either because they are isolated in both the TROSY-spectra of G21V/G23V hVDAC1 and *wt* hVDAC1 (which is true for Threonine-6, Glycine-38 or Glycine-145, for example) or they are isolated in the spectrum of *wt* hVDAC1 and there is a clear maximum in the immediate proximity in the mutant spectrum. Even if it might be located in the region where the broadened resonances are completely merged (which is true for Cysteine-127, Alanine-151 or Tryptophane-210, for example). Also, there are some resonances from the *wt* hVDAC1 spectrum where this kind of assignment is only possible with high uncertainty.



**Figure 3.20: Topology map of G21V/G23V hVDAC1 with possible assignments based on similar resonance positions in comparison to wt hVDAC1.** Residues that have a possible assignment that is relatively clear are colored in dark gray. Resonances where the assignment is very ambiguous are colored in light gray.

All in all, even based on this very non-conservative approach there are just 106 resonances that could be identified as single resonance and are placed at a similar frequency like resonances in the existing *wt* hVDAC1 spectra. The overview of these resonances is shown in Figure 3.20. They comprise 38.1 % of the assignable residues in the whole sequence or 41.7 % of the residues in the

$\beta$ -barrel, respectively. Interestingly, there is a clear prevalence of these possible assignments in the region where in the high-resolution 3D structures of hVDAC1 the N-terminal  $\alpha$ -helix is connected to the  $\beta$ -barrel. This points to that the  $\alpha$ -helix is still stabilizing the barrel in that region to some extent.

To get more insight into why the resonances in the  $^1\text{H}$ - $^{15}\text{N}$ -TROSY spectrum are broadened, several more experiments were performed. The recording of the  $^1\text{H}$ - $^{15}\text{N}$ -TROSY spectra was repeated at a lower temperature than the usually used  $37^\circ\text{C}$  as this could slow down possible motion in the protein to make the broadened lines split up into several lines. As it turns out, reducing the temperature does not help to better resolve the broadened resonances. Instead it quenches the signal yielded from the protein as barely any resonances are retained at  $5^\circ\text{C}$ . (See Figure 3.21)

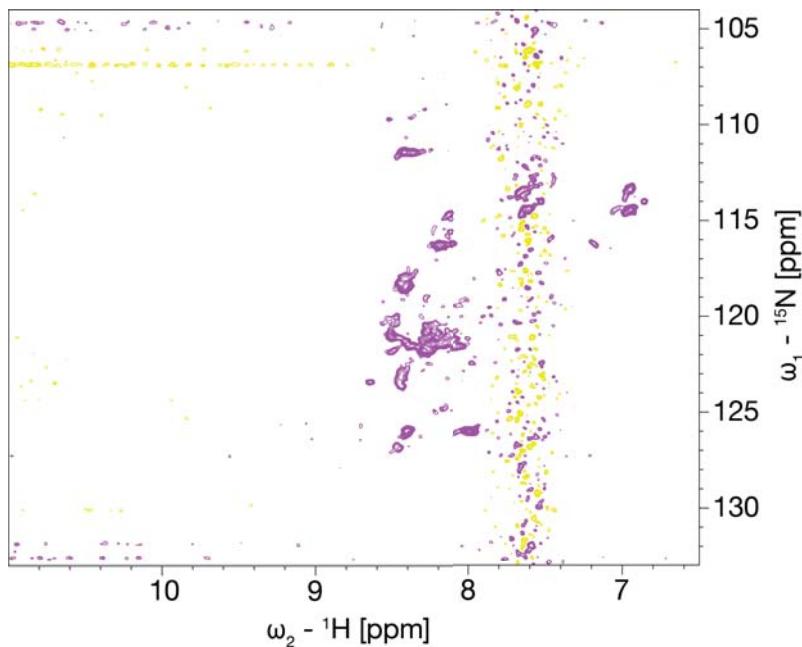


Figure 3.21:  $^1\text{H}$ - $^{15}\text{N}$ -TROSY spectrum of G21V/G23V hVDAC1 at  $5^\circ\text{C}$ .

To also exclude the possibility that the lines in the  $^1\text{H}$ - $^{15}\text{N}$ -TROSY spectrum are broadened because for some reason the G21V/G23V hVDAC1 mutant is prone to oligomerization or dimerization and the emerging oligomers are to large to be detected by transverse relaxation-optimized spectroscopy (as it was observed for T19C/C127S/C232S hVDAC1 by Saskia Villinger)<sup>[74]</sup>,  $^1\text{H}$  1D (1-1) echo experiments were performed to estimate the overall rotational correlation time of the protein.<sup>[114]</sup> Bigger entities than the usual combination of one hVDAC1 channel and the surrounding

### 3 Results

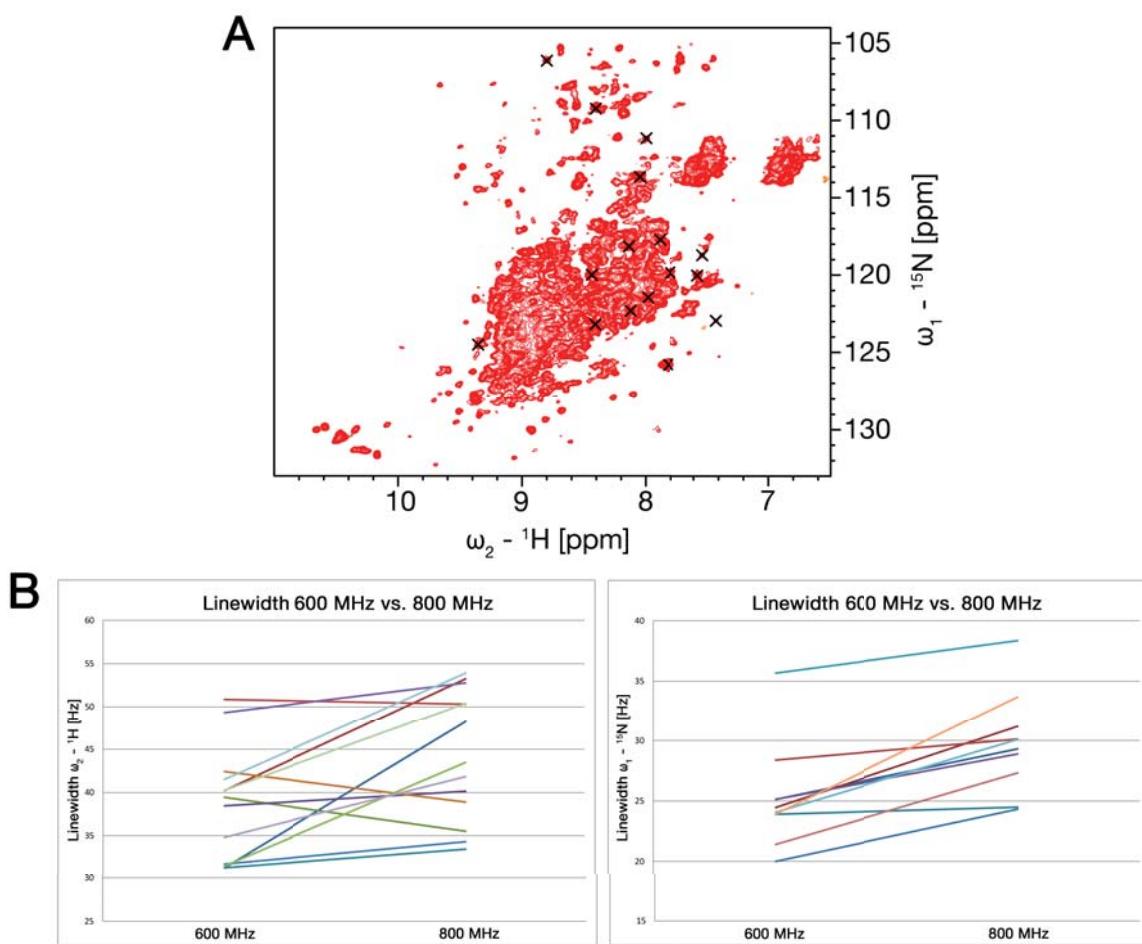
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LDAO micelle, weighing usually roughly 140 kDa, should have a significantly elevated rotational correlation time. However, this could be ruled out. Based on the measurements, for G21V/G21V hVDAC1 rotational correlation times of 26 ns in the folded and 37 ns in the unfolded region ( $^1\text{H}$  shift of 9.25 ppm and 7.5 ppm as representative positions, respectively) were estimated, which fits well to previously reported values for the *wt* hVDAC1 of 23 ns in the folded and 35 ns in the unfolded region, respectively. (For the dimer-forming T19C/C127S/C232S hVDAC1 values of 50–60 ns were reported.)

Finally, the behavior of the resonances that could be found in the  $^1\text{H}$ - $^{15}\text{N}$ -TROSY spectrum was examined in regard of the linewidth these resonances are showing in dependence of the magnetic field used for measurement. Solely based on the TROSY effect one would expect that the lines should become sharper due to the improved compensation of the transversal relaxation time  $T_2$  when switching from 600 MHz to 800 MHz measurement frequency <sup>[149,150]</sup>. However, as can be seen in figure 3.22, the opposite is the case.

For this analysis, not only completely isolated resonances were taken into account, but also some resonances in the region where the single resonances are due to shifts and line broadening to a large extent not identifiable anymore. The linewidths were determined by CcpNmr Analysis. All resonances had thus to be strong enough in both analyzed spectra that their linewidth could be calculated by the program without distortions due to neighboring resonances. The analyzed resonances were refined and cases where shoulders or neighboring resonances distorted the result, are excluded. As Figure 3.22 B shows, there are some few resonances that don't broaden with increasing field strength, but the overall tendency is clearly visible: higher field strength is broadening the resonances.

All in all it was concluded that the resonances in G21V/G23V hVDAC1 are broadened due to chemical exchange that is intermediate on the NMR timescale, which does not allow a deeper analysis of the spectra yielded. This could indeed be seen in line with the results of Geula *et al.*, as they showed that the mutant is switching between open and closed state to a large part totally independent of the applied potential. If this is happening in a speed which is intermediate on the NMR timescale, this would totally explain the results that we got. Also the fact that certain resonances don't broaden



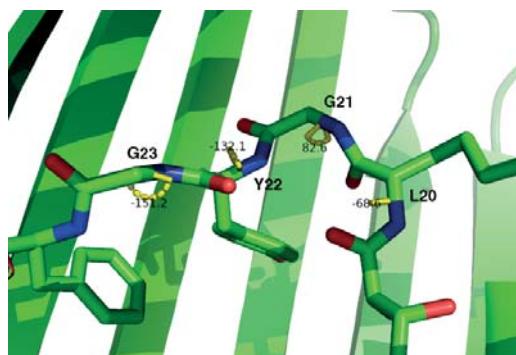
**Figure 3.22: Linewidths of several residues in G21V/G23V hVDAC1.** A: Resonances that were chosen for analysis are marked with “x”. B: Linewidths of the respective resonances at 600 and 800 MHz in proton (left) and nitrogen dimension (right). Although there are some outliers the overall trend is that the linewidth of every resonance is increasing with increasing field strength.

with increasing field could just mean that they are less affected by the motion that underlies the broadening effect.

Nevertheless, to check the results described by Geula *et al.*, in collaboration with Roland Benz (Department of Life Sciences and Chemistry, Jacobs University Bremen gGmbH) we performed own BLM measurements on G21V/G23V hVDAC1. Due to lack of material, the voltage dependency of the protein could not be investigated. Still it could be shown that there are some pores with the usual conductance of 3.9 nS in 1 M KCl and a lot of pores with lower conductance – being in line with the results published. (Data not shown.)

### 3.3.3.3 Experiments on G21V hVDAC1

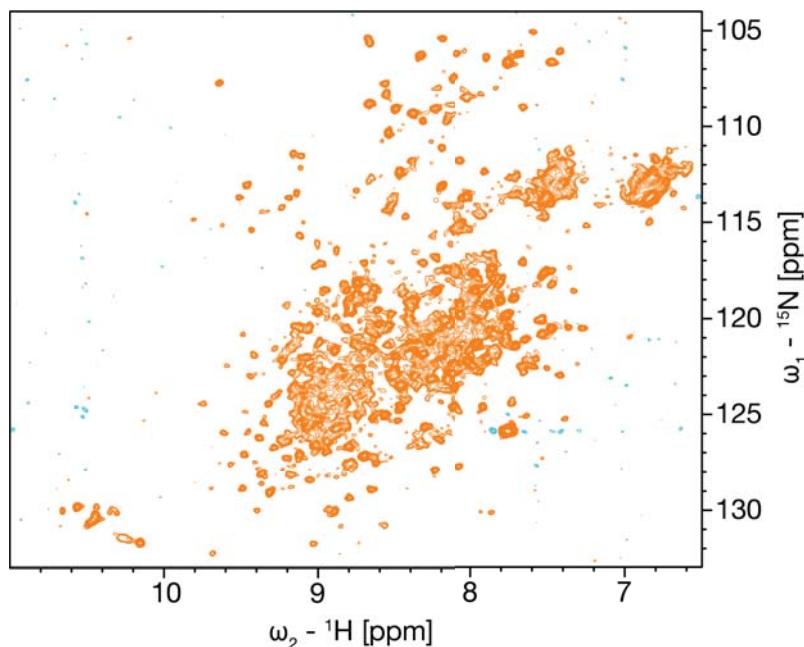
After the result that spectra of G21V/G23V hVDAC1 are broadened due to intermediate exchange in NMR and are thus not usable for deeper analysis as described in the last chapter, the next step was to have a look into which of the two mutated Glycines or introduced Valines, respectively, might be more important for the properties Geula *et al.* described for the mutant. When checking the conformation of the two Glycines in the crystal structure of mVDAC1, it turns out that Glycine-23 adopts a conformation that also L amino acids can adopt, while Glycine-21 has a conformation that, when replaced by Valine, only D-Valine could adopt. ( $\phi(G21) = 82.6^\circ$ ,  $\phi(G23) = -151.2^\circ$ , See Figure 3.23) Therefore we hypothesized that the mutation G21V could have bigger responsibility for the mutant properties and at the same time by only replacing one instead of two Glycines in the linker, the stiffening effect could be weakened such that a timescale of movement is reached that makes it possible to study the mutant by NMR spectroscopy again.



**Figure 3.23: Glycine-21 and Glycine-23 in the 3D structure of mVDAC1.** While Glycine-23 adopts a L like configuration ( $\phi(G23) = -151.2^\circ$ ), Glycine-21 could only be replaced by D-Valine without changing the backbone angles ( $\phi(G21) = 82.6^\circ$ ).

G21V hVDAC1 could be expressed in the pET28a expression vector and was studied with  $^1\text{H}$ - $^{15}\text{N}$ -TROSY experiments. As it turns out, similar like it was the case with G21V/G23V hVDAC1, also this mutant delivers spectra that show severe resonance broadening, although this broadening effect is far less strong than with G21V/G23V hVDAC1 and the  $^1\text{H}$ - $^{15}\text{N}$ -TROSY spectrum is overall of significantly better quality. (See Figure 3.24)

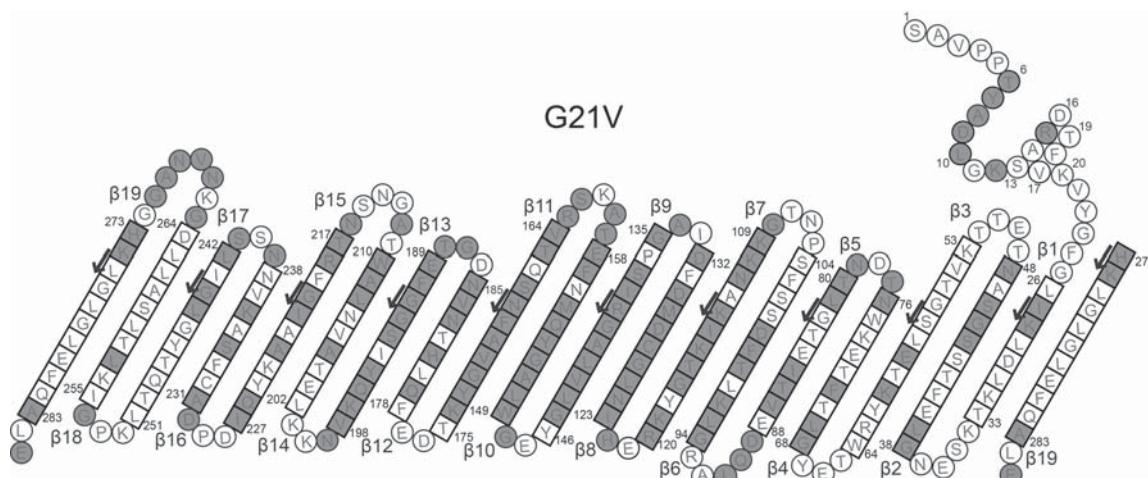
As described for G21V/G23V hVDAC1 a tentative assignment by comparing the available 2D  $^1\text{H}$ - $^{15}\text{N}$ -TROSY spectra was also performed for G21V hVDAC1. The number of residues for which



**Figure 3.24:**  $^1\text{H}$ - $^{15}\text{N}$ -TROSY spectrum of G21V hVDAC1 at 37 °C. The resonances are dispersed in the right range for folded proteins, but are broadened due to intermediate exchange.

for which assignments can be made by matching the amide signals with those from *wt* hVDAC1 is increased in G21V hVDAC1 to 144 residues, which corresponds to 51.8 % of the whole protein sequence or 56.3 % of the  $\beta$ -strands. Again, in the region where in the high resolution 3D structures of mVDAC1 and hVDAC1 the  $\alpha$ -helix is anchored to the barrel, *i.e.* in the  $\beta$ -strands 7 to 13, the number of these possible assignments is significantly increased. Still, the number of possible assignments based on this approach is far from what can be assigned in *wt* hVDAC1 or E73V hVDAC1. (See Figure 3.25)

The  $^1\text{H}$ - $^{15}\text{N}$ -TROSY spectra of G21V hVDAC1 are of better quality than these of G21V/G23V hVDAC1, but still show signs of broadening due to intermediate exchange (which was understood as movement causing the special gating properties described by Geula *et al.*). Based on this fact it was hypothesized that it might – contrary to G21V/G23V hVDAC1 – be possible to reach a better assignment. To do this and to confirm the assignments that are until now only based on similar resonance positions a 3D TROSY-HNCA spectrum was recorded. After recording this spectrum it turned out that this assumption was wrong, as the recorded TROSY-HNCA does not show any signals above the noise level. Additionally, after recording the TROSY-HNCA, the sample presented itself com-



**Figure 3.25: Topology map of G21V hVDAC1 with possible assignments based on similar resonance positions in comparison to wt hVDAC1.** Residues that have a possible assignment are colored in dark gray.

pletely unfolded in the subsequently recorded  $^1\text{H}$ - $^{15}\text{N}$ -TROSY, pointing to that the G21V mutation is decreasing the sample's stability at the measurement temperature of 37 °C dramatically.

Also the G21V hVDAC1 mutant has been studied with BLM measurements in collaboration with Roland Benz, Jacobs-University Bremen. In these measurements, only the normal gating channels were taken into account. G21V hVDAC1 has an open channel conductance of  $3.8 \pm 0.16$  nS in 1 M KCl, which is slightly lower than for *wt* hVDAC1. Furthermore, it shows significantly more distinct gating, meaning that it is gating at way lower potentials than usual: the voltage dependency starts already at around 20 mV and is completely shaped at potentials of around 50 mV. Interestingly, for this mutant one could not observe channels that are already closed at zero potential.

### 3.3.3.4 Development of a Quintuple hVDAC1 mutant

After the results with G21V/G23V hVDAC1 and G21V hVDAC1, the available information was reinterpreted. From the  $^1\text{H}$ - $^{15}\text{N}$ -TROSY spectra of these two mutants it was obvious that the barrel is too much destabilized such that intermediate exchange is causing severe resonance broadening in these spectra. On the other hand, it is known since several years, that the E73V mutation is stabilizing the barrel and improving the spectra quality dramatically. This had theoretically been proven by Villinger *et al.*<sup>[132]</sup> and can practically be seen in that this spectra quality lead to the

dramatically improved NOE network, the possibility to record a TROSY-HNCACB spectrum and the new high resolution 3D structure of E73V hVDAC1 described in this thesis.

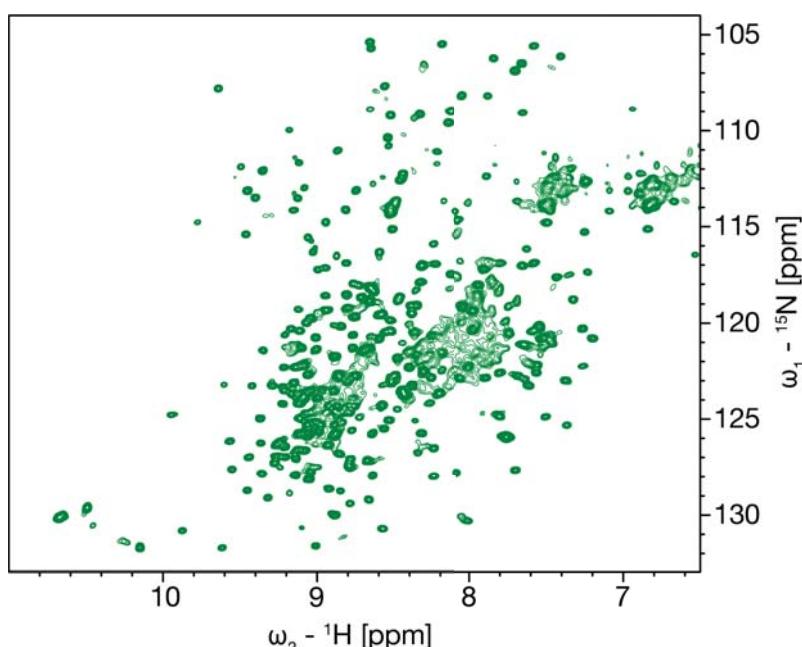
Furthermore, the BLM measurements performed by Roland Benz showed, that it is necessary to have both the G21V and the G23V mutation present in the sample to be able to observe channels that are in the closed state at low or zero potentials applied to the membrane, as this could only be observed for G21V/G23V hVDAC1 but not for G21V hVDAC1.

Additionally, the fact that G21V hVDAC1 presented itself completely unfolded after the measurement of the TROSY-HNCA, calls for stabilization of the mutant not only dynamically but also thermodynamically such that the stability over time at 37 °C has to be improved. For this, additionally to the E73V mutation the two native Cysteines were removed, which is taking the system the possibility to denaturize or dimerize, respectively, by forming disulfide bonds. As mentioned earlier the removal of the Cysteines has already been shown to have no influence on the gating properties of the channel.<sup>[146]</sup> Given the fact that Cysteine-127 is pointing to the micelle, Cysteine-232 is pointing to the water inside the pore and with the aim to replace the Cysteines with the most similar other amino acids possible it was decided to replace Cysteine-127 by Alanine (which is apolar and thus disturbing the micelle less) and Cysteine-232 by Serine (which is polar and thus better fitting to the water filled interior of the channel).

All in all this leads to a Quintuple hVDAC1 construct, namely G21V/G23V/E73V/C127A/C232S hVDAC1, which was designed to combine the desired special gating properties with the necessary stability for NMR measurements.

### 3.4 Experiments on Quintuple hVDAC1 and development of a model of the closed state of hVDAC1

Quintuple hVDAC1 was expressed in the pET28a expression vector as it was done with G21V/G23V hVDAC1 and G21V hVDAC1. Unlike these two mutants, Quintuple hVDAC1 indeed delivers  $^1\text{H}$ - $^{15}\text{N}$ -TROSY spectra in which the resonances are not broadened due to intermediate exchange. (See Figure 3.26)



**Figure 3.26:**  $^1\text{H}$ - $^{15}\text{N}$ -TROSY spectrum of Quintuple hVDAC1 at 37 °C. The dispersion of the resonances is indicating a correct fold of the protein. Different from G21V and G21V/G23V hVDAC1, the resonances are not broadened in a way which renders interpretation of the spectra impossible.

Based on the good spectral behavior of Quintuple hVDAC1, assignment of the protein was possible and because of the importance of this mutant a lot of effort has been put into it. To assign Quintuple hVDAC1,  $^1\text{H}$ - $^{15}\text{N}$ -TROSY spectra and already existing assignments for *wt* hVDAC1 were used first. To support the assignment an E73V/C127A/C23S hVDAC1 mutant was also expressed in the pET28a vector as a wild-type-like reference sample (as in comparison Quintuple hVDAC1 and E73V/C127A/C23S hVDAC1 just differ by the G21V/G23V mutation). This is the mutant where as described in Chapter 3.1.1 recording a TROSY-HNCACB spectrum was successful for the first time. This way, a good reference spectrum and reference assignment was available that could then

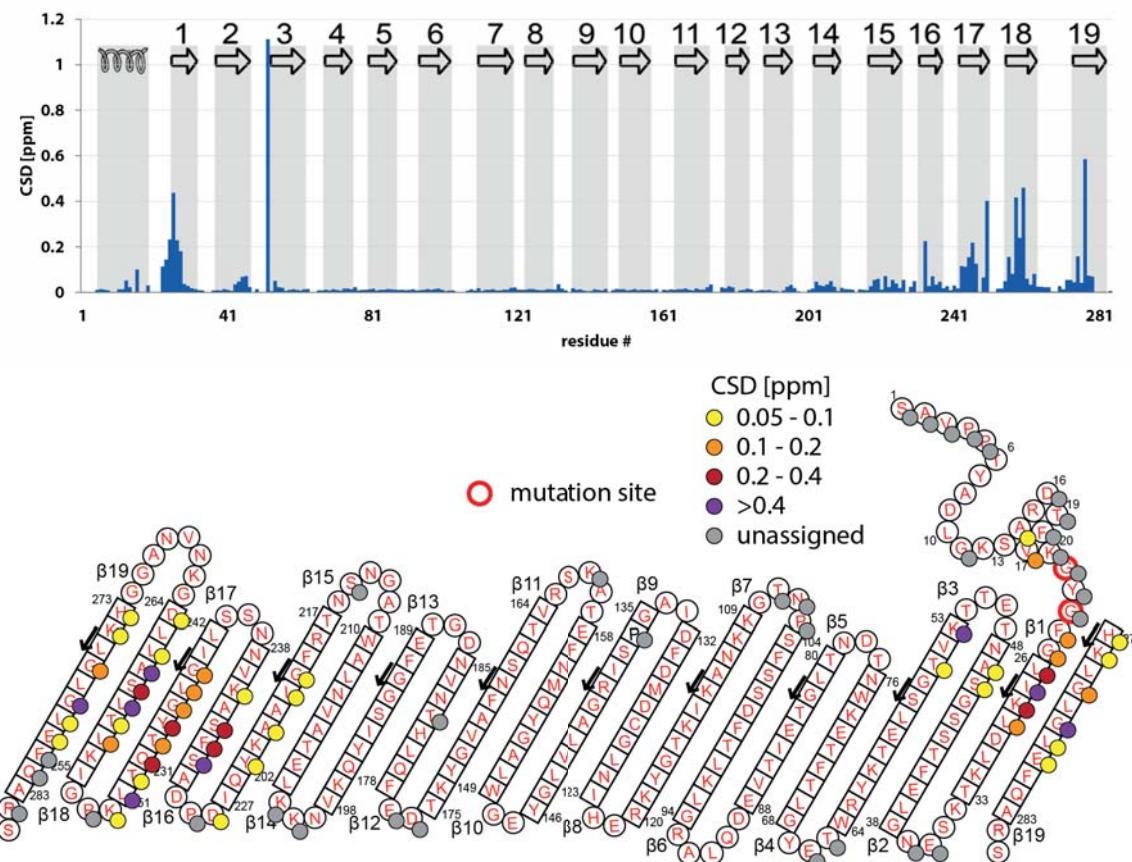
be used to assign Quintuple hVDAC1. Additionally, the assignment was performed with the help of further recorded spectra: a mixed-time parallel evolution HMQC-NOESY as described and previously recorded for other hVDAC1 mutants by Saskia Villinger<sup>[74,111]</sup> to yield spatial connections between the residues, a TROSY-HNCA to yield the sequential connections and – after it turned out with E73V/C127A/C232S hVDAC1, that recording a TROSY-HNCACB spectrum is possible – also a TROSY-HNCACB spectrum.

With the help of these spectra, it was possible to nearly reach a level of assigned backbone resonances similar to E73V/C127A/C232S hVDAC1. Presently, the assignment statistics are as follows: There are 254 assigned amide backbone groups in the overall protein, corresponding to 91.4 % of the assignable amide groups (for comparison 88.1 % are assigned in *wt* hVDAC1 and 92.1 % are assigned in E73VC127AC232S hVDAC1). In the  $\beta$ -strands, 189 amide backbone groups are assigned, which is corresponding to 98.4 % of the assignable amide groups (*wt* hVDAC1: 94.8 %, E73V/C127A/C232S hVDAC1: 98.4 %). Thus, the backbone amide assignment in Quintuple hVDAC1 is of even better quality than it can be reached with *wt* hVDAC1 and only slightly worse than in the reference E73V/C127A/C232S hVDAC1. With the TROSY-HNCA and the TROSY-HNCACB also the  $C_{\alpha}$  and  $C_{\beta}$  resonances could be assigned. However, in the triple-resonance spectra the differences in overall sample quality, sample stability and signal strength are, due to the generally decreased sensitivity in these kind of spectra and the required measurement time, more pronounced, such that here the level of assignment is significantly lower for Quintuple hVDAC1. Presently, 90.2 % of the  $C_{\alpha}$  and 54.1 % of the  $C_{\beta}$  atoms are assigned, which is less than for E73V hVDAC1 or E73V/C127A/C232S hVDAC1 ( $C_{\alpha}$ : 95.4 %,  $C_{\beta}$ : 87.4 %), respectively. Importantly, there is generally only one set of resonances that could be observed in the Quintuple hVDAC1 spectra.

To now assess differences between Quintuple hVDAC1 and E73V/C127A/C232S hVDAC (which will from now on be used as a wild-type-like reference mutant as the only difference between these two mutants is the G21V/G23V mutation that is inducing the protein to be partly closed), the chemical shift differences (CSD) and intensity changes between the two mutants were investigated. As can be seen in Figure 3.27, only small CSDs can be observed in the  $\beta$ -strands 4 to 13 of the barrel. The largest CSDs are located in the  $\beta$ -strands 14 to 19 and  $\beta$ -strand 1 as well as in the proximity

### 3 Results

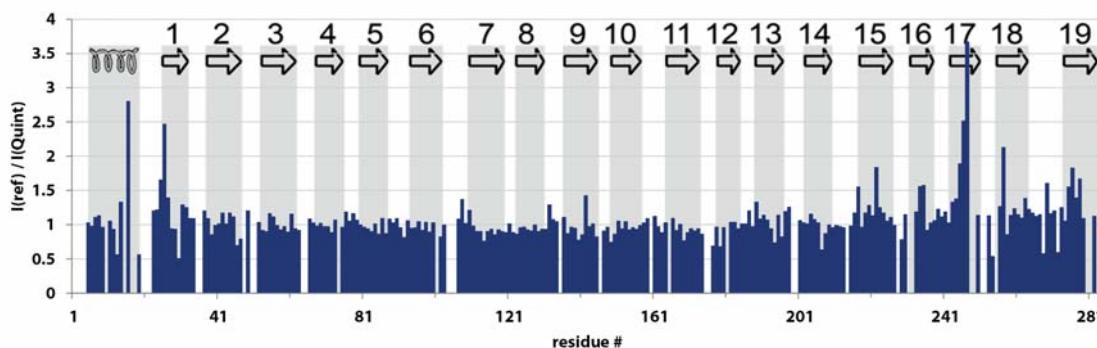
of the loop from  $\beta$ -strand 2 to 3, indicating conformational changes in these regions. The  $\alpha$ -helix nevertheless seems to be only affected in the C-terminal end, which anyway has to be taken with care, as due to the missing sequential links in the helix assignment of a possible second conformation is difficult to impossible. However, this is strongly suggesting that the N-terminal helix is not unfolding upon the changes that can be observed in this mutant.



**Figure 3.27: CSD of Quintuple hVDAC1 in comparison to E73V/C127A/C232S hVDAC1.** Chemical shift differences induced by the G21V/G23V mutations in dependence of the residue number (top) and mapped to the topology map of hVDAC1 (bottom).

Still, the CSDs observable in the last  $\beta$ -strands point to conformational changes there. Additionally, the change of intensity values to asses possible changes in dynamics were investigated. The respective graph can be seen in Figure 3.28. Obviously, some residues in the  $\beta$ -strands 15 to 19 are severely broadened in the Quintuple hVDAC1 mutant, pointing to increased dynamics in these strands. The same is true for residues in the linker and in  $\beta$ -strand 1.

Taken together, by analyzing the  $^1\text{H}$ - $^{15}\text{N}$ -TROSY spectra, first experimental evidence is pointing to some conformational and dynamical changes in the C-terminal  $\beta$ -strands of the  $\beta$ -barrel as well



**Figure 3.28: Intensity of the resonances of Quintuple hVDAC1 in comparison to E73V/C127A/C232S hVDAC1.**

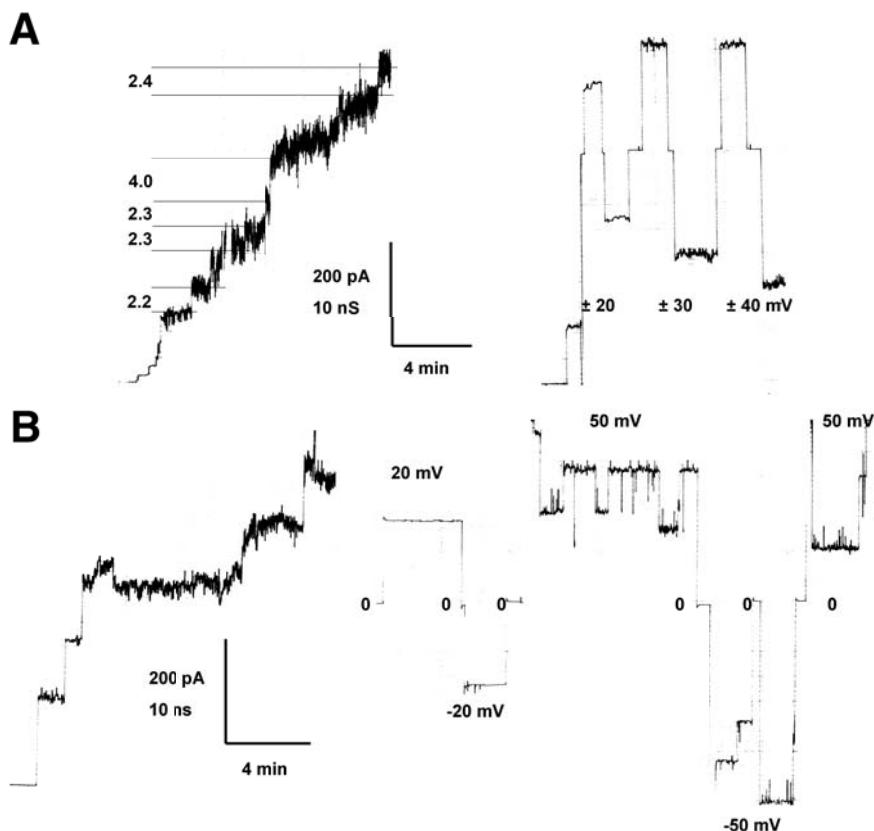
The intensity ratio of the resonances of the two mutants E73V/C127A/C232S hVDAC1 and Quintuple hVDAC1 is plotted against the residue number. Ratios greater than 1 indicate weaker resonances in the Quintuple hVDAC1.

as the linker between barrel and helix and the proximity of the point where to linker is connected to the barrel.

### 3.4.1 Quintuple hVDAC1 behaves like a $\Delta N$ construct in BLM measurements

As the experiments on Quintuple hVDAC1 are following the assumption that this mutant is exhibiting the same or very alike gating properties like the G21V/G23V hVDAC1 mutant described by Geula *et al.*, BLM measurements had to be performed with the mutant to confirm this. The BLM measurements were performed by Roland Benz (Department of Life Sciences and Chemistry, Jacobs University Bremen gGmbH), with a part of the NMR sample containing Quintuple hVDAC1 reconstituted in LDAO micelles.

As depicted in Figure 3.29, besides pores with the usual conductance of around 4 nS, Quintuple hVDAC1 shows a lot of pores with lower conductance of slightly above 2 nS at small potentials that are usually not sufficient to induce closing of the channel (10 to 20 mV). While the former channels can close at higher potentials of  $\pm 50$  mV, the latter type of channels does never show closure. The channels with conductances of around 4 nS are obviously Quintuple hVDAC1 channels in the open state and thus can close when a higher potential is applied. On the other hand the channels with conductances of slightly above 2 nS resemble channels that are already closed at lower potentials and thus they are not showing gating when the applied potentials are increased. Notably, the channels with closed state-like conductance are showing relatively noisy traces. Overall, the BLM

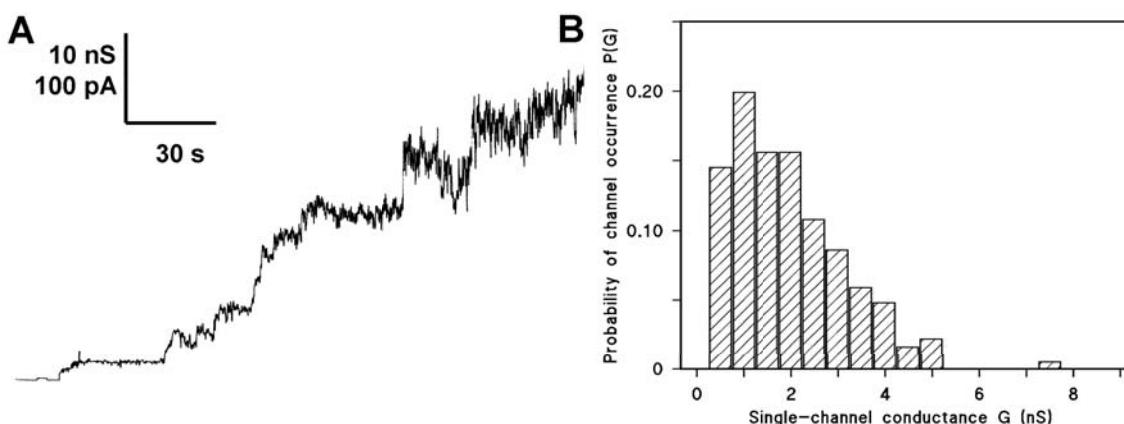


**Figure 3.29: BLM measurements on Quintuple hVDAC1.** A: There are a lot of channels with closed state-like conductances that do not show gating at higher potentials. B: There are also channels with usual open state-like conductances that show gating at higher potentials.

traces are showing that Quintuple hVDAC1 has – compared to *wt* hVDAC1 – significantly altered gating properties with the protein being in the closed state at unusual voltages. This is confirming that Quintuple hVDAC1 shows the desired properties that are similar to G21V/G23V rVDAC1 like it was described by Geula *et al.*

Similar traces as Quintuple hVDAC1 (Figure 3.29 A) have already been described for a completely different VDAC mutant. In this group, a  $\Delta(1-31)$ -hVDAC2 construct has been studied.<sup>[151]</sup> hVDAC2 is eleven residues longer than hVDAC1, having the extension at the N-terminal end (see sequence alignment in Appendix C). The sequence identity of this construct with the corresponding part of hVDAC1 is 74.9 % and the percentage of strongly similar amino acids is 91.6 % according to Clustal Omega<sup>[152]</sup>, such that the  $\Delta(1-31)$ -hVDAC2 construct is basically the same as a  $\Delta(1-20)$ -hVDAC1 construct – being a hVDAC variant where the N-terminal helix is deleted. Such a construct has

been shown by computational electrophysiology simulations to have a Cation-selective state with a conductance of around 2 nS at an ellipticity of approximately 0.5.<sup>[91]</sup>



**Figure 3.30: BLM measurements on  $\Delta(1\text{--}31)\text{-hVDAC2}$ .** Illustration is reprinted from Gattin *et al.*<sup>[151]</sup> with permission from SpringerLink.

For the  $\Delta(1\text{--}31)\text{-hVDAC2}$  construct, roughly 2 nS are the most often observed conductance value in BLM measurements. However,  $\Delta(1\text{--}20)\text{-hVDAC1}$  and  $\Delta(1\text{--}31)\text{-hVDAC2}$  could not be studied by liquid state NMR spectroscopy due to too big instability in the barrel caused by the missing N-terminal helix. Taking this together, Quintuple hVDAC1 partly behaves like a  $\Delta N$  construct in BLM measurements, although it still possesses the N-terminal helix and is still being able to be studied by liquid state NMR. The  $\Delta N$  construct as well as Quintuple hVDAC1 show conductances of around 2 nS and by computational methods it was shown that this state most likely is Cation-selective – exactly like the closed state of the protein.

All this is pointing towards that the N-terminal helix in Quintuple hVDAC1 moves away from its original position, causing  $\Delta N$  construct-like BLM traces, and this movement resembles the closed state of the protein. Notably, this seems to be contradictory to the fact that no elevated CSD values are visible in the open-state contact region between  $\alpha$ -helix and  $\beta$ -barrel in liquid state NMR spectra. This, however, could also be caused by the different environments of the protein in the two measurement methods (micelles in liquid state NMR and phospholipid membranes in BLM measurements).

### 3 Results

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#### 3.4.2 Quintuple hVDAC1 in solid state NMR measurements

With the knowledge that Quintuple hVDAC1 behaves like a  $\Delta(1-31)$ -hVDAC2 construct in BLM measurements and given the fact, that the similar  $\Delta(1-20)$ -hVDAC1 construct could be studied with solid state NMR but not with liquid state NMR, the decision was taken to also produce a Quintuple hVDAC1 sample that was reconstituted into DPhPC liposomes for solid state NMR measurements.

As described by Schneider *et al.*<sup>[113]</sup>,  $\Delta(1-20)$ -hVDAC1 and *wt* hVDAC1 show several differences in the solid state spectra due to the N-terminal helix missing in the former construct. In particular, in a  $^{13}\text{C}$ - $^{13}\text{C}$  PDSD spectrum with a mixing time of 300 ms, the Leu-142-Val-143-Leu-144 motif could be assigned together with some sequential, long-range and internal contacts of these three residues for *wt* hVDAC1. Valine-143 forms a hydrophobic patch together with Leucine-150 that points to the interior of the barrel and is anchor point for Leucine-10 of the N-terminal helix. In the  $\Delta(1-20)$ -hVDAC1 construct however, nearly all the sequential, internal and long-range contacts for these three residues are missing.

A similar 300 ms  $^{13}\text{C}$ - $^{13}\text{C}$  PDSD spectrum was recorded for Quintuple hVDAC1 by ShengQi Xiang. The overall quality of the resulting spectrum was good, such that the resonances in question were examined. The results are shown in Table 3.1.

**Table 3.1:** Assignability of PDSD contacts

resonance	Assignable in		
	<i>wt</i> hVDAC1	$\Delta(1-20)$ -hVDAC1	Quint-hVDAC1
L142CA-CB	yes	no	unclear
V143CA-L10CG	yes	no	no
V143CA-L144CG	yes	no	yes
V143CB-L10CG	yes	no	no
V143CB-CG1	yes	no	yes
V143CB-CG2	yes	yes	yes
V143CB-L144CG	yes	no	yes
L144CA-CB	yes	no	no

Obviously, Quintuple hVDAC1 does not behave completely identical to  $\Delta(1-20)$ -hVDAC1, but quite similar. While the internal and sequential contacts involving  $C_\gamma$  atoms are retained, all other contacts are not visible anymore. (The  $C_\alpha$ - $C_\beta$  contact of Leucine-142 is unclear, see below.) That means that in particular the contacts from the N-terminal helix to the  $\beta$ -barrel are missing, indicating once again, that the N-terminal  $\alpha$ -helix has moved from its original position at least partly. As with the BLM measurements, this seems to contradict the fact that no elevated CSD values are visible in the open-state contact region between  $\alpha$ -helix and  $\beta$ -barrel in liquid state NMR spectra. However, also in this case the two environments in question into which hVDAC1 is reconstituted are very different. (Micelles in liquid state NMR and liposomes in solid state NMR.) This points to that the environment of the protein plays a large role in to which extent the special properties of Quintuple hVDAC1 are visible. More membrane-like environments seem to result in more strongly altered protein properties.

Yet, the impact of the movement of the helix is not as strong as its complete removal, as not all contacts in the region of the hydrophobic patch are broadened beyond detection as it happens in  $\Delta(1-20)$ -hVDAC1, but just some.

As it was possible to measure an HNCACB on Quintuple hVDAC1 in liquid state, both measurement methods can be compared regarding the  $C_\alpha$  and  $C_\beta$  resonances. Indeed, for both Leucine-142 and Leucine-144,  $C_\alpha$  and  $C_\beta$  are assigned in the HNCACB. While for Leucine-142 no broadening of the  $C_\beta$  resonance is observed, the opposite is the case for Leucine-144, where the height of the  $C_\beta$  resonance is just 60 % of the expected height (based on the HNCACB of E73V/C127A/C232S hVDAC1 and the height of the  $C_\alpha$  resonance; data not shown). So at least for Leucine-144, the broadening of the  $C_\alpha$ - $C_\beta$  contact beyond detection can be justified with the liquid state spectra. On the other hand, as Leucine-142 is showing once again, liquid state and solid state NMR spectra do not necessarily deliver completely identical results.

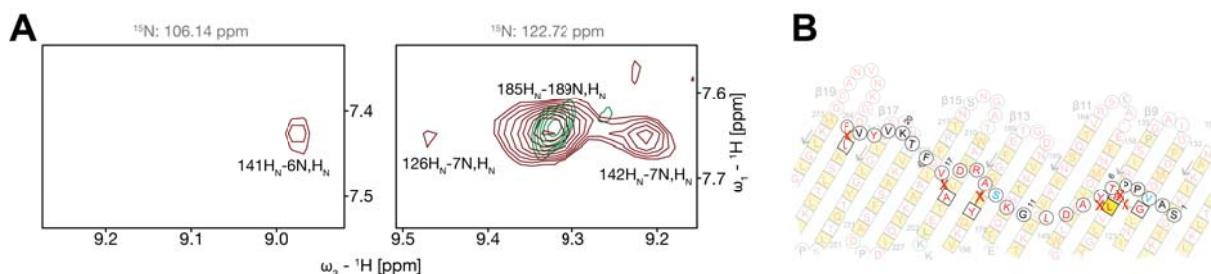
To rule out the possibility that the missing PDSD contacts are caused by the resonances shifting into bulk regions near them, the respective  $^{13}\text{C}$  chemical shifts in the liquid state HNCACB experiment were used. For Leucine-142 the  $C_\beta$  resonance in the Quintuple HNCACB actually shifts by approximately  $-0.2$  ppm in comparison to E73V/C127A/C232S hVDAC1, and as the respective

resonance is a shoulder of the Leucine bulk, the possibility that it is shifting into the latter is existing. Thus, in the above table the visibility of this resonance in the Quintuple PDSD is assessed as unclear. For Leucine-144 however,  $C_\beta$  is barely shifting at all (not more than 0.02 ppm) and  $C_\alpha$  shifts by 0.25 ppm upfield, which would put the resonance even further away from the bulk. Accordingly, the possibility that the respective PDSD contact is relocated into the bulk is very unlikely. (The respective chemical shift values can be found in Appendices B.2 and B.3.)

### 3.4.3 Missing and new NOE contacts of Quintuple hVDAC1

To further test the hypothesis that the N-terminal helix is displaced in Quintuple hVDAC1 and to probe for spatial information in Quintuple hVDAC1, a 3D  $^{15}\text{N}$  HMQC-NOESY was recorded with the usual mixing time of 240 ms. The NOE contacts between N-terminal helix and  $\beta$ -barrel were taken into special consideration, as they are a main source of information about the position of the N-terminal helix.

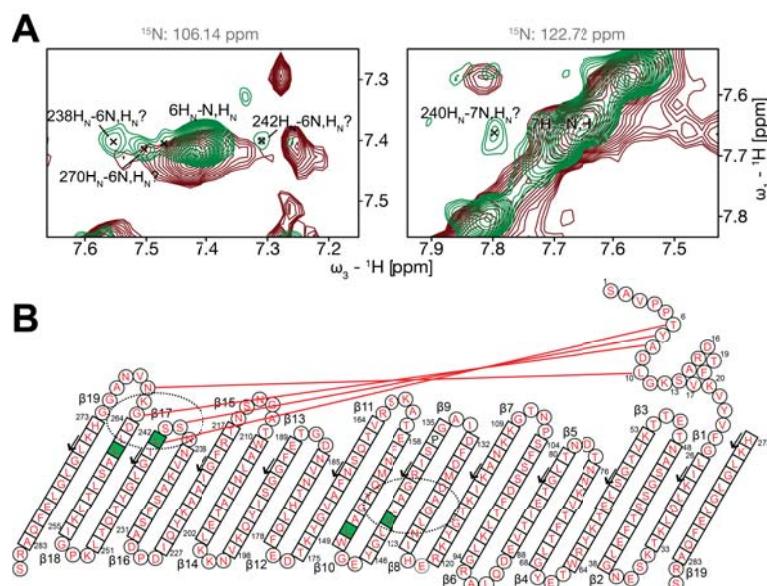
As it turns out, all 7 previously in E73V hVDAC1 assigned NOE contacts between N-terminal helix and  $\beta$ -barrel are broadened below the noise level, see Figure 3.31.



**Figure 3.31: Missing NOEs in Quintuple hVDAC1.** A: HMQC-NOESY spectra of Quintuple hVDAC1 (green) and E73V hVDAC1 (brown), showing the missing contacts of Threonine-6 (left) and Tyrosine-7 (right) while other correlations are visible. (Missing: A141-T6, G126-Y7, L142-Y7, Visible: N185-E189) B: Map of the missing NOE contacts plotted to the corresponding fragment of the topology map of hVDAC1.

As shown, other contacts in the spectrum are well retained. It should be noted that the signal strength in the two spectra is quite different, the spectrum of Quintuple hVDAC1 has a significantly worse S/N ratio. Based on the respective diagonal peaks and the reference resonances in the NOESY spectrum of E73V hVDAC1, there are the following contacts that are, however, expected to be visible: A141-T6 (expected Peak height  $2.01 \cdot 10^6$  a.u.), L142-Y7 (expected  $1.54 \cdot 10^6$  a.u.), A14-

Y195 (expected  $1.74 \cdot 10^6$  a.u.) and V17-A205 (expected  $2.28 \cdot 10^6$  a.u.). The graphically determined maximum noise level in the spectrum is  $1.42 \cdot 10^6$  a.u. However, except of the contact L142-Y7 where the data height at the position of the expected resonance is  $1.29 \cdot 10^6$  a.u. – slightly below the noise cutoff, which could mean that this peak is retained, but just influenced by noise – all other resonances are clearly no longer visible. Given this, the broadening of the resonance is pointing to that the N-terminal helix is partially displaced and partially still in place, which would be in line with the BLM and solid state measurements presented in the last chapters. (Also the results from these measurement methods did rather show a partial than a complete displacement of the helix.) Taken together, the previously reported NOEs between N-terminal helix and  $\beta$ -barrel are missing or at least broadened significantly. On the other hand, for the residues in the N-terminal helix there are new resonances visible above the noise level. Examples for these are shown in figure 3.32 A – the additional resonances are marked with “x” in the green Quintuple hVDAC1 spectra.



**Figure 3.32: Examples for additional NOEs in Quintuple hVDAC1.** A: HMQC-NOESY spectra of Quintuple hVDAC1 (green) and E73V hVDAC1 (brown), showing possible new contacts of Threonine-6 (left, possible contacts to Asparagine-238, Leucine-242, Alanine-270) and Tyrosine-7 (right, possible contact to Serine-240). B: Map of some possible new NOE contacts plotted to the topology map of hVDAC1. Residues in green are residues with hydrophobic sidechain that are pointing to the interior of the pore. The old contact region between N-terminal helix and  $\beta$ -barrel and a possible new region based on the experimental results are circled.

It was not possible to assign the here described resonances safely until now. A set of possible assignments is shown in Figure 3.32 B: T6-L242, Y7-S240, A8-G265, L10-N267. These possible

assignments, i.e. the possible new contacts for the residues in the  $\alpha$ -helix, are clustered in the loop from  $\beta$ -strand 16 to 17 and in the loop from  $\beta$ -strand 18 to 19.

This leads to the assumption that the  $\alpha$ -helix, which is – based on results from BLM and solid state NMR measurements – expected to move away from its original position at least partially, is replaced to the C-terminal  $\beta$ -strands. This would be in line with the results described at the beginning of this Chapter (significantly elevated CSDs and intensity changes in the last  $\beta$ -strands in comparison to E73V/C127A/C232S hVDAC1). Based on this, a new contact region between  $\alpha$ -helix and  $\beta$ -barrel was identified and was marked in Figure 3.32 C. However, it should again be noted that in the open-state contact region (also marked in Figure 3.32 C) no elevated CSDs could be found in the liquid state NMR spectra of Quintuple hVDAC1. In the proposed new contact region the barrel possesses a pattern of two amino acids with hydrophobic sidechains that are pointing to the interior of the barrel, namely Leucine-242 and Leucine-262. A similar pattern of Valine-143 and Leucine-150 is forming a hydrophobic patch, where Leucine-10 of the  $\alpha$ -helix is anchored in the open state of the pore. Thus, this pattern of Leucine-242 and Leucine-262 could be the anchor point for the helix in a second state of the protein. (The two residues are colored in green in Figure 3.32 C.) It is hypothesized that this second state is a closed state of hVDAC1 – as this is the state in which the protein is situated in BLM measurements to a large amount. The pattern of Leucine-242 and Leucine-262 is furthermore conserved in a high number of VDAC isoforms (see Appendix C).

#### 3.4.4 Probing the new orientation of the N-terminal helix

To accumulate more experimental evidence about the N-terminal helix moving from the old contact site at  $\beta$ -strands 8 to 10 to the proposed new contact site at  $\beta$ -strands 16 to 18, the movement and new orientation of the helix was probed with MTSL labeling.

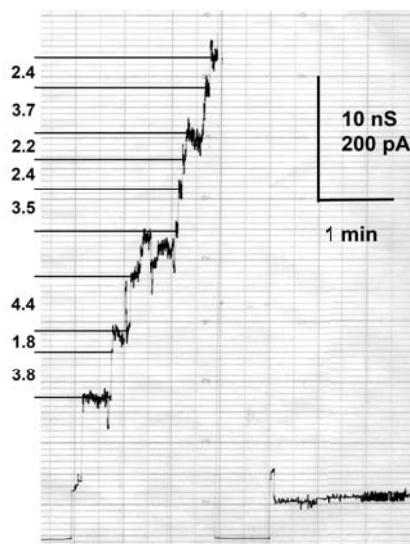
Based on Quintuple hVDAC1, an additional mutation was introduced at residue 2. Alanine-2 was mutated to Cysteine and the MTSL label was attached at that position. The resulting hexuple mutant (A2C-Quintuple hVDAC1) should show less resonance broadening due to the attached MTSL label at the old position of the N-terminal helix or of the N-terminus, respectively, than *wt* hVDAC1 labeled at this position. On the contrary, the region where the N-terminal helix is displaced to, should

### 3.4 Experiments on Quintuple hVDAC1 and development of a model of the closed state of hVDAC1

be broadened more. As mentioned earlier, the wild-type-like reference sample used for experiments on Quintuple hVDAC1 is E73V/C127A/C232S hVDAC1, thus the reference mutant for the MTSL labeling is A2C/E73V/C127A/C232S hVDAC1.

E73V/C127A/C232S hVDAC1 and Quintuple hVDAC1 have been described earlier in this thesis. The two MTSL-labeled mutants were expressed in the pET28a expression vector as usual and were labeled with MTSL prior to the refolding to avoid dimerization (see Materials and Methods for further details). Both MTSL mutants yielded nice  $^1\text{H}, ^{15}\text{N}$ -TROSY spectra without signs of resonance broadening due to intermediate chemical exchange or dimerization. Assignment of these mutants was done based on the existing assignments for E73V/C127A/C232S hVDAC1 and Quintuple hVDAC1, respectively.

To confirm that the A2C mutation and the attached MTSL label are not changing the gating properties of the channel, the MTSL labeled A2C-Quintuple hVDAC1 mutant was tested in BLM measurements. The BLM measurements were performed by Roland Benz (Department of Life Sciences and Chemistry, Jacobs University Bremen gGmbH), as usual with a part of the liquid state NMR sample reconstituted in LDAO micelles.



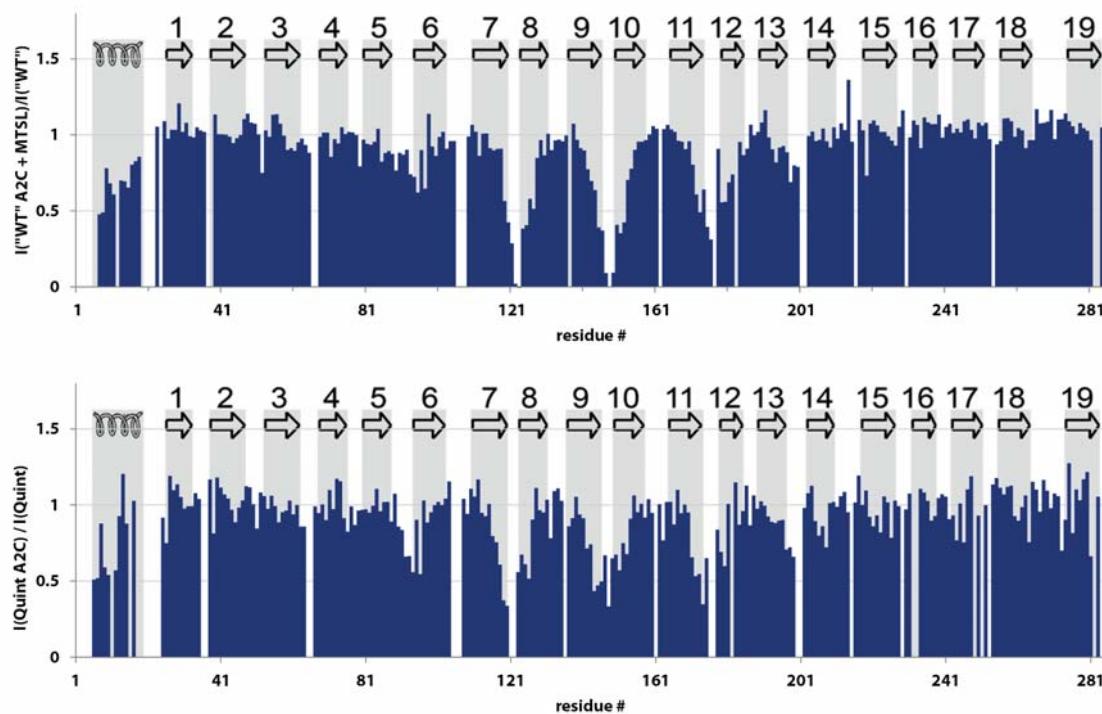
**Figure 3.33: BLM measurements on MTSL labelled A2C-Quintuple hVDAC1.** Similar to Quintuple hVDAC1 there are channels with conductances around 4 nS, resembling the open state of the protein, as well as channels with conductances around 2 nS, resembling a closed state of the protein.

Similar to Quintuple hVDAC1, also A2C-Quintuple hVDAC1 labeled with MTSL shows a great amount of channels with conductances of around 2 nS or slightly above 2 nS at low potentials that

### 3 Results

are usually not large enough to induce a closure of the channel, but also pores with 4 nS can be observed. (See Figure 3.33) This is exactly the behavior of the mutant that was expected and it is the same as for Quintuple hVDAC1. It is thus concluded that the A2C mutation and the MTS label at this position are not interfering with the gating properties of the channel and that A2C-Quintuple hVDAC1 is still stabilizing a closed state of the pore like Quintuple hVDAC1 is doing it.

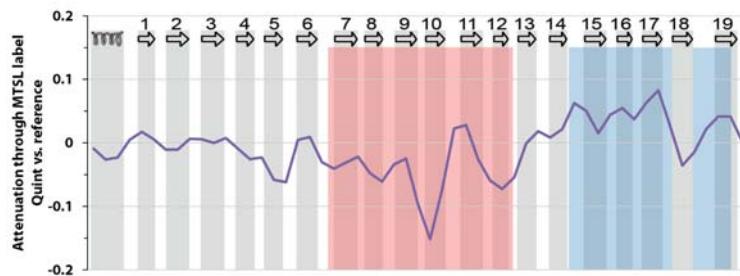
The effect of the MTS labels was now determined for both sets of mutants and the resulting intensity ratios are given in Figure 3.34.



**Figure 3.34: Comparison of PRE effects between Quintuple hVDAC1 and E73V/C127A/C232S hVDAC1.** The PRE effects induced by an MTS label introduced at residue 2 are shown by plotting the intensity ratios of the resonances of labeled and unlabeled E73V/C127A/C232S hVDAC1 (top) and Quintuple hVDAC1 (bottom) against the residue number.

Obviously in both sets of mutants the MTS label induces PREs in a region, where the label should be located if the helix is still at its old position, *i.e.* the position that it has in the open state of the pore. However, there are some residues in the C-terminal  $\beta$ -strands that also show broadening due to PRE in the Quintuple/A2C-Quintuple hVDAC1 mutant pair. As the results for this particular mutant pair are visibly more influenced by noise than the results for the other mutant pair, the difference

between the two mutant pairs was enhanced in the following way: The difference between the PRE effects of the two mutant pairs was calculated as a binomial filtered floating average of the intensity change of 3 five-residue groups, respectively. (See Chapter 2.2.5.) The result of this can be seen in Figure 3.35.



**Figure 3.35: Difference of PRE effects between Quintuple hVDAC1 and E73V/C127A/C232S hVDAC1.** The difference between the PRE effects shown in Figure 3.34 is calculated as a binomial filtered floating average of the intensity of 3 five-residue groups. Negative values indicate less pronounced PRE effects in Quintuple hVDAC1, while positive values indicate stronger PRE effects in Quintuple hVDAC1.

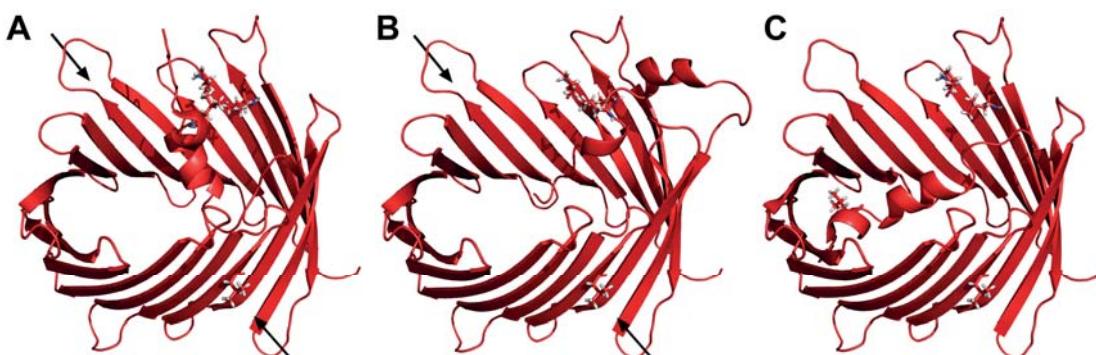
Obviously, the PRE effect is weaker at the old contact region for the mutant pair based on Quintuple hVDAC1 (resulting in a negative difference in Figure 3.35 around  $\beta$ -strand 10) and stronger in the proposed new contact region (resulting in a positive difference in Figure 3.35 around  $\beta$ -strand 17). This further supports the assumption that the N-terminal helix is replaced from its original position in Quintuple hVDAC1.

Yet, the difference between the two PRE profiles is far less pronounced than expected from BLM measurements and the results from solid state NMR that were presented in the last chapters. Based on the BLM experiments, at least 50 % of the Quintuple hVDAC1 channels should be situated in a closed state. Based on the PRE profile, however, probably not more than 10 % of the channels are closed, as the maximal attenuation of the residue intensities caused by the MTSL label is around 10 % of the normalized intensities in both regions. Obviously the micelle environment is intrinsically pushing the system to the open state, as based on methods where hVDAC1 is reconstituted into more membrane like environments far bigger amounts of closed hVDAC1 are observed.

Still, based on the collected results presented in the last chapters, a model of hVDAC1 in the closed state was derived that will be described in the following chapter.

### 3.4.5 A model for the closed state of hVDAC1

Taking all experimental data together, there is clear evidence that in Quintuple hVDAC1 the N-terminal  $\alpha$ -helix is replaced from its old position at the middle of the  $\beta$ -barrel to a new position. All the data available (CSD values, Intensity ratio Quintuple vs. E73V/C127A/C232S hVDAC1, NOE contacts, sequence analysis, PRE effects from MTSI labeling) are suggesting, that this new position of the N-terminal helix is in the C-terminal end of the  $\beta$ -barrel, in the  $\beta$ -strands 16 to 18.



**Figure 3.36: Models of the closed state of hVDAC1 incorporated in Quintuple hVDAC1.** A: Model where the N-terminal  $\alpha$ -helix is “bending back” to the C-terminal  $\beta$ -strands. B: Model in which the linker pulls the helix out of the barrel. In both models Leucine-10 is connected to the hydrophobic patch formed of Leucine-242 and Leucine-262. Arrows indicate further elliptic deformation following the displacement of the helix. Models are based on the NMR structure of E73V hVDAC1. C: NMR structure of E73V hVDAC1 with the  $\alpha$ -helix attached at the open-state contact site. Residues Leucine-10, Valine-73, Leucine-242 and Leucine-262 are shown by stick representations.

This thesis thus proposes a model, where a) the linker between helix and barrel is – coming from the  $\beta$ -barrel – conformationally largely unchanged until the G21VG23V mutation, which induces the  $\alpha$ -helix to bend back to the loops between the  $\beta$ -strands 16 to 17 and 18 to 19. Or b), the linker between helix and barrel is, due to the G21V/G23V mutation, displaced to out of the barrel, pulling the N-terminal helix into the new position attached to  $\beta$ -strands 16 to 18. Both models are depicted in Figure 3.36.

In either model, Leucine-10 is positioned such that it is now attached to the hydrophobic patch that is consisting of Leucine-242 and Leucine-262. A detailed discussion of the two models will be provided in Chapter 4.3.1.

Modeling was done with PyMol<sup>[71]</sup>, starting from the NMR structure of E73V (PDB ID: 5JPD). The resulting PDB files can be found in Appendix D.

## 4 Discussion

### 4.1 A14C/C127A/S193C/C232A hVDAC1 is not closed in NMR samples

Opening and closing of VDAC is a process that usually happens on the timescale of seconds (closing) and microseconds to milliseconds (opening) in BLM measurements.<sup>[45]</sup> It has been phenomenologically shown that the opening of the channels can, however, be slowed down by holding the channel in the closed state for longer times (which is done by applying elevated potentials for longer times).<sup>[45]</sup>

Exact values for the state switching behavior of VDAC were determined by Mertins *et al.*, they yielded an average dwelling time of  $0.17 \pm 0.03$  minutes in either of the states for mVDAC1. Contrary to that, the mutant A14C/C127A/S193C/C232A mVDAC1 usually takes a far longer time to switch into the closed state. It takes about  $30.61 \pm 6.39$  minutes (SEM), meaning that the process of closing is slowed down by two orders of magnitude, or factor 180, respectively, in this mutant. Afterwards it fails to reopen again.<sup>[52]</sup>

A kinetic description of the opening and closing process of VDAC has been provided by Marco Colombini<sup>[153]</sup>. The switching between the open and the closed state of the channel can be described with the transition state theory:  $k = \frac{k_B T}{h} \cdot \exp\left(\frac{-\Delta G}{RT}\right)$  with  $\Delta G$  being the height of the energy barrier,  $k_B$  the Boltzmann constant,  $h$  the Planck constant,  $R$  the Gas constant and  $T$  the temperature. A deceleration of the closing process like observed by Mertins *et al.* by the factor of 180 would then be equivalent to a raise of this energy barrier  $\Delta G$  for the reaction between open and closed state of 12.9 kJ/mol at 298 K. Based on his own findings<sup>[154]</sup>, according to Colombini the influence of the

applied voltage on the velocity of the closing process can be corrected with  $E = nFV$ ,  $F$  being the Faraday constant,  $V$  the applied potential and  $n$  the number of charges moving. In the case of VDAC the observed values can be fitted to  $n = 3$ .  $\Delta G$  is then corrected by  $E$  and the resulting corrected rate constants for the closing of the channel at different applied potentials fit very well to the observed values. The values Mertins *et al.* reported were measured at 40 mV.

Following the model of Colombini, the difference between 40 mV (applied potential by Mertins *et al.*) and 0 mV (potential in the NMR samples) would be equivalent to a further raise of the energy barrier between open and closed state by 11.6 kJ/mol relative to the open state. Going back to the transition state theory, this would be equivalent to a further reduction of the rate constant of closing of the channel by a factor of 107, meaning that A14C/C127A/S193C/C232A hVDAC1 has a dwelling time in the open state of at least 54.6 hours in NMR samples.

The energy difference between the open and the closed state of *wt* VDAC amounts to around 10 kJ/mol.<sup>[93]</sup> According to Boltzmann distribution in *wt* hVDAC1 the closed state is populated to about 2 % also at zero potential<sup>[93]</sup>. Given the extremely high dwelling time of A14C/C127A/S193C/C232A mVDAC1 at zero voltages it is thus very questionable, whether this population is enriched to values that would render the closed state observable by conventional TROSY experiments, as also overall denaturation of the protein over time has to be taken into account. There are ways to detect “invisible” states with NMR: CPMG relaxation dispersion experiments usually only work as long as the rates of exchange between the “visible” ground state and the “invisible” excited state lie between  $\approx 200$  and  $2000\text{ s}^{-1}$  and the population of the excited state is at least 0.5 %.<sup>[155]</sup> CEST experiments are able to access slower processes, with exchange rates of  $\approx 20$  to  $200\text{ s}^{-1}$ . DEST experiments can probe exchange processes with even lower rates of  $\approx 1$  to  $100\text{ s}^{-1}$ .<sup>[156]</sup> However, the rate of exchange in A14C/C127A/S193C/C232A hVDAC1 is obviously slower than all of these measurement methods require.

Spectroscopically, the mutant does not show elevated CSD values in regions that are distant from the mutation sites. This does not point to large conformational changes, like they are expected for the protein when it switches into the closed state. Quite the contrary to this, the mutant shows a CSD profile that is very similar to C127S/C232S hVDAC1, a mutant that does not show an unusual

gating behavior in BLM measurements at all.<sup>[146]</sup> Thus also based on this line of evidence there is no hint that A14C/C127A/S193C/C232A hVDAC1 is in a different state than it is usually manifested in NMR sample – which is the open state of the protein.

There is even the open question, whether the closed state Mertins *et al.* are seeing in their publication is really the usual closed state like it is described for *wt* hVDAC1. The fact that the mutant is trapped in the closed state contradicts the usual behavior that opening of the channel is occurring even faster than closing.<sup>[45]</sup> Furthermore, there is plethora of experimental and theoretical evidence that movement of the N-terminal helix is essential for the channel to reach a cation selective closed state. This starts with the findings that the N-terminus is part of a voltage-sensor that is moving during closure of the channel and that it is leaving the pore at least partially.<sup>[53,87,88]</sup> It continues with theoretical calculations that show that a large movement of the helix is necessary to explain the voltage dependence that is seen and that the channel can only reach a cation selective state under large deformation and without the N-terminal helix in the interior of the pore.<sup>[91,94]</sup> Furthermore, BLM measurements on a large number of mutants do also point in that direction.<sup>[90,91]</sup>

Basically there is only one publication from 2012 which suggests that VDAC can reach a closed state without movement of the N-terminal helix: Teijido *et al.* reported a L10C/C127A/A170C/C232A mVDAC1 mutant, that according to them is showing undisturbed gating.<sup>[157]</sup> Similar to the mutant studied in this thesis, the N-terminal  $\alpha$ -helix is affixed to the  $\beta$ -barrel, particularly to  $\beta$ -strand 11. The authors conclude their findings with two possible explanations, namely that either the N-terminal helix is staying inside the pore during gating or that it moves in an concerted fashion together with the rest of the voltage sensor to reach the outside of the lumen. However,  $\beta$ -strand 11 does not belong to the regions that could form the voltage sensor (but as probed by mutations that affect the voltage gating, the neighboring  $\beta$ -strands 9 and 10 do).<sup>[59]</sup> Furthermore, movement of just one or two  $\beta$ -strands during gating would imply that a large number of hydrogen bonds to these strands has to be broken during gating. As mentioned above, the gating can however be described with an energy difference of 10 kJ/mol.<sup>[93]</sup> To break one hydrogen bond in a  $\beta$ -sheet alone already 6.6 kJ/mol are needed.<sup>[158]</sup> This renders the possibility that the N-terminal  $\alpha$ -helix is moving together with one or two  $\beta$ -strands very unlikely. Additionally, the au-



#### 4 Discussion

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thors do not report measurements about the selectivity of their construct in the closed state that they are seeing.<sup>[157]</sup> Lastly, their claim that the mutant behaves “indistinguishably from the endogenous protein”<sup>[157]</sup> is in disagreement with their own data. Their data clearly shows that the mutant even closes at lower positive potentials than *wt* mVDAC1 – a behavior that remains unexplained. All in all, the publication by Teijido *et al.* does not seem to be reliable basis on which to reason that there is the possibility for the VDAC channel to close in the usual way when the N-terminal  $\alpha$ -helix is covalently bound to the  $\beta$ -barrel.

Taking all this together the most likely explanation for the behavior of A14C/C127A/S193C/C232A hVDAC1 in BLM measurements is indeed, that the  $\beta$ -barrel is collapsing, *i.e.* losing its overall structural integrity completely, due to the applied potential and that the channel closure one can see is not a real gating but just the macroscopic effect of the channel collapse. Consequently, the spectra A14C/C127A/S193C/C232A hVDAC1 delivers are spectra of the protein in the open state.



## 4.2 The structure of hVDAC1 and the N-terminal $\alpha$ -helix in micelles

Based on the three 3D structures of VDAC1 published in 2008<sup>[46,48,70]</sup> it was previously argued whether the visible difference of the appearance of the N-terminal  $\alpha$ -helix is due to different resolutions of the structures or due to dynamics of the helix. Especially the tilt of the  $\alpha$ -helix of  $\approx 90^\circ$  between the X-ray structure of mVDAC1 and the combined NMR/X-Ray structure of hVDAC1 and the different length of the helix in these two structures raised suspicion.<sup>[74]</sup> However, as mentioned in the introduction, with the new data available, this question can most likely be answered.

Based on revised chemical shift data of E73V hVDAC1 already the by TALOS-N predicted backbone torsion angles  $\psi$  and  $\phi$  indicate a kink in the N-terminal helix at position 10. (See Figure 3.5) This is confirmed by the refined NOE network, in which the first part of the helix from residues 6 to 10 is showing the characteristic (i,i+1) and (i,i+2) NOE connections. The same is true for the second part of the helix from residues 13 to 17. Between these two parts, however, the (i,i+1) NOE connections are missing.<sup>[115]</sup> Furthermore, the distance of the two residues directly before and after the kink in the X-ray structure is 5.0 Å and in the new NMR structure is 4.8 Å, which is very similar to the usual pitch of an  $\alpha$ -helix of 5.4 Å. The resolution of the combined NMR/X-ray structure by our group was only 4.1 Å, which explains why the kink can very easily be modeled as a further turn of the  $\alpha$ -helix without any problems. The presence of the kink in the N-terminal  $\alpha$ -helix, that could not be resolved in the combined NMR/X-ray by our group in 2008, is thus encoded in the NMR data and not just stemming from different dynamics of the helix in micelles and crystals.

The same arguments are true for the tilt of the N-terminal  $\alpha$ -helix that was previously observed when comparing the different structures. Refining the NOE network lead to different connections between  $\alpha$ -helix and  $\beta$ -barrel compared to what was assumed and assigned previously. This was described in detail in Chapter 3.1.3. It is very likely that this revision of the contacts between the two characteristic parts of the protein lead to the helix being positioned in the E73V hVDAC1 NMR structure the same way as in the mVDAC1 X-ray structure. The reasoning for this is that information about the linker between  $\alpha$ -helix and  $\beta$ -barrel is still very sparsely, thus the  $\alpha$ -helix is not pushed into

## 4 Discussion

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one certain orientation by this very part of the protein. Rather the connection information between  $\alpha$ -helix and  $\beta$ -barrel have a large impact on how the helix is modeled. The new assignments and connections are not only more numerous than before (7 versus 3) but also the chemical shifts of the assigned resonances fit better to the shifts of the respective connected residues. Thus, the outcome with the  $\alpha$ -helix having the same position, length and rotation in both the old X-ray and the new NMR liquid state structure is really encoded in the NOE data.

Concluding this, this answers the question raised in the thesis of Saskia Villinger, where the differences between the appearance of the  $\alpha$ -helix in the X-ray and the combined NMR/X-ray structure are stemming from. They are clearly caused by the different resolution of these two structures. A better resolved NMR structure is able to depict the  $\alpha$ -helix in the same accuracy as it was depicted in the X-ray structure previously.

### 4.2.1 The shape of the $\beta$ -barrel

The overall shape of the  $\beta$ -barrel is very much determined by the relaxation rates  $R_1$  and  $R_2$  or the anisotropy of the diffusion tensor of the protein that was determined by the relaxation rates, respectively. The resulting lowest energy structure shows a clear elliptic deformation with the minor axis of the ellipse going through Valine-73. This deformation of the barrel is resembling one of the two extremes yielded from a combination of four PCA eigenmodes generated from MD simulations using the X-ray structure of mVDAC1 and three modified versions (modifications at E73: deprotonated charged, E73V and E73Q) thereof.<sup>[132]</sup> (See Figure 3.11) The MD simulations revealed that E73V mVDAC1 embedded in a lipid bilayer is only slightly less circular than *wt* mVDAC1 (i.e. the X-ray structure of mVDAC1), having a relative excursion of no more than 0.2 nm along the eigenvector of this first eigenmode with respect to the latter. In absolute values the most probable excursion along this vector is  $\approx 0.7$  nm for E73V mVDAC1 in a phospholipid bilayer.

The newly published and in this thesis described E73V hVDAC1 NMR structure, however, matches to an excursion of 6 nm along this eigenvector. An ellipticity that large was only observed for the deprotonated charged modification ( $E73^-$ ) of mVDAC1 in the MD simulations – and even for this modification this is the largest observed excursion along the eigenvector, reached only with low

probability.<sup>[132]</sup> It is exactly this deprotonated charged modification that in the MD simulations also perturbated the DMPC membrane in which it was embedded. The membrane was thinning out at the position of E73 by roughly 0.5 nm with respect to the E73V mVDAC1. Thinning of the membrane may reduce the lateral pressure the membrane is exerting on the protein on that particular side – it could also increase the lateral pressure on the protein from the opposite side –, which then in turn can undergo conformational changes and in particular this global deformation more easily. Detergent micelles do usually also exert a different lateral pressure on IMPs than membranes.<sup>[159]</sup> The large deformation of E73V hVDAC1 in LDAO micelles, *i.e.* the large excursion along the calculated eigenvector, is thus pointing to an altered lateral pressure exerted on the protein.

Interestingly, the effect of the different lateral pressure is much more pronounced in E73V hVDAC1 in detergent micelles than in E73<sup>+</sup> in DMPC membranes. The 3D NMR structure, which is of course a structure that is resembling a mean structure out of an ensemble of present structures, is way more elliptic than E73<sup>+</sup> mVDAC1 in the MD simulations. The most probable excursion along the eigenvector of the global deformation mode is no more than 2.8 nm for E73<sup>+</sup> mVDAC1 – 3.2 nm less than E73V hVDAC1 in detergent micelles. It was this same global elliptic deformation mode that was shown to being able to lead to a closed state of the protein.<sup>[91]</sup> This obviously raises the question, whether exactly this might be the state of E73V hVDAC1 in LDAO micelles. According to Zachariae *et al.* an ellipticity of 0.5 is needed to reach a cation selective state with halved conductance in comparison to the circular channel. However, fitting the lowest energy structure of E73V hVDAC1 to an ellipse delivers a fit with an ellipticity of just 0.2 – and the long and short axis of the ellipse are swapped with respect to what was reported by Zachariae *et al.* Furthermore, the NMR data suggest that the N-terminal  $\alpha$ -helix is tightly anchored to the  $\beta$ -barrel wall, although it should leave the pore lumen according to the simulations.<sup>[91]</sup>

#### 4.2.2 The old NMR structure in the light of the new findings

These findings also shed light on the NMR structure of hVDAC1 that was published in 2008.<sup>[48]</sup> The definition of the barrel in the deposited structures (PDB ID: 2K4T) is only very modest. While the lowest energy structure that is always depicted in comparisons of the 3D structures from 2008 is

#### 4 Discussion

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nearly circular, a large number of the 20 deposited structures are very deformed, partly reaching even higher ellipticities than E73V hVDAC1. Some of the 20 deposited structures are not deformed as suggested by the global mode that was calculated, but the biggest part of the structures is. Together with the knowledge that the E73V mutation is increasing the overall stability of the  $\beta$ -barrel of the protein this leads to the conclusion that the high conformational space the 20 structures of 2K4T are occupying is an expression of the (with respect to E73V hVDAC1) unstabilized  $\beta$ -barrel together with the different lateral pressure the LDAO micelles are exerting on the protein with respect to membranes.

It is thus not that E73V hVDAC1 in micelles is more elliptic than *wt* hVDAC1 in micelles, but rather is the E73V mutation stabilizing the  $\beta$ -barrel to a single of the anyway existing deformed protein conformations.



## 4.3 Quintuple hVDAC1 as a model for the closed state

The assumption that Quintuple hVDAC1 delivers spectra of the pure closed state of the protein has one big flaw. Similar to A14C/C127A/S193C/C232A hVDAC1, large parts of the protein do not show significantly elevated CSD values with respect to the reference (in this case E73V/C127A/C232S hVDAC1, see Figure 3.27). There are especially no elevated CSDs in the  $\beta$ -strands 4 to 13, which is an indication against the possibility that large conformational changes are happening in the  $\beta$ -barrel. To completely rule this out, however, further measurements about the relaxation behavior of the residues in these strands – similar to the ones that were determined for E73V hVDAC1 – are needed. Nevertheless, the observable shifts point to some conformational changes in the beginning of  $\beta$ -strand 1, in the loop from  $\beta$ -strand 2 to 3 and in the last five  $\beta$ -strands of the barrel. For “just” two mutation sites that are additionally very close to each other, the range of these conformational changes seems to be quite large.

Additionally, there are several lines of evidence that are not based on the simple 2D TROSY spectrum, that Quintuple hVDAC1 behaves differently from previously studied *wt*-like hVDAC1 mutants. There are the following points: a) There are significant intensity changes of the resonances in Quintuple hVDAC1 with respect to E73V/C127A/C232S hVDAC1. Resonances in the C-terminal end of the  $\alpha$ -helix and  $\beta$ -strands 1, 9 and 15 to 19 as well as some in the loops from  $\beta$ -strands 6 to 7 and 8 to 9 are significantly broadened, pointing to changes in dynamics in these regions of the protein. b) In BLM measurements Quintuple hVDAC1 is often behaving like a  $\Delta(1-31)$ -hVDAC2 construct and not always like *wt* hVDAC1. It is thus to a significant percentage behaving like a mutant which is lacking the N-terminal  $\alpha$ -helix which implements the closed state. c) In solid state PDSD spectra Quintuple hVDAC1 is lacking several intra- und inter-residue correlations that are visible in *wt* hVDAC1 and are missing in  $\Delta(1-20)$ -hVDAC1, too. Taking these two points together, according to solid state NMR measurements Quintuple hVDAC1 shows features previously described for a construct lacking the N-terminal  $\alpha$ -helix and is in the closed state electrophysiologically. d) In liquid state HMQC-NOESY spectra the contacts between N-terminal  $\alpha$ -helix and  $\beta$ -barrel are missing or broadened beyond the noise level, respectively. Instead, possible new resonances are emerging for

the residues in the  $\alpha$ -helix that can tentatively be assigned to residues in the loops between  $\beta$ -strands 16 to 17 and 18 to 19, where a pattern of two hydrophobic residues is pointing to the interior of the pore. This in turn could have the same function as the respective pattern in  $\beta$ -strands 9 and 10. e) MTSL labeling at position 2 shows that resonances of residues in the  $\beta$ -strands 7 to 12 are less broadened in Quintuple hVDAC1 with respect to E73V/C127A/C232S hVDAC1, while resonances of residues in the  $\beta$ -strands 15 to 17 and 19 are showing stronger PRE effects.

As calculated and discussed in Chapter 4.1, based on the small energy difference between open and closed state, in *wt* hVDAC1 roughly 2 % of the channels should be in the closed state of the protein. The findings listed in the last paragraph point to that Quintuple hVDAC1 is situated in the closed state to a higher percentage, roughly 10 %, in liquid state samples, i.e. if reconstituted into detergent micelles. This quantitative conclusion will be derived in the following.

The finding that G21V/G23V hVDAC1 delivers non-interpretable liquid state NMR spectra can be explained with what has been discussed in Chapter 4.2: The detergent micelles exert a different lateral pressure on the protein in comparison to phospholipid bilayers. This weakens the structural integrity of the protein in the micelles. This would explain that even E73V hVDAC1 (with the E73V mutation that is stabilizing the  $\beta$ -barrel) adopts a structure that is considerably more elliptic compared to what is expected for E73V hVDAC1 embedded in lipid bilayers by MD simulations. The structure rather resembles an extreme case of a global deformation mode that is expected for charged E73<sup>-</sup> hVDAC1.<sup>[115,132]</sup> This points to that especially in detergent micelles the N-terminal  $\alpha$ -helix has a crucial function of stabilizing the protein in its overall structure. Interestingly as mentioned in Chapter 4.2.1, the long and short axis of the ellipse in the NMR structure of E73V hVDAC1 are swapped with respect to an elliptic collapsed structure that was found under low uniaxial membrane membrane stress (approximately  $-10 \text{ mN/m}$ ) in MD simulations.<sup>[91]</sup>

This in turn can explain why it is not possible to study a  $\Delta(1-20)$ -hVDAC1 construct reconstituted into detergent micelles: The barrel without the helix is destabilized that strongly, that the different lateral pressure of the detergent micelles with respect to phospholipid membranes leads to a barrel collapse. The observations made with G21V/G23V hVDAC1 are very analogous to that: based on BLM measurements it is expected that a significant relative share of the protein is in its closed



state.<sup>[145]</sup> If this is comprising movement of the helix, the same destabilization effect like what was observed for  $\Delta(1-20)$ -hVDAC1 is kicking in, resulting in non-interpretable spectra. The different lateral pressure that lipid bilayers are exerting on the protein can however stabilize the conformation of the  $\beta$ -barrel, resulting in the possibility to record interpretable solid state spectra and BLM measurements. Detergent micelles are thus inherently demanding the protein to be mostly open, because it is otherwise destabilized and collapsing.

This problem is tackled by combining the G21V/G23V mutation with the E73V mutation. While the former is inducing the movement of the N-terminal helix, respectively pushing the protein towards the closed state, the latter is stabilizing the barrel and ensures that the protein can still be studied by liquid state NMR. The result is a protein which is still mostly in the open state, but where due to the G21V/G23V mutation the energy difference between the open and the closed state is further reduced, thus a larger relative share of the channels is in the closed state of the protein than usual.

This assumption allows an explanation for the observations made with liquid state NMR. If the protein is still mostly open the fact that for large parts of the protein no elevated CSD values can be observed can be understood. The larger share of protein in the closed state, however, is manifesting itself in the CSDs in the last  $\beta$ -strands 15 to 19 and the resonance intensity changes observed (and pointing to changes in dynamics) in the very same  $\beta$ -strands.

How large the relative share of Quintuple hVDAC1 in the closed state is can be estimated with the MTS defense experiments. The comparison of the PRE effects of a label at residue 2 in Quintuple hVDAC1 and E73V/C127A/C232S hVDAC1 (See Figure 3.35) shows a maximal difference of the intensity ratios of roughly 0.1. The values presented there are normalized to the peak intensity of the single amino acid. Assuming that the maximal difference of the intensity ratios is stemming from residues that are affected in the strongest possible way by the MTS defense label, *i.e.* that are broadened beyond detection if the MTS defense label is near them in one state and are completely unaffected if the label is far away from them in the other state, this corresponds to approximately 10 % population difference of the open and closed state between Quintuple hVDAC1 and E73V/C127A/C232S hVDAC1.

DAC1. Thus it can be estimated that the share of Quintuple hVDAC1 situated in the closed state of the protein in detergent micelles is roughly 10 %, as already stated above.

The results from solid state NMR and BLM experiments do also fit into this explanation. Both of these systems are better able to stabilize the channel also in the moments when the helix is not in the interior of the pore. Thus, in these two systems Quintuple hVDAC1 is appearing to be closed to an even higher extent: far more than 10 % of the BLM traces are showing Quintuple hVDAC1 to be closed at low applied potentials as described by Geula *et al.*<sup>[145]</sup> and shown by our own measurements (see Chapter 3.4.1). The solid state PDSD spectra are also suggesting a higher share of protein without N-terminal helix in the interior as the mentioned missing contacts are quite prominent in *wt* hVDAC1 (see Chapter 3.4.2). However, that open channels can be observed in the BLM measurements and that not all PDSD contacts in question are invisible in solid state NMR experiments also suggests, that in both methods also open channels are still present.

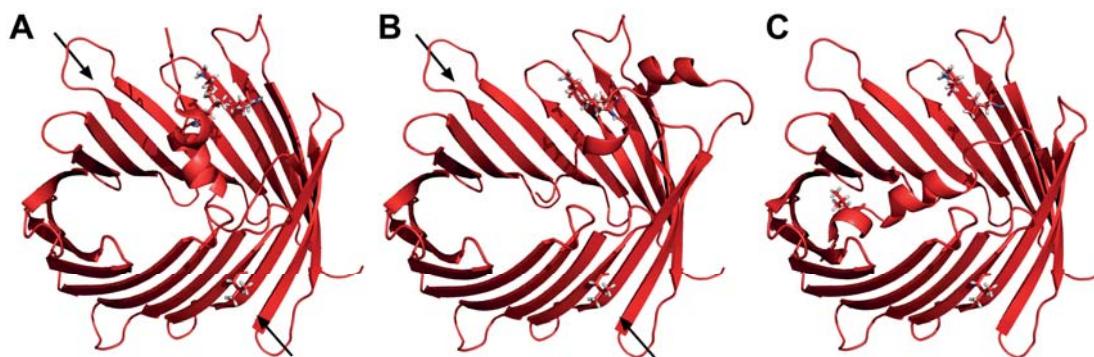
At the same time all these results are pointing to that the N-terminal  $\alpha$ -helix is indeed moving between the open and the closed state of the channel, as the results are delivering a coherent picture with exactly this assumption. However, the question arises where exactly the N-terminal helix is moving in the closed state of the protein.

All experimental evidence presented in this thesis suggests that the C-terminal  $\beta$ -strands 15 to 19 are the place where the helix is moving. These strands are where the largest CSD values and the biggest resonance intensity changes are observed. The tentative new NOE contacts of the helix residues are pointing there. Also the PRE effects observed suggest a stronger effect of the MTS label in these strands in Quintuple hVDAC1. Lastly, there is a unique and often conserved pattern of two residues with hydrophobic sidechains that are pointing to the interior of the channel in  $\beta$ -strands 17 and 18 (Leucine-242 and Leucine-262), which could form a hydrophobic patch Leucine-10 is anchored to in the closed state of the protein – similar to the hydrophobic patch formed by Valine-143 and Leucine-150.

This lead to the two possible models shown in Figure 3.36; the helix is either “bending back” to the C-terminal  $\beta$ -strands or pulled out by the linker between helix and barrel.

### 4.3.1 The conformation of the N-terminal helix in the closed state of Quintuple hVDAC1

Both the models presented in this thesis seem to reasonably fulfill the experimental restraints that were found, although some of them fit better to one or the other of the two models. The two models are shown here together with the an open state of the protein as it is observed with E73V hVDAC1.



**Figure 4.1: Models of the closed state of hVDAC1 incorporated in Quintuple hVDAC1.** A: Model where the N-terminal  $\alpha$ -helix is “bending back” to the C-terminal  $\beta$ -strands. B: Model in which the linker pulls the helix out of the barrel. In both models Leucine-10 is connected to the hydrophobic patch formed of Leucine-242 and Leucine-262. Arrows indicate further elliptic deformation following the displacement of the helix. Models are based on the NMR structure of E73V hVDAC1. C: NMR structure of E73V hVDAC1 with the  $\alpha$ -helix attached at the open-state contact site. Residues Leucine-10, Valine-73, Leucine-242 and Leucine-262 are shown by stick representations.

Model A in Figure 4.1 comprises a movement of the  $\alpha$ -helix that leads to it bending back to the loops of the C-terminal  $\beta$ -strands (“bending model”). The second part of the  $\alpha$ -helix would then be in closer spatial proximity to the C-terminal  $\beta$ -strands. This could cause conformational changes in these  $\beta$ -strands, which would explain the elevated CSD values there. If this also causes changes in dynamics, it would also explain the altered residue intensities. As is visible in the structural model, this special conformation would also induce a pull on the linker between  $\alpha$ -helix and  $\beta$ -barrel, resulting in conformational changes in the first  $\beta$ -strand – again explaining the observed elevated CSD values. Depending on the exact conformation of the first part of the helix, with this model furthermore the tentative NOE assignments could be confirmed, as the first part of the helix can be located there where it can explain the possible new NOE contacts. The model would, however, be problematic when trying to explain the cation selectivity of the closed state, as the N-terminal helix is, although significantly tilted, still completely located in the interior of the pore. Also, the

position of the MTS defense would be quite far outside of the pore: The distance from Alanine-2 would be around 28 Å from the middle of  $\beta$ -strand 17 (Glycine-246) in this model ( $C_\alpha$ - $C_\alpha$  distance). As the PRE effect of the MTS defense is detectable up to approximately 25 Å, this renders it unclear whether the observed PRE profile can be explained with the MTS defense in this position.

Model B, where the linker between helix and barrel is more or less pulling the helix away from its old position (“pulling model”), would better explain the observed PRE profile. In this model, the above mentioned distance between Alanine-2 and Glycine-246 would be around 13 Å ( $C_\alpha$ - $C_\alpha$  distance). Adding the length of the MTS defense label, it seems to be very likely, that large parts of the C-terminal  $\beta$ -strands undergo broadening due to PRE in this state. The model would also be more reasonable to form a state that has cation selectivity, as the helix in this case is visibly moved to the very rim of the pore. This is obviously better resembling the “empty barrel” that was used for the MD simulations which came out with a collapsed, cation-selective state. Furthermore, this model would probably also fit better to the observations that the N-terminal helix as voltage sensor is moving to the surface of the pore during gating. As is visible in the figure, also this model requires conformational changes in the N-terminal  $\beta$ -strands, explaining the elevated CSD values there. The elevated CSD values and intensity changes in the C-terminal  $\beta$ -strands can be explained with the proximity of the N-terminal part of the  $\alpha$ -helix to these  $\beta$ -strands. The pulling model, however, would not confirm the tentative NOE connections, as the N-terminal part of the  $\alpha$ -helix would not be located in the proximity of the loops from  $\beta$ -strand 16 to 17 and 18 to 19, where the tentative NOE contacts are located. As mentioned, it would rather be in the middle of these  $\beta$ -strands.

Both models can explain that by the suggested movement of the N-terminal  $\alpha$ -helix the  $\beta$ -barrel becomes less stabilized and thus collapses to a very elliptic state which is needed to explain the lower conductance and inverted ion selectivity of the closed state. Solely based on the new positions of the  $\alpha$ -helix, however, it can not be predicted whether this elliptic, collapsed state resembles the ellipse observed in the NMR structure of E73V<sup>[115]</sup> (long axis running through  $\beta$ -strands 1 and 9) or the one found with MD simulations on the empty barrel under uniaxial membrane stress (long axis running through  $\beta$ -strands 4 and 14).<sup>[91]</sup>



In the light of the results of Choudhary *et al.*<sup>[94]</sup>, the pulling model might be a bit more feasible. They showed that a large displacement of the  $\alpha$ -helix is needed to explain the observed voltage dependence of the gating process. In the pulling model the helix is rather moving in its entirety for a significant distance, while the bending model involves a smaller movement of the helix.

Both models are equally able to explain the results from Marco Colombini.<sup>[87]</sup> He determined a number of residues in scVDAC1 that are influencing the steepness of the voltage gating process in scVDAC1 and thus proposed that they belong to the voltage sensor, that is moving during the gating process. They will be discussed in detail in the following. Aspartate-15 and Lysine-19 are conserved among the VDACs from different species, are part of the N-terminal  $\alpha$ -helix and are thus obviously moving in the models presented here. Lysine-46 in scVDAC1 is equivalent to Serine-46 in hVDAC1. This residue is located in the C-terminal end of  $\beta$ -strand 2, which is also involved into the movements that are induced by the respective rearrangements of the linker between  $\alpha$ -helix and  $\beta$ -barrel in both proposed models. Lysine-61, Lysine-65 and Lysine-84 are equivalent to Lysine-61, Threonine-65 and Glutamate-84 in hVDAC1. These residues are located in the proximity of one of the axes of the proposed elliptic deformation that is connected with the closure of the channel. Thus, upon this elliptic deformation they are located in the region which is exhibiting the largest displacements – explaining why they have been identified as moving during the gating. The same argument is true for Glutamate-145 and Glutamate-152 (equivalent to Glutamate-147 and Glutamine-154 in hVDAC1), with the only difference that they are located near the second of the two axes of the proposed ellipse. Thus, for all these residues the findings of Colombini can be explained with the models proposed in this thesis. The only residue where the explanation of its movement has to remain unclear is Aspartate-282 (equivalent to Glutamine-282), as this residue does not move in both models proposed here.

Which of the two suggested models is more realistic has to be probed with further experiments – see Chapter 5. The possibility that each of the suggested models implements one of the two gating processes of VDAC (the one at positive and the one at negative potential) might be imaginable too.



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## 5 Conclusion & Outlook

The results of this thesis shed further light into the gating process of VDAC and increase the overall understanding of how VDAC is behaving in lipid bilayers and reconstituted into detergent micelles and which influence these environments have on the channel gating. Also, new information about structural changes during the gating process have been obtained.

The liquid state NMR assignment, which is the basis for further analysis of the channel with the means of NMR spectroscopy, was once again revised and improved in this thesis. The current assignment statistics are as follows: for *wt* hVDAC1, 88.1 %/94.8 % (whole protein/barrel) of the NH backbone resonances are assigned now. For E73VC127AC232S hVDAC1, a mutant in which the overall barrel is stabilized, the respective values are 92.1 % and 98.4 % (whole protein/barrel). For Quintuple hVDAC1, a mutant in which the closed state of the protein is stabilized, 91.4 % of the whole protein's NH backbone groups and 98.4 % of the NH backbone groups in the barrel are assigned.

Furthermore, by recording TROSY-HNCACB experiments, the assignment of the C<sub>β</sub> resonances in the protein was finally possible. This lead to 87.4 % of the C<sub>β</sub> resonances being assigned in E73VC127AC232S hVDAC1 and 54.1 % of the same being assigned in Quintuple hVDAC1.

Lastly, the existing NOE network in E73V hVDAC1 was dramatically improved. 900 new H<sub>N</sub>-H<sub>N</sub> NOE contacts raised the overall number of contacts to 1431, which is a nearly threefold improvement with respect the previous values.

Based on the improved NOE network, the improved overall assignment and previously recorded relaxation data, a new 3D structure of E73V hVDAC1 reconstituted into LDAO micelles was determined and published. The structure features an elliptic β-barrel that implements the maximal ex-

## 5 Conclusion & Outlook

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cursion of a previously calculated global elliptic deformation mode of the channel. The N-terminal  $\alpha$ -helix is, as in the X-ray structure from 2008, tightly bound to the  $\beta$ -barrel and has a short kink from residues 10 to 12. As shown in the Discussion, the ellipticity of the  $\beta$ -barrel is the manifestation of both the different lateral pressure, the detergent micelles are exerting on the barrel, and the E73V mutation.

A publication from 2012 lead to the development of a G21V/G23V/E73V/C127A/C232S hVDAC1 mutant, called Quintuple hVDAC1. It has been shown that this mutant is significantly stabilizing the closed state of the protein. Based on the NMR structure of E73V hVDAC1 and previously published results about the gating mechanism of the channel, seemingly contradictory results about how large the relative share of closed channels in this mutant is, have been understood: reconstitution in detergent micelles is causing an inherent necessity of the channel to be open. Based on MTSL labeling experiments and as shown in the Discussion, an estimated 10 % of the Quintuple hVDAC1 channels in liquid state NMR are situated in a closed state of the channel.

Several lines of evidence point to a) a movement of the N-terminal  $\alpha$ -helix upon gating and b) that the new location of the helix is at the cytosolic side of the channel at the C-terminal  $\beta$ -strands. Based on these results, two possible models for the movement of the helix upon gating have been developed and are proposed in this thesis. One of the models comprises a movement of the  $\alpha$ -helix such that it is bending back to the respective C-terminal  $\beta$ -strands. The second model proposes that the  $\alpha$ -helix is pulled out of the channel by the linker between  $\alpha$ -helix and  $\beta$ -barrel.

Further experiments or calculations are recommended to gain more insight into which of the models is a better representation of a closed state of the channel. For example, the PRE profile which is expected based on MTSL labeling at residue 2 from both of these models could be calculated and compared with the already known experimental PRE profile.

To circumvent the problems the different lateral pressure of the detergent micelles is causing, it could be tried to study Quintuple hVDAC1 reconstituted into covalently circularized nanodiscs. This system has already been shown by Nasr *et al.*<sup>[160]</sup> to deliver very nice spectra with VDAC. If these nanodiscs are better stabilizing the channel when the  $\alpha$ -helix has moved from its original position, this system could further improve the results on Quintuple hVDAC1. Also solid state NMR

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experiments on 2D crystals of Quintuple hVDAC1 could deliver further insights, first experiments on this system seem to be very promising.

Additionally, the observed resonance intensity changes in the C-terminal  $\beta$ -strands in Quintuple hVDAC1 should be further investigated. It should be checked with Het-NOE or CPMG measurements, in how far the dynamics of these  $\beta$ -strands change in Quintuple hVDAC1 with respect to *wt* hVDAC1 and E73V hVDAC1, respectively, and also, which implication that has for the channel and for the closed state of the channel.



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# Appendix A Pulse programs

In the following the pulse program used for the acquisition of the TROSY-HNCACB and the pulse programs used for measuring the  $^{15}\text{N}$  relaxation data are given.

## A.1 Pulse program of TROSY-HNCACB

```
;trhncacbetgp3d
;avance-version (07/08/13)
;TROSY-HNCACB
;3D sequence with
;    inverse correlation for triple resonance
;        via TROSY and inept transfer steps
;
;    F1(H) -> F3(N) -> F2(Caliph.,t1) -> F3(N,t2) -> F1(H,t3)
;
;on/off resonance Ca and C=O pulses using shaped pulse
;phase sensitive (t1)
;phase sensitive using Echo/Antiecho gradient selection (t2)
;using constant time in t2
;with H-1 180degree pulses in t1
;uncompensated version d25=d26
;(use parameterset TRHNCACBETGP3D)
;
;T. Schulte-Herbrueggen & O.W. Sorensen, J. Magn. Reson. 144,
; 123 - 128 (2000)
;A. Eletsky, A. Kienhoefer & K. Pervushin,
;  J. Biomol. NMR 20, 188-180 (2001)
;(M. Salzmann, G. Wider, K. Pervushin, H. Senn & K. Wuethrich,
;  J. Am. Chem. Soc. 121, 844-848 (1999))
;
;$CLASS=HighRes
;$DIM=3D
;$TYPE=
;$SUBTYPE=
;$COMMENT=
;
prosol relations=<triple>
;
#include <Avance.incl>
#include <Grad.incl>
#include <Delay.incl>
;
define list<gradient> EA5 = { 0.8750 1.0000}
```

## Appendix A Pulse programs

---

```

define list<gradient> EA7 = { 1.0000 0.6667}
define list<gradient> EA9 = { 0.6595 1.0000}

;p2=p1*2
;p22=p21*2"
;d11=30m"
;d12=20u"

;d19=1.75m"           ;shorter 1h-15n-inept delay
;d23=11m"
;d25=2.3m"
;d26=2.3m"
;d28=3.6m"

;d0=3u"
;d10=3u"
;d30=d23/2-d25/2-p14/2-p16/2-d16/2-2u"

"in0=inf1/4"
"in10=inf2/4"

"in30=in10"

"DELTA=d0*4+p2*2+larger(p14,p22)-p14-4u"
;"DELTAl=d26-p16-d16"
;"DELTAl=d19-p16-d16"           ;shorter 1h-15n-inept delay
;"DELTAl=d23-p16-d16"
;"DELTAl=d23-d16"               ;deleted gradient at this place
;"DELTAl=d28-p16-d16"
;"DELTAl=d23/2-d25/2-p14/2-d10+p21*2/3.1416"
;"DELTAl=d23/2-d25/2-p14/2-d10-p16-d16-p11+p21*2/3.1416-8u"
;"DELTAl=d25-p16-d16"
;"DELTAl=d26-p11-p16-d16-8u"

"spoff2=0"
"spoff3=0"
"spoff5=bf2*(cnst21/1000000)-o2"
"spoff8=0"

aqseq 321

1 d11 ze
2 d11
3 d1 p11:f1
  50u UNBLKGRAD

  4u p10:f1
  (p11:sp1 ph2:r):f1
  4u
  4u p11:f1

  4u p13:f3      ;new line, loading power level on 15n
  (p1 ph3):f1
  p16:gp1
  d16
  DELTAl
  (center (p2 ph2) (p22 ph1):f3 )
  DELTAl
  p16:gp1
  d16
  (p1 ph2):f1

  (p21 ph1):f3

```

```

d16 ;deleted gradient here
DELT A2
(center (p14:sp3 ph1):f2 (p22 ph1):f3 )
DELT A2
d16
(p21 ph2):f3

(p13:sp2 ph6):f2
p16:gp3
d16
DELT A3
(p14:sp3 ph1):f2
DELT A3
p16:gp3
d16
(p13:sp8 ph2):f2

d0
(p2 ph1):f1
d0
(center (p14:sp5 ph1):f2 (p22 ph1):f3 )
d0
(p2 ph1):f1
d0
(p14:sp3 ph1):f2
DELT A
(p14:sp5 ph1):f2
4u

(p13:sp2 ph9):f2
p16:gp4
d16
DELT A3
(p14:sp3 ph1):f2
DELT A3
p16:gp4
d16
(p13:sp8 ph10):f2

(p21 ph5):f3
d30
(p14:sp5 ph1):f2
d30
(center (p14:sp3 ph1):f2 (p22 ph1):f3 )
DELT A4
d10
(p14:sp5 ph1):f2
d10
DELT A5
p16:gp5*EA5
d16 p10:f1
(p11:sp1 ph3:r):f1
4u
4u p11:f1

(p1 ph7)
p16:gp6
d16
DELT A6
(center (p2 ph2) (p22 ph2):f3 ) ;deleted 13c pulse here
DELT A6
p16:gp6
d16
(p1 ph1)

p16:gp7*EA7
d16
4u

(p21 ph1):f3

```



## Appendix A Pulse programs

---

```
p16:gp8
d16
DETA7 p10:f1
(p11:spl ph4:r):f1
4u
4u p11:f1
(center (p2 ph2) (p22 ph2):f3 )
4u p10:f1
(p11:spl ph4:r):f1
4u
DETA7
p16:gp8
d16 p11:f1
(p21 ph8:r):f3

p16:gp9*EA9
d16
4u BLKGRAD

go=2 ph31
d11 mc #0 to 2
F1PH(rd10 & rd30 & rp5 & rp31 & dp9 & dp10, id0)
F2EA(igrad EA5 & igrad EA7 & igrad EA9 & ip7*2 & ip8*2, id10 & dd30 & ip5*2 & ip31*2)

exit

ph1=0
ph2=1
ph3=2
ph4=3
ph5=0 2
ph6=0 0 2 2
ph7=3
ph8=3
ph9=0
ph10=1
ph31=0 2 2 0

;p10 : 120dB
;p11 : f1 channel - power level for pulse (default)
;p13 : f3 channel - power level for pulse (default)
;sp1: f1 channel - shaped pulse 90 degree (H2O on resonance)
;sp2: f2 channel - shaped pulse 90 degree (Cali on resonance)
;sp3: f2 channel - shaped pulse 180 degree (Cali on resonance)
;sp5: f2 channel - shaped pulse 180 degree (C=O off resonance)
;sp8: f2 channel - shaped pulse 90 degree (Cali on resonance)
;
for time reversed pulse
;p1 : f1 channel - 90 degree high power pulse
;p2 : f1 channel - 180 degree high power pulse
;p11: f1 channel - 90 degree shaped pulse [1 msec]
;p13: f2 channel - 90 degree shaped pulse
;p14: f2 channel - 180 degree shaped pulse
;p16: homospoil/gradient pulse [1 msec]
;p21: f3 channel - 90 degree high power pulse
;p22: f3 channel - 180 degree high power pulse
;d0 : incremented delay (F1 in 3D) [3 usec]
;d1 : relaxation delay; 1-5 * T1
;d10: incremented delay (F2 in 3D) [3 usec]
;d11: delay for disk I/O [30 msec]
;d16: delay for homospoil/gradient recovery
;d23: 1/(4J(NCa)) [11 msec]
;d25: 1/(4J' (NH)) [2.3 msec]
;d26: 1/(4J(NH)) [2.3 msec]
;d28: 1/(8J(CaCb)) [3.6 msec]
;d30: decremented delay (F2 in 3D) = d23/2-d25/2-p14/2-p16/2-d16/2-2u
;cnst21: CO chemical shift (offset, in ppm)
;cnst23: Caliphatic chemical shift (offset, in ppm)
;o2p: Caliphatic chemical shift (cnst23)
```

## A.1 Pulse program of TROSY-HNCACB

```
;inf1: 1/SW(Cali) = 2 * DW(Cali)
;inf2: 1/SW(N) = 2 * DW(N)
;in0: 1/(4 * SW(Cali)) = (1/2) DW(Cali)
;nd0: 4
;in10: 1/(4 * SW(N)) = (1/2) DW(N)
;ndl0: 4
;in30: = in10
;NS: 8 * n
;DS: >= 16
;td1: number of experiments in F1
;td2: number of experiments in F2      td2 max = 2 * d30 / in30
;FnMODE: States-TPPI (or TPPI) in F1
;FnMODE: echo-antiecho in F2

;use gradient ratio: gp 1:gp 2:gp 3:gp 4:gp 5:gp 6:gp 7:gp 8:gp 9
;                           2:   2:   2: -80:   2:   30: 45:30.13

;for z-only gradients:
;gpz1: 2%
;gpz2: 2%
;gpz3: 2%
;gpz4: 2%
;gpz5: -80%
;gpz6: 2%
;gpz7: 30%
;gpz8: 45%
;gpz9: 30.13%

;use gradient files:
;gpnam1: SINE.100
;gpnam2: SINE.100
;gpnam3: SINE.100
;gpnam4: SINE.100
;gpnam5: SINE.100
;gpnam6: SINE.100
;gpnam7: SINE.100
;gpnam8: SINE.100
;gpnam9: SINE.100

;Processing
;PHC0(F1): 90
;PHC0(F2): 22.5

;$Id: trhncacbetgp3d,v 1.4.2.1 2007/09/14 16:17:35 ber Exp $
```



## A.2 Pulse program for measurement of $^{15}\text{N}$ - $R_1$ relaxation rates at 900 MHz

```
; 15N-T1 relaxation experiment with TROSY read-out
; for 15N, 15N13C, 2H15N and 2H15N13C labelled proteins
; modified from NL by savi 15/08/2012
; see footnotes
; as pseudo-3D

#include <Avance.incl>
#include <Grad.incl>
#include <Delay.incl>

define list<loopcounter> t1list=<$VCLIST>

define loopcounter COUNTER1
define loopcounter COUNTER2

;#define LABEL_CN ; switch on for 13C labelled samples
#define TEMP_COMPENSATION

"in0=inf2/2"

#ifdef LABEL_CN
"do=97u-p4*2+p7*0.66-p1*0.5"
"d25=20m-p15*0.5-p4*4-8u"
#else
"do=100u+p7*0.66-p1*0.5"
"d25=20m-p15*0.5"
#endif /*LABEL_CN*/

"dl1=30m"
"d21=500u-p30*0.5"
"DELTA=2.65m"
"DELTA1=2.65m"
"DELTA2=2.65m-p22-p11-300u"
"DELTA3=2.65m-p23-p10-300u"
"DELTA4=260u-p24-p1*0.66"

"d27=p24+35u"
"d30=d21*4+p2*2"

"l1=1"
"l2=1"

"cnst21=176"
"cnst22=56"
"cnst18=-1"
"cnst23=9"

"spoff4=bf2*((cnst22-cnst21)/1000000)"

aqseq 312
```

## A.2 Pulse program for measurement of $^{15}\text{N}$ - $R_1$ relaxation rates at 900 MHz

---

```

1      ze
d11

; ----- start -----
3m
3m
2      d11 do:f2
1m LOCKH_OFF
2m BLKGRAD
10u p11:f1
10u p14:f2
10u p17:f3

"COUNTER1=t1list"
"COUNTER2=cnst23*4"

(p7 ph0):f3      ; purge pulse for 15N magnetization before d1
5u

;----- Temperature compensation and d1 recovery delay-----
# ifdef TEMP_COMPENSATION

    "d17=d1-40u-((d21*2+p30)*cnst23*4)"    ; cnst23 is the highest repeat used
in T2 experiment +1

10u fq=cnst18(bf ppm):f3
10u p130:f3

10      d21
(p30 ph16):f3          ; 15N pulses are applied almost on-resonance
d21
lo to 10 times COUNTER2

10u
10u fq=0:f3
d17

# else
    d1
# endif
1m UNBLKGRAD
10u p17:f3

;----- kill steady state 15N -----
(p7 ph0):f3
5u
p20:gp6
200u

;----- first INEPT Hz-> 2HxNz -----
(p1 ph0):f1
5u
DELTAL gron0 ; soft gradient to prevent radiation damping
5u groff
(center(p1*2 ph0):f1 (p7*2 ph0):f3)
5u
DELTAL gron0
5u groff

;----- rephase 2HxNz to Nz----- -----
(p1 ph5):f1  (p7 ph0):f3
5u
DELTAL1 gron1 ; soft gradient to prevent radiation damping
5u groff
(center (p1*2 ph0):f1 (p7*2 ph0):f3)
5u
DELTAL1 gron1

```



## Appendix A Pulse programs

---

```
5u groff
(p7 ph6):f3 ; phase-cycle Nz, -Nz for Freeman-Hill decay
5u
;-----
(p1 ph2):f1 ; purge pulse to kill any residual HzNz
5u
p21:gp7 ; cleaning gradient
100u
100u p10:f1
;-----15N T1 relaxation period-----
if "COUNTER1==0" goto 77 ; jump to 77 for zero relaxation data point, (0 in t1list)

70      d25*0.5
# ifdef LABEL_CN
    3u p14:f2
    (p4*2 ph0 3u 3u p12 p4*2:sp4 ph0):f2
# endif
    d25*0.5
    (p15:sp5 ph0):f1
    d25*0.5
# ifdef LABEL_CN
    3u p14:f2
    (p4*2 ph0 3u 3u p12 p4*2:sp4 ph0):f2
# endif
    d25*0.5
    lo to 70 times COUNTER1 ; delay=t1list*2*d25 (40ms)

;-----Echo/ Anti-echo encoding for TROSY read-out-----
77      3u
    3u p14:f2
    3u p11:f1
    if "l1==1"
    {
    (p7 ph7):f3
    10u
    p25:gp5
    200u
    (p7*2 ph7):f3
    10u
    p25:gp5*-1
    }
    else
    {
    (p7 ph17):f3
    10u
    p25:gp5*-1
    200u
    (p7*2 ph17):f3
    10u
    p25:gp5
    }
;-----t1 (15N) evolution period -----
d0
# ifdef LABEL_CN
    (p4*2 ph0 3u 3u p12 p4*2:sp4 ph0):f2
# endif
    d0
;-----start TROSY read-out-----
    if "l1==1"
    {
    (p1 ph1):f1 ; Echo
    3u
    3u p10:f1
    (p11:sp11 ph11:r):f1
    6u
    }
    else
    {
    (p1 ph3):f1 ; Anti-Echo
```

## A.2 Pulse program for measurement of $^{15}\text{N}$ - $R_1$ relaxation rates at 900 MHz

---

```

3u
3u p10:f1
(p11:sp11 ph13:r):f1
6u
}
5u p11:f1
;goto 999 ; optimization of water supression
DELTAA2
p22:gp2
300u
(center (p1*2 ph0):f1 (p7*2 ph0):f3)
7u
p22:gp2
DELTAA2
300u p10:f1
-----
(p11:sp12 ph12:r):f1
5u
3u p11:f1
if "l1==1"
{
(p1 ph0):f1 (p7 ph1):f3 ; Echo
}
else
{
(p1 ph0):f1 (p7 ph3):f3 ; Anti-Echo
}
;goto 999 ; for optimization of water supression
DELTAA3
p23:gp3
200u
100u p110:f1
(center(p10 ph10:r 5u p11 p1*2 ph0 5u p110 p10 ph10:r):f1 (p7*2 ph0 d27):f3)
5u
;goto 999 ; for optimization of water supression
p23:gp3
DELTAA3
DELTAA4
(p7 ph0):f3
5u
p24:gp4 ; Echo/Anti-echo decoding gradient
999
5u
5u p131:f2
20u BLKGRAMP
-----
; ----- Acquisition and looping for T1 and E/AE -----
go=2 ph31 cpds2:f2
1m do:f2
1m LOCKH_OFF
d11 do:f2 mc #0 to 2
    F1QF(tllist.inc & iu2)
    F2EA(tllist.res & iul & ru2, id0 & ru1)
d30
1m BLKGRAD
exit

ph0=0
ph1=1
ph2=2
ph3=3
ph5=1
ph6=1 1 1 1 3 3 3 3
ph10=2
ph11=3
ph12=0
ph13=1
ph16=1
ph7=1 0 3 2
ph17=1 2 3 0

```

## Appendix A Pulse programs

---

```

ph31=1 2 3 0 3 0 1 2

;-----NOTES-----
;olp = 4.7 ppm
;o2p=176 ppm (CO)
;o3p=119 ppm

;NS=8*n
;in0=inf/2
;SW=1/(2*in0)
;echo-antiecho in N15 (process as Complex in NmrDraw before splitting the spectra)

; 1H pulses

;p1: 90 deg hard 1H pulse @p11
;p11: 1H 90 deg
;p10: 120 dB
;p10: 1200u (@ 600 MHz) 90 deg soft rectangular water flip-back pulse
;p11: 1900u (@ 600 MHz) 90 deg Sinc1.1000 water flip-back pulse (sp11,sp12)
;p15: 2000u (@ 600 MHz) 180 deg IBurp2 pulse on 1H (sp15)
;sp5: 180 deg IBurp2 pulse on 1H (sp15)
;sp11: 90 deg Sinc1.1000 water flip-back pulse
;sp12: 90 deg Sinc1.1000 water flip-back pulse
;spnam5: IBurp2
;spnam11: Sinc1.1000
;spnam12: Sinc1.1000
;spoffs5: 2340Hz @ 600 MHz (8.6 ppm) , should be centered in amide region but not touch the water

; 13C pulses

;p4: 13CO selective 180 deg (23.7*2us @ 600 MHz) @p14
;p12: 120 dB
;p14: 13C 90 deg

;sp4: 13CA selective 180 deg (23.7*2us @ 600 MHz)
;CPDPRG2: garp (aq C' decoupling)
;pcpd5: C' decoupling (140u or 280u @p131)
;p131: C' decoupling power

;15N pulses
;p7 : 90 deg hard 15N pulse @p17
;p18 : maximum duration of spin-lock; temperature compensation
;p17 :15N 90 deg

; gradients
;p20: 1000u
;p21: 200u
;p22: 300u
;p23: 1000u
;p24: 60.8u Echo/Anti-echo decoding gradient
;p25: 300u Echo/Anti-echo half-encoding gradient

;for z-only gradients
;gpz0: 3%
;gpz1: 2%
;gpz2: 10%
;gpz3: 50%
;gpz4: 33%
;gpz5: -33%
;gpz6: 30%
;gpz7: -50%

;gpnam2 SINE.10
;gpnam3 SINE.50
;gpnam4 SINE.10
;gpnam5 SINE.10

```

## A.2 Pulse program for measurement of $^{15}\text{N}$ - $R_1$ relaxation rates at 900 MHz

---

```
;gpnam6 SINE.10  
;gpnam7 SINE.10
```

## A.3 Pulse program for measurement of $^{15}\text{N}$ - $R_2$ relaxation rates at 900 MHz

```
; 15N-T2 relaxation experiment with TROSY read-out
; for 15N, 15N13C, 2H15N and 2H15N13C labelled proteins
; written according to NL by savi 15/08/2012
; see footnotes

#include <Avance.incl>
#include <Grad.incl>
#include <Delay.incl>

define list<loopcounter> cpmglist=<$VCLIST>

define loopcounter vc_max

define loopcounter COUNTER1
define loopcounter COUNTER2
define loopcounter COUNTER3

;

```
#define LABEL_CN ; switch on for 13C labelled samples
#define TEMP_COMPENSATION
```



"in0=inf2*0.5"

# ifdef LABEL_CN
"d0=97u-p4*2+p7*0.66-p1*0.5"
#else
"d0=100u+p7*0.66-p1*0.5"
#endif /*LABEL_CN*/

"dl1=30m"
"d21=500u-p30*0.5"
"DELTA=2.65m"
"DELTA1=2.65m"
"DELTA2=2.65m-p22-p11-300u"
"DELTA3=2.65m-p23-p10-300u"
"DELTA4=260u-p24-p1*0.66"
"DELTA5=d21-p1*2.15-3u"
"DELTA6=d21-p1*2.15-13u"
"DELTA7=d21-10u"
"DELTA8=d21-13u-p4*2"

"d27=p24+35u"

"l1=1"
"l2=1"
"l3=td1"

"cnst21=176"
"cnst22=56"
"cnst18=-1"

"spoff4=bf2*((cnst22-cnst21)/1000000)"

aqseq 312

1      ze
1m
```

### A.3 Pulse program for measurement of $^{15}\text{N}$ - $\text{R}_2$ relaxation rates at 900 MHz

---

```

"vc_max=0"

; ----- Count maximum number of cpmg points from cpmglist -----
9      20u
      if "cpmglist > vc_max"
      {
        20u
        "vc_max=cpmglist"
      }

      3m cpmglist.inc
      lo to 9 times 13

      3m cpmglist.res

; ----- start -----
2      d11 do:f2
      1m LOCKH_OFF
      3m
3      1m
      1m
4      3m
5      2m BLKGRAD
      10u p11:f1
      10u p14:f2
      10u p17:f3

; ----- set counters for cpmg loop and temperature correction -----
      20u
      "COUNTER1=cpmglist-1"
      20u
      "COUNTER2=(vc_max-cpmglist+2)*4"
      20u
      "COUNTER3=cpmglist-2"
      20u

      (p7 ph0):f3      ; purge pulse for  $^{15}\text{N}$  magnetization before d1
      5u

;-----temperature compensation and d1 recovery delay-----
# ifdef TEMP_COMPENSATION

      "d17=d1-40u-((d21*2+p30)*COUNTER2)"

      10u fq=cnst18(bf ppm):f3
      10u pl30:f3

      if "COUNTER2>=1"
      {
10      d21
      (p30 ph16):f3
      d21
      lo to 10 times COUNTER2
      }
      ; 15N pulses are applied far off-resonance
      10u
      10u fq=0:f3
      d17

# else
      d1
# endif
      1m UNBLKGRAD
      10u p17:f3

;----- kill steady state 15N -----

```



## Appendix A Pulse programs

---

```
(p7 ph0):f3
5u
p20:gp6
200u

;----- first INEPT Hz-> 2HxNz -----
(p1 ph0):f1
5u
DELTAn gron0 ; soft gradient to prevent radiation damping
5u groff
(center(p1*2 ph0):f1 (p7*2 ph0):f3)
5u
DELTAn gron0
5u groff

;----- rephase 2HxNz to Nz-----
(p1 ph5):f1 (p7 ph0):f3
5u
DELTAn1 gron1 ; soft gradient to prevent radiation damping
5u groff
(center (p1*2 ph0):f1 (p7*2 ph0):f3)
5u
DELTAn1 gron1
5u groff
(p7 ph6):f3 ; phase-cycle Nz, -Nz for Freeman-Hill decay
5u

;----- (p1 ph2):f1 ; purge pulse to kill any residual HzNz
5u
p21:gp7 ; cleaning gradient
100u
100u

;-----15N T1 relaxation period-----
;----- excitation pulse -----
(p7 ph18):f3 ; rotate -Nz to Ny

;---- N-1 loop -----
if "COUNTER1==0" goto 20
if "COUNTER1>=1"
{
14 5u
DELTAn7 p130:f3
5u
(p30 ph16):f3
d21
lo to 14 times COUNTER1
}

;---- 1 times with 1H decoupling composite pulse -----
d21
(p30 ph16):f3
DELTAn5
(p1 ph0 3u p1*2.3 ph1 3u p1 ph0):f1
5u
DELTAn6 gron9
5u groff
(p30 ph16):f3
5u
DELTAn7 gron9
5u groff

;---- N-2 loop -----
if "COUNTER3>=1"
{
15 5u
DELTAn7 gron9
5u groff
(p30 ph16):f3
```

### A.3 Pulse program for measurement of $^{15}\text{N}$ - $\text{R}_2$ relaxation rates at 900 MHz

---

```

5u
DETA7 gron9
5u groff
lo to 15 times COUNTER3
}

;----1x with 13C decoupling in the middle -----
5u
DETA7 gron9
5u groff
(p30 ph16):f3
5u
DETA8 gron9
5u groff
(p4*2 ph0 3u 3u p12 p4*2:sp4 ph0):f2
5u
DETA8 gron9*-1
5u groff
(p30 ph16):f3
5u
DETA7 gron9*-1
5u groff

;----N-2 loop -----
if "COUNTER3>=1"
{
16
5u
DETA7 gron9*-1
5u groff
(p30 ph16):f3
5u
DETA7 gron9*-1
5u groff
lo to 16 times COUNTER3
}

;---- 1x with 1H decoupling composite pulse -----
5u
DETA7 gron9*-1
5u groff
(p30 ph16):f3
5u
DETA6 gron9*-1
5u groff
(p1 ph0 3u p1*2.3 ph1 3u p1 ph0):f1
DETA5
(p30 ph16):f3
d21

;---- N-1 loop -----
if "COUNTER1>=1"
{
17
5u
DETA7 p130:f3
5u
(p30 ph16):f3
5u
DETA7 p17:f3
5u
lo to 17 times COUNTER1
}
goto 21

;--- if counter1 is 0 -----
20
5u
DETA7 p130:f3
5u
(p30 ph16):f3
DETA5
(p1 ph0 3u p1*2.3 ph1 3u p1 ph0):f1

```

---

## Appendix A Pulse programs

---

```

;----1x with 13C decoupling in the middle -----
5u
DETA6 gron9
5u groff
(p30 ph16):f3
5u
DETA8 gron9
5u groff
(p4*2 ph0 3u 3u p12 p4*2:sp4 ph0):f2
5u
DETA8 gron9*-1
5u groff
(p30 ph16):f3
5u
DETA6 gron9*-1
5u groff
(p1 ph0 3u p1*2.3 ph1 3u p1 ph0):f1
DETA5
(p30 ph16):f3
5u
DETA7 p17:f3
5u
;-----

;---- flip back to Nz/-Nz-----
21
(p7 ph8):f3 ; rotate back to -Nz

5u
p21:gp8 ; cleaning gradient
100u
100u

;----Echo/ Anti-echo encoding for TROSY read-out-----
77
3u p14:f2
3u p11:f1
if "l1==1"
{
(p7 ph7):f3
10u
p25:gp5
200u
(p7*2 ph7):f3
10u
p25:gp5*-1
}
else
{
(p7 ph17):f3
10u
p25:gp5*-1
200u
(p7*2 ph17):f3
10u
p25:gp5
}
;---- t1 (15N) evolution period -----
d0
#ifndef LABEL_CN
(p4*2 ph0 3u 3u p12 p4*2:sp4 ph0):f2
#endif
d0
;---- start TROSY read-out-----
if "l1==1"
{
(p1 ph1):f1 ; Echo
3u

```

### A.3 Pulse program for measurement of $^{15}\text{N}$ - $\text{R}_2$ relaxation rates at 900 MHz

---

```

3u p10:f1
(p11:sp11 ph11:r):f1
6u
}
else
{
(p1 ph3):f1 ; Anti-Echo
3u
3u p10:f1
(p11:sp11 ph13:r):f1
6u
}
5u p11:f1
;goto 999 ; optimization of water supression
DELT A2
p22:gp2
300u
(center (p1*2 ph0):f1 (p7*2 ph0):f3)
7u
p22:gp2
DELT A2
300u p10:f1
;-----
(p11:sp12 ph12:r):f1
5u
3u p11:f1
if "l1==1"
{
(p1 ph0):f1 (p7 ph1):f3 ; Echo
}
else
{
(p1 ph0):f1 (p7 ph3):f3 ; Anti-Echo
}
;goto 999 ; for optimization of water supression
DELT A3
p23:gp3
200u
100u p110:f1
(center(p10 ph10:r 5u p11 p1*2 ph0 5u p110 p10 ph10:r):f1 (p7*2 ph0 d27):f3)
5u
;goto 999 ; for optimization of water supression
p23:gp3
DELT A3
DELT A4
(p7 ph0):f3
5u
p24:gp4 ; Echo/Anti-echo decoding gradient
999
5u
5u p131:f2
20u BLKGRAMP
go=2 ph31 cpds2:f2
1m do:f2
1m LOCKH_OFF

d11 do:f2 mc #0 to 2
F1QF(cpmglist.inc & iu2)
F2EA(cpmglist.res & iul & ru2, id0 & rul)

1m
1m BLKGRAD
exit

ph0=0
ph1=1
ph2=2
ph3=3
ph5=1
ph6=1 1 1 1 3 3 3 3
ph8=0

```



## Appendix A Pulse programs

---

```
ph10=2
ph11=3
ph12=0
ph13=1
ph16=3
ph7=1 0 3 2
ph17=1 2 3 0
ph18=2
ph31=1 2 3 0 3 0 1 2

;-----NOTES-----
;olp = 4.7 ppm
;o2p=176 ppm (CO)
;o3p=119 ppm

;NS=8*n
;in0=inf/2
;SW=1/(2*in0)
;echo-antiecho in N15 (process as Complex in NmrDraw before splitting the spectra)

; 1H pulses

;p1: 90 deg hard 1H pulse @p11
;p11: 1H 90 deg
;p10: 120 dB
;p10: 1200u (@ 600 MHz) 180 deg soft rectangular water flip-back pulse
;p11: 1900u (@ 600 MHz) 90 deg Sinc1.1000 water flip-back pulse (sp11,sp12)
;p15: 2000u (@ 600 MHz) 180 deg IBurp2 pulse on 1H (sp15)
;sp5: 180 deg IBurp2 pulse on 1H (sp15)
;sp11: 90 deg Sinc1.1000 water flip-back pulse
;sp12: 90 deg Sinc1.1000 water flip-back pulse
;spnam5: IBurp2
;spnam11: Sinc1.1000
;spnam12: Sinc1.1000
;spoffss5: 2340Hz @ 600 MHz (8.6 ppm) , should be centered in amide region but not touch the water

; 13C pulses

;p4: 13CO selective 180 deg (23.7*2us @ 600 MHz) @p14
;p12: 120 dB
;p14: 13C 90 deg

;sp4: 13CA selective 180 deg (23.7*2us @ 600 MHz)
;CPDPRG2: garp (aq C' decoupling)
;pcpd5: C' decoupling (140u or 280u @p131)
;p131: C' decoupling power

;15N pulses
;p7 : 90 deg hard 15N pulse @p17
;p18 : maximum duration of spin-lock; temperature compensation
;p17 :15N 90 deg
;p18: 15N spin-lock power

; gradients
;p20: 1000u
;p21: 200u
;p22: 300u
;p23: 1000u
;p24: 60.8u Echo/Anti-echo decoding gradient
;p25: 300u Echo/Anti-echo half-encoding gradient

;for z-only gradients
;gpz0: 3%
;gpz1: 2%
;gpz2: 10%
;gpz3: 50%
;gpz4: 33%
```

### A.3 Pulse program for measurement of $^{15}\text{N}$ - $R_2$ relaxation rates at 900 MHz

---

```
;gpz5: -33%
;gpz6: 30%
;gpz7: -50%
;gpz8: 40%

;gpnam2 SINE.10
;gpnam3 SINE.50
;gpnam4 SINE.10
;gpnam5 SINE.10
;gpnam6 SINE.50
;gpnam7 SINE.10
;gpnam8 SINE.10
```



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# Appendix B Assignment tables of important hVDAC1 mutants

## B.1 Chemical shift assignment of *wt* hVDAC1

**Table B.1:** Backbone chemical shifts of *wt* hVDAC1

	H <sub>i</sub>	N	C <sub>αi</sub>	C <sub>αi-1</sub>	C' <sub>i-1</sub> *		H <sub>i</sub>	N	C <sub>αi</sub>	C <sub>αi-1</sub>	C' <sub>i-1</sub> *
M1	-	-	-	-	-	Y22	6.608	113.7	55.82	44.27	-
A2	-	-	-	-	-	G23	-	-	-	-	-
V3	-	-	-	-	-	F24	7.081	120.2	56.17	-	-
P4	-	-	-	-	-	G25	8.141	110.2	45.40	56.19*	-
P5	-	-	-	-	-	L26	7.487	118.3	53.09	45.24*	172.9
T6	7.411	106.1	60.20	61.18	174.4	I27	8.696	120.9	59.72	52.97	176.6
Y7	7.659	122.5	62.08	60.25	176.2	K28	8.760	126.8	54.87	59.87	175.0
A8	8.404	119.1	53.59	62.13*	178.7	L29	8.725	124.1	53.35	54.91	173.9
D9	7.237	115.2	53.61	53.61	177.6	D30	8.475	123.1	53.49	53.49	174.4
L10	7.366	125.4	56.04	53.43*	177.5	L31	9.112	123.9	53.67	53.67	173.9
G11	-	-	-	-	-	K32	8.611	125.3	55.05*	-	-
K12	7.278	122.2	59.54	44.58*	175.5	T33	9.017	115.9	60.13	55.06	172.2
S13	-	-	-	-	-	K34	8.131	123.7	55.18	60.01	172.9
A14	8.320	121.9	54.86	-	-	S35	8.820	120.3	57.00	55.04	175.8
R15	7.307	117.2	59.70	54.91*	-	E36	-	-	-	-	-
D16	8.559	118.6	-	-	-	N37	-	-	-	-	-
V17	7.495	120.4	66.26	-	-	G38	7.862	106.5	45.22	52.78	175.4
F18	-	-	-	-	-	L39	7.617	122.5	54.99	45.29	-
T19	-	-	-	-	-	E40	8.841	124.5	54.95	54.95	-
K20	-	-	-	-	-	F41	8.743	124.5	56.43	54.97	-
G21	-	-	-	-	-	T42	8.886	120.3	61.10*	-	-

*Appendix B Assignment tables of important hVDAC1 mutants*

	H <sub>i</sub>	N	C <sub>αi</sub>	C <sub>αi-1</sub>	C' <sub>i-1</sub> *		H <sub>i</sub>	N	C <sub>αi</sub>	C <sub>αi-1</sub>	C' <sub>i-1</sub> *
S43	9.262	121.9	56.62	-	-	L81	8.601	127.3	53.30	62.84	173.7
S44	8.631	120.4	56.48	56.48	173.0	G82	-	-	-	-	-
G45	9.095	111.4	44.98	-	173.3	T83	-	-	-	-	-
S46	9.087	116.0	56.64	44.92	172.0	E84	-	-	-	-	-
A47	9.163	123.8	49.39	56.62	172.3	I85	9.196	127.3	60.03	-	-
N48	8.320	122.3	52.52	49.44	175.3	T86	9.076	123.7	60.77	59.97	174.5
T49	-	-	-	-	-	V87	9.321	124.4	59.88	60.72	173.1
E50	8.517	119.6	57.47	64.31	-	E88	-	-	-	-	-
T51	8.095	114.1	-	-	-	D89	8.721	118.1	57.14	55.17	-
T52	7.848	108.3	-	-	-	Q90	7.847	116.6	55.36	-	175.2
K53	7.525	120.1	56.34	64.14*	172.2	L91	8.141	117.3	58.26	-	-
V54	8.595	125.1	59.77	56.41	176.8	A92	-	-	-	-	-
T55	8.760	118.5	59.78	59.78	175.2	R93	8.839	124.0	57.58	-	-
G56	8.302	109.4	45.33	59.76*	172.4	G94	9.415	115.4	45.08	57.62	177.3
S57	9.156	113.4	57.12	-	-	L95	7.909	122.7	53.72	45.09	173.7
L58	9.128	122.3	54.03	57.14	172.4	K96	9.678	132.3	55.18	53.77	175.3
E59	9.059	124.6	54.60	-	-	L97	8.716	127.0	53.52	55.09	-
T60	9.046	120.4	60.70	54.55	-	T98	9.152	120.4	61.15	53.55	174.6
K61	8.909	126.9	54.52	60.69	172.7	F99	9.470	127.0	55.96	61.11	172.9
Y62	9.343	125.1	56.92	54.52	-	D100	8.648	128.8	52.30	56.01	-
R63	8.073	124.9	-	-	-	S101	8.759	119.1	56.57	52.24	174.5
W64	8.940	126.4	54.15	-	-	S102	8.695	118.8	57.07	56.55	-
T65	-	-	-	-	-	F103	8.913	122.3	55.73	57.17	-
E66	-	-	-	-	-	S104	7.810	120.7	52.95	55.79	-
Y67	6.572	114.5	-	-	-	P105	-	-	-	-	-
G68	7.572	105.1	46.71	-	-	N106	-	-	-	-	-
L69	7.336	120.5	53.83	46.67	175.7	T107	-	-	-	-	-
T70	9.086	120.6	62.08	53.74	176.7	G108	8.230	111.9	45.27	-	-
F71	9.335	128.0	55.93	62.04	-	K109	7.468	120.5	56.43	45.28	173.7
T72	-	-	-	-	-	K110	8.463	124.4	54.80	56.43	175.8
E73	8.940	126.2	54.09	-	-	N111	8.571	120.1	52.65	54.75	175.2
K74	-	-	-	-	-	A112	8.577	124.9	51.31	52.65	176.7
W75	-	-	-	-	-	K113	9.008	119.7	54.55	51.34	-
N76	7.567	122.4	51.75	-	174.1	I114	8.795	120.4	59.25	54.67	175.7
T77	8.013	108.4	63.58	-	-	K115	9.281	126.7	54.62	59.34	174.9
D78	7.701	121.0	53.88	-	-	T116	8.688	117.1	60.13	54.77	175.9
N79	8.441	114.7	54.25	53.94*	176.0	G117	9.161	113.6	44.48	60.13	174.5
T80	7.393	113.0	62.66	-	174.1	Y118	9.153	123.5	56.55	44.51	171.1

B.1 Chemical shift assignment of wt hVDAC1

	H <sub>i</sub>	N	C <sub>αi</sub>	C <sub>αi-1</sub>	C' <sub>i-1</sub> *		H <sub>i</sub>	N	C <sub>αi</sub>	C <sub>αi-1</sub>	C' <sub>i-1</sub> *
K119	7.615	125.0	52.90	56.51	172.3	F157	9.554	127.5	56.15	52.24	173.4
R120	8.721	118.1	53.63	52.96	173.0	E158	8.076	127.7	54.87	56.23	173.5
E121	8.476	120.7	59.41	53.83	174.5	T159	7.645	117.0	64.89	54.83	177.1
H122	7.883	112.3	59.44	59.44	177.1	A160	9.152	125.0	54.44	64.96	177.1
I123	7.940	118.0	59.92	59.49	173.5	K161	-	-	-	-	-
N124	8.976	125.3	52.78	59.91	171.9	S162	7.486	114.7	58.36	54.63	175.3
L125	9.471	128.8	53.25	52.76	174.5	R163	6.834	115.0	53.71	58.34	173.1
G126	9.494	113.6	44.76	53.25	175.8	V164	8.815	124.3	62.01	53.76	174.5
C127	8.533	121.3	58.11	44.73	171.9	T165	8.634	118.8	61.27	61.93	177.0
D128	9.383	129.9	52.41	58.08	172.7	Q166	7.442	121.0	55.31	61.32*	174.6
M129	9.406	123.0	54.42	52.41	175.0	S167	8.336	119.0	57.05	55.31	174.6
D130	8.559	124.9	52.78	54.35	173.0	N168	9.120	127.2	51.53	56.93	172.7
F131	8.521	123.0	57.88	52.73	176.0	F169	8.615	118.2	56.10	51.46	171.9
D132	7.394	122.8	54.05	57.81	-	A170	8.830	122.7	51.02	56.08	173.4
I133	8.448	124.2	62.95	54.06	-	V171	8.226	116.8	60.14	51.03	176.2
A134	8.469	123.4	52.45	63.03	176.8	G172	9.445	113.0	45.70	60.16	175.5
G135	7.679	106.2	44.51	52.44	177.9	Y173	8.595	119.4	56.59	45.63	170.1
P136	-	-	-	-	-	K174	8.778	129.4	55.50	56.66	173.8
S137	8.836	118.2	57.37	62.90	177.1	T175	8.647	118.1	59.56	55.38	174.0
I138	9.103	122.5	59.58	57.36	172.7	D176	-	-	-	-	-
R139	9.043	128.4	53.86	59.58	173.4	E177	-	-	-	-	-
G140	8.741	113.3	43.76	53.83	176.1	F178	7.526	119.9	56.32	64.17*	173.9
A141	8.948	123.9	50.90	43.77	170.8	Q179	8.692	122.6	53.71	56.25	174.1
L142	9.236	123.8	53.64	50.85	174.8	L180	9.350	127.7	53.44	53.63	173.8
V143	9.006	125.2	60.91	53.60	174.1	H181	9.165	128.9	-	-	-
L144	9.358	126.1	51.99	60.94	174.7	T182	-	-	-	-	-
G145	8.178	105.4	44.64	52.00	176.2	N183	8.862	116.4	52.99	59.87*	171.8
Y146	8.501	122.9	58.87	44.73	173.0	V184	8.931	118.4	59.97	52.97	172.7
E147	9.043	125.8	57.14	58.90	173.6	N185	9.320	129.2	51.23	60.00	175.0
G148	8.117	108.8	45.22	56.94*	175.3	D186	9.032	125.1	55.85	51.32	174.9
W149	8.180	121.4	57.20	45.12	173.3	G187	8.648	105.3	46.01	55.75	175.9
L150	8.961	123.5	53.52	57.13	177.3	T188	7.644	109.0	62.47	46.11	173.3
A151	8.871	121.6	50.70	53.52	175.4	E189	7.636	122.8	54.83	62.41	-
G152	9.629	107.7	45.22	50.68	175.5	F190	8.878	125.6	55.77	54.76	174.3
Y153	9.209	122.1	56.03	45.27	171.2	G191	8.543	110.3	44.44	55.74	175.7
Q154	8.219	127.9	53.09	56.00	171.9	G192	8.635	105.6	45.42	44.49	171.7
M155	8.750	121.3	52.93	52.93	172.2	S193	9.783	114.7	56.14	45.45	171.4
N156	7.983	119.4	52.23	52.88	174.3	I194	8.784	119.0	59.60	56.26	173.3

*Appendix B Assignment tables of important hVDAC1 mutants*

	H <sub>i</sub>	N	C <sub>αi</sub>	C <sub>αi-1</sub>	C' <sub>i-1</sub> *		H <sub>i</sub>	N	C <sub>αi</sub>	C <sub>αi-1</sub>	C' <sub>i-1</sub> *
Y195	8.780	127.4	55.61	59.65	173.4	F233	9.249	127.8	55.61	56.29	-
Q196	8.264	125.5	53.65	55.58	172.9	S234	9.321	124.4	57.54	55.60	-
K197	8.916	130.4	55.71	53.61	174.5	A235	8.036	122.7	50.72	57.45	-
V198	8.585	131.0	64.83	55.73	176.1	K236	9.050	114.8	54.61	50.74	175.7
N199	8.159	113.5	-	-	-	V237	8.967	116.9	59.47	54.69	174.1
K200	-	-	-	-	-	N238	7.585	122.3	50.80	59.48	174.0
K201	-	-	-	-	-	N239	8.475	113.5	54.89	50.69	176.5
L202	7.548	122.1	54.07	-	-	S240	7.820	116.9	58.27	55.11*	175.2
E203	8.842	125.5	54.11	54.11	-	S241	8.158	109.5	59.62	58.37*	172.8
T204	8.399	112.2	59.39	54.13	174.8	L242	7.299	118.5	53.75	59.76	172.3
A205	9.038	122.4	50.75	59.37	171.6	I243	9.045	126.6	59.45	53.72	176.7
V206	8.967	116.9	59.47	50.75	174.6	G244	9.300	114.5	43.99	59.49	174.8
N207	8.885	122.5	51.96	59.34	174.4	L245	9.391	122.2	53.31	44.02	171.4
L208	8.705	121.4	54.08	51.94	172.5	G246	9.185	110.4	44.61	53.45	-
A209	9.189	126.3	51.86	54.11	175.2	Y247	9.101	125.4	56.26	44.68	170.9
W210	8.991	121.3	57.64	51.88	175.4	T248	8.367	124.3	60.74	56.22	174.0
T211	8.519	120.5	60.35	57.78	173.3	Q249	9.045	126.0	52.52	60.76*	-
A212	8.670	127.0	53.22	60.33	173.7	T250	9.250	126.0	62.56	52.49	-
G213	8.406	110.1	45.02	53.29	-	L251	8.931	130.1	-	-	-
N214	7.973	119.2	52.39	45.00	-	K252	8.776	126.6	-	-	-
S215	-	-	-	-	-	P253	-	-	-	-	-
N216	8.371	120.4	53.04	59.05	174.3	G254	8.586	110.9	47.03	62.53	-
T217	8.384	119.4	62.66	53.03	174.8	I255	8.246	121.6	59.89	46.97	-
R218	9.040	125.7	54.96	62.68	174.2	K256	8.694	127.0	54.82	59.97	175.0
F219	8.043	119.1	55.84	54.92	175.1	L257	8.809	127.7	52.98	54.71	-
G220	8.576	107.7	45.04	55.81	173.5	T258	9.597	122.8	61.12	53.15	174.3
I221	8.815	118.6	59.59	45.01	171.0	L259	9.251	129.4	52.95	61.22	-
A222	8.872	126.2	50.13	59.59	172.1	S260	8.815	115.3	56.71	52.93	-
A223	9.210	119.3	50.22	50.22	175.3	A261	9.547	120.6	51.14	56.83	172.8
K224	8.769	122.5	56.06	50.13	-	L262	8.735	124.7	52.44	51.07	173.3
Y225	9.863	128.9	55.27	56.19	-	L263	9.495	126.1	52.33	52.33	175.2
Q226	9.012	131.6	54.39	55.29	173.7	D264	9.850	124.5	52.31	52.31	175.3
I227	7.825	129.9	64.61	54.31	173.8	G265	8.465	112.3	46.01	52.32	176.8
D228	8.439	118.4	52.68	64.39	175.6	K266	8.141	117.3	56.85	46.03	173.8
P229	-	-	-	-	-	N267	7.434	117.4	51.79	56.83	177.1
D230	8.517	114.4	53.71	65.64	176.1	V268	8.511	122.0	65.05	51.80	175.0
A231	7.679	124.2	51.74	51.61	174.9	N269	8.322	117.0	54.34	65.08	177.4
C232	8.468	119.0	56.10	51.68*	175.1	A270	7.497	120.6	51.68	54.39	175.7

### B.1 Chemical shift assignment of wt hVDAC1

	H <sub>i</sub>	N	C <sub>αi</sub>	C <sub>αi-1</sub>	C' <sub>i-1</sub> *		H <sub>i</sub>	N	C <sub>αi</sub>	C <sub>αi-1</sub>	C' <sub>i-1</sub> *
G271	7.692	106.7	44.71	51.64	177.7	L279	8.684	124.5	53.33	44.17	171.7
G272	8.525	109.1	45.56	44.75	174.0	E280	8.813	125.7	-	53.50*	-
H273	7.251	120.7	54.53	45.49	173.9	F281	-	-	-	-	-
K274	8.854	119.3	54.71	54.71	176.0	Q282	-	-	-	-	-
L275	8.667	121.5	53.51	54.82	175.2	A283	8.301	126.4	51.22	53.95	174.5
G276	9.098	110.9	45.06	53.56	178.0	R284	-	-	-	-	-
L277	8.222	120.9	53.83	45.15	171.0	S285	7.816	125.6	57.05	55.73	174.1
G278	9.597	115.8	44.28	53.74	-						

\*: Shift obtained from RGS-wt hVDAC1



## B.2 Chemical shift assignment of E73VC127AC232S hVDAC1

**Table B.2:** Backbone chemical shifts of E73VC127AC232S hVDAC1

	Hi	Ni	C <sub>αi</sub>	C <sub>αi-1</sub>	C <sub>βi</sub>	C <sub>βi-1</sub>	C' i-1*
M1	-	-	-	-	-	-	-
A2	-	-	-	-	-	-	-
V3	-	-	-	-	-	-	-
P4	-	-	-	-	-	-	-
P5	-	-	-	-	-	-	-
T6	7.419	106.1	60.39	61.62	69.85	31.49	174.4
Y7	7.690	122.6	62.12	60.16	39.22	70.15	176.1
A8	8.403	119.0	53.84	62.11	-	38.97	178.9
D9	7.262	115.2	53.82	53.82	39.98	-	177.7
L10	7.369	125.3	56.13	53.60	39.01	40.05	177.4
G11	-	-	-	-	-	-	-
K12	7.278	122.2	59.48	44.91	32.64	-	-
S13	8.101	115.4	-	-	-	-	-
A14	8.328	122.0	55.22	62.70*	-	-	-
R15	7.322	117.4	59.67	55.33	29.58	-	-
D16	8.550	118.5	-	-	-	-	-
V17	7.534	120.5	66.55	56.90*	31.23	-	-
F18	-	-	-	-	-	-	-
T19	-	-	-	-	-	-	-
K20	-	-	-	-	-	-	-
G21	-	-	-	-	-	-	-
Y22	6.601	113.3	55.69	44.56	39.91	-	-
G23	-	-	-	-	-	-	-
F24	7.031	120.2	56.15*	-	-	-	174.5
G25	8.153	110.1	45.54	56.29	-	38.03	177.0
L26	7.485	118.4	53.28	45.29*	46.15	-	172.8
I27	8.699	120.9	60.02	52.93	38.24	45.97	176.5
K28	8.777	126.9	54.95	59.83	36.25	38.12	175.1
L29	8.753	124.2	53.30	55.07	45.70	36.33	173.9
D30	8.484	123.3	53.77	53.30	45.40	45.40	174.4
L31	9.113	123.6	53.58	53.58	44.90	45.40	173.9
K32	8.542	125.0	55.23	53.92	35.71	44.35	-
T33	9.044	116.3	60.28	55.35	70.04	35.88	172.1
K34	8.217	123.6	55.39	60.41	35.02	69.97	172.7

B.2 Chemical shift assignment of E73VC127AC232S hVDAC1

	Hi	Ni	C <sub>αi</sub>	C <sub>αi-1</sub>	C <sub>βi</sub>	C <sub>βi-1</sub>	C' i-1*
S35	8.767	119.7	56.91	55.36	64.83	35.24	175.6
E36	-	-	-	-	-	-	-
N37	-	-	-	-	-	-	-
G38	7.852	106.2	45.26	53.02	-	37.65	-
L39	7.815	122.5	55.30	45.37	42.84	-	-
E40	8.936	124.3	54.96	55.30	32.40	42.73	175.9
F41	8.810	124.6	56.48	54.87*	42.08	32.14	174.7
T42	8.856	120.1	60.80	56.35	71.24	42.08	174.6
S43	9.218	120.6	56.67	61.05	66.44	71.57	173.3
S44	8.604	120.4	56.62	56.62	66.01	66.46	172.9
G45	9.108	111.3	45.08	56.53	-	65.93	173.1
S46	9.094	116.0	56.78	45.08	66.45	-	172.1
A47	9.213	124.0	49.68	56.61	21.26	66.30	172.3
N48	8.305	122.4	52.74	49.66	39.41	21.06	175.5
T49	8.495	118.6	-	52.54*	-	-	-
E50	8.523	119.8	57.65	64.17*	30.19	67.47	174.2
T51	8.107	114.2	56.46*	-	-	-	176.5
T52	7.841	108.2	-	-	-	-	-
K53	7.548	120.2	56.58	64.02	36.14	68.56	172.3
V54	8.650	125.8	60.04	56.57	33.06	36.02	176.6
T55	8.785	118.7	59.83	59.83	71.47	33.06	175.2
G56	8.345	109.3	45.59	59.84	-	71.25	172.6
S57	9.100	113.1	57.33	45.50	66.27	-	171.4
L58	9.089	122.2	55.79	57.31	45.76	66.45	172.3
E59	9.046	123.5	54.77	55.71	33.45	46.23	174.8
T60	9.085	119.9	60.81	54.77	70.67	33.41	175.2
K61	8.864	126.8	54.20	60.85	36.67	70.83	171.8
Y62	9.237	124.2	57.09	54.24	42.83	36.15	174.2
R63	7.706	127.6	54.37	57.16	31.23	42.85	173.6
W64	9.152	127.9	53.46	54.38	30.91	31.23	-
T65	-	-	-	-	-	-	-
E66	-	-	-	-	-	-	-
Y67	6.486	114.0	56.20	59.14	39.20	28.20	176.2
G68	7.588	105.5	47.05	56.37	-	39.42	173.7
L69	7.276	120.2	53.66	46.67*	45.67	-	175.6
T70	9.126	120.4	62.33	53.71	70.52	45.74	176.6
F71	9.291	127.3	55.64	62.28	42.52	-	172.6
T72	9.022	120.7	61.25	55.53	70.79	42.73	174.0

*Appendix B Assignment tables of important hVDAC1 mutants*

	Hi	Ni	C <sub>αi</sub>	C <sub>αi-1</sub>	C <sub>βi</sub>	C <sub>βi-1</sub>	C' i-1*
V73	9.000	125.6	60.61	61.45	33.67	71.06	173.6
K74	9.123	125.7	54.47	60.80	36.23	33.58	173.6
W75	9.127	124.1	55.79	54.35	32.48	36.13	174.1
N76	7.603	123.2	51.39	55.63	41.28	32.72	174.7
T77	8.041	108.0	63.71	51.50	68.22	41.31	-
D78	7.652	121.1	54.31	63.64*	39.09	-	175.2
N79	8.523	115.2	54.02	54.20	37.23	39.04	176.0
T80	7.226	112.6	62.59	54.37	69.46	36.93	174.0
L81	8.658	128.0	53.24	62.60	44.80	69.48	173.4
G82	9.373	112.2	44.45	53.09	-	-	175.8
T83	8.826	114.1	59.59	44.50	71.66	-	172.6
E84	8.900	124.6	54.53	59.60	33.27	71.55	172.7
I85	9.146	126.6	60.27	54.53	41.25	33.07	175.2
T86	9.009	123.8	60.61	60.23	71.83	41.16	174.5
V87	9.378	124.9	59.90	60.62	33.76	71.66	173.0
E88	9.064	128.1	55.34	59.78	33.23	33.83	174.5
D89	8.781	117.9	57.32	55.45	38.58	33.05	175.4
Q90	7.913	117.1	55.31	57.18	28.77	-	175.3
L91	8.340	117.8	58.53	55.22	40.48	28.78	177.3
A92	7.588	116.8	51.25	58.42	20.66	41.18	176.8
R93	8.779	123.9	57.52*	51.05*	-	-	176.3
G94	9.467	115.4	45.37	57.79	-	28.41	177.3
L95	7.901	122.8	54.00	45.01*	44.46	-	173.8
K96	9.624	131.7	55.09	54.06	35.64	44.74	175.3
L97	8.794	127.0	53.86	55.10	45.29	35.66	174.0
T98	9.179	120.2	61.18	53.86	72.00	45.22	174.7
F99	9.457	127.0	56.09	61.07	41.15	72.28	172.9
D100	8.678	129.2	52.47	56.18	44.55	40.82	173.0
S101	8.672	118.8	56.90	52.32	66.38	-	174.8
S102	8.659	118.2	56.82	56.93	66.29	66.29	174.8
F103	9.112	121.3	56.06	56.89	42.59	66.30	173.1
S104	7.981	120.3	53.45	56.05	63.72	42.59	173.5
P105	-	-	-	-	-	-	-
N106	-	-	-	-	-	-	-
T107	-	-	-	-	-	-	-
G108	8.229	111.7	45.22*	-	-	-	-
K109	7.524	120.7	56.78	45.46	32.51	-	173.8
K110	8.444	123.7	54.94	56.69	34.74	32.49	175.9

	Hi	Ni	C <sub>αi</sub>	C <sub>αi-1</sub>	C <sub>βi</sub>	C <sub>βi-1</sub>	C' <sub>i-1</sub> *
N111	8.596	119.5	52.82	55.05	41.88	34.82	175.0
A112	8.585	124.3	51.44	52.74	21.88	41.62	173.5
K113	9.053	119.7	54.79	51.59	36.68	21.85	174.9
I114	8.781	120.6	59.43	54.83	40.46	36.46	175.4
K115	9.276	126.3	54.71	59.28	33.95	40.62	175.0
T116	8.818	116.8	59.99	54.66	71.37	33.39	175.8
G117	9.136	113.5	44.76	59.99	-	71.55	174.6
Y118	9.161	123.1	56.90	44.78	42.26	-	171.0
K119	7.507	124.8	52.96	56.87	34.62	-	172.2
R120	8.734	117.9	53.79	52.99	30.80	34.54	173.1
E121	8.623	120.8	59.61	53.68*	27.96	30.69	174.6
H122	7.905	112.4	59.48	59.80	32.95	28.09	177.0
I123	7.946	118.0	59.95	59.32	42.05	32.56	173.5
N124	8.988	125.3	52.86	59.95	42.02	41.12	171.9
L125	9.465	128.7	53.39	52.86	45.79	40.94	174.5
G126	9.408	113.4	45.35	53.60	-	-	175.8
A127	8.413	122.3	51.39	45.40	20.47	-	171.9
D128	9.179	123.7	52.10	51.40	42.87	20.65	172.6
M129	9.433	123.2	54.53	52.25	35.66	42.92	175.0
D130	8.580	125.5	53.02	54.53	42.86	35.69	173.1
F131	8.532	122.8	58.00	53.02	37.94	42.87	176.0
D132	7.386	122.9	54.05	58.01	42.94	38.22	174.6
I133	8.451	123.6	63.02*	53.86*	-	-	-
A134	8.457	123.5	52.72	63.13	18.23	-	176.8
G135	7.671	106.5	44.45	52.63	-	18.04	177.8
P136	-	-	-	-	-	-	-
S137	8.826	118.4	57.49	63.07	65.41	31.97	177.2
I138	9.147	122.2	59.48	57.36	40.92	65.21	172.8
R139	8.935	128.6	53.99	59.50	31.75	41.31	173.4
G140	8.756	113.1	43.97	54.09	-	31.58	176.1
A141	9.095	123.9	51.11	43.59	22.74	-	170.8
L142	9.220	123.5	53.84	50.95	46.13	22.57	174.9
V143	9.018	125.1	60.95	53.79	34.70	46.07	-
L144	9.374	126.2	52.30	61.11	45.16	34.93	174.8
G145	8.199	105.5	44.79	52.45	-	-	176.3
Y146	8.506	122.7	58.86	44.83	41.41	-	173.0
E147	9.070	125.8	57.09	58.96	33.11	41.17	173.7
G148	8.143	109.0	45.27	56.83*	-	-	175.2

*Appendix B Assignment tables of important hVDAC1 mutants*

	Hi	Ni	C <sub>αi</sub>	C <sub>αi-1</sub>	C <sub>βi</sub>	C <sub>βi-1</sub>	C' i-1*
W149	8.207	121.4	57.29	45.21	30.31	-	173.4
L150	8.977	123.4	53.80	57.29	46.89	30.59	177.3
A151	8.893	121.7	50.90	53.77	21.73	46.93	175.4
G152	9.649	107.8	45.51	50.99	-	21.91	175.3
Y153	9.227	122.3	56.04	45.57	43.63	-	171.2
Q154	8.254	127.9	53.28	56.05	31.46	43.66	172.0
M155	8.780	121.3	53.05	53.29	35.16	31.37	172.2
N156	7.988	119.3	52.66	53.07	42.69	34.99	174.3
F157	9.561	127.6	56.32	52.56	42.14	42.68	173.4
E158	8.095	127.7	55.00	56.30	30.02	42.00	173.5
T159	7.667	117.0	65.21	55.02	68.01	29.92	177.0
A160	9.103	124.9	54.36	64.91*	-	-	177.1
K161	-	-	-	-	-	-	-
S162	7.505	114.7	58.44	54.79	61.75	32.86	175.2
R163	6.847	115.1	53.94	57.81	33.26	61.73	173.0
V164	8.831	124.4	62.18	53.99	30.92	33.23	174.4
T165	8.672	118.8	61.43	62.21	68.63	31.13	177.0
Q166	7.471	121.0	55.62	61.41	32.52	-	174.6
S167	8.371	119.0	57.23	55.52	63.70	32.48	174.7
N168	9.146	127.0	51.65	57.12	41.28	63.87	172.7
F169	8.647	118.3	56.20	51.65	35.83	41.39	172.0
A170	8.865	122.8	51.15	56.51	23.46	36.12	173.4
V171	8.250	116.8	60.35	51.15	35.39	-	176.2
G172	9.456	113.1	45.79	60.33	-	-	175.4
Y173	8.609	119.5	56.70	45.81	43.25	-	170.1
K174	8.813	129.4	55.76	56.60	35.90	-	173.8
T175	8.647	117.9	59.69	55.30*	71.30	-	174.0
D176	-	-	-	-	-	-	-
E177	8.439	124.7	58.00	60.69	31.20	-	-
F178	7.553	120.0	56.40	57.97	42.04	-	173.9
Q179	8.734	122.7	53.88	56.52	31.87	41.94	174.1
L180	9.358	127.7	53.59	53.91	45.18	-	173.9
H181	9.178	128.8	55.01*	-	-	-	-
T182	-	-	-	-	-	-	-
N183	8.884	116.6	53.14	59.86*	-	-	171.8
V184	8.958	118.5	60.07	53.05	32.96	42.95	172.7
N185	9.323	129.1	51.50	59.97	38.07	32.84	175.0
D186	9.072	125.2	55.74	51.25	40.90	-	174.9

	Hi	Ni	C <sub>αi</sub>	C <sub>αi-1</sub>	C <sub>βi</sub>	C <sub>βi-1</sub>	C' i-1*
G187	8.664	105.3	46.02	55.80*	-	40.44	-
T188	7.659	109.0	62.69	46.02	71.27	-	173.2
E189	7.649	122.8	54.86	62.46*	30.80	71.41	172.5
F190	8.903	125.9	55.82	54.48*	41.12	30.97	174.3
G191	8.554	110.3	44.63	55.89	-	40.97	175.6
G192	8.660	105.7	45.57	44.58	-	-	171.7
S193	9.777	114.8	56.11	45.53	66.95	-	171.5
I194	8.799	119.2	59.88	56.07*	41.64	-	173.3
Y195	8.803	127.5	55.57	59.51	42.21	-	173.4
Q196	8.305	125.6	53.92	55.58	31.81	42.56	172.9
K197	8.907	130.3	55.91	53.87	31.44	31.82	174.4
V198	8.599	130.8	64.91	55.90	31.89	31.38	-
N199	8.178	113.6	52.56	-	37.52	-	-
K200	-	-	-	-	-	-	-
K201	-	-	-	-	-	-	-
L202	7.569	122.1	54.23	57.01	44.48	34.42	175.2
E203	8.903	125.8	54.49	54.15	34.11	44.49	-
T204	8.438	112.3	59.51	54.23	72.49	33.68	174.9
A205	9.094	122.6	50.99	59.64	23.67	72.47	171.6
V206	8.990	117.0	59.46	50.87	35.62	24.13	174.6
N207	8.916	122.7	52.32	59.55	42.32	35.55	174.4
L208	8.746	121.8	54.21	52.32	46.07	42.32	172.6
A209	9.191	126.3	52.18	54.16	22.34	45.85	175.3
W210	9.012	121.4	57.92	51.99	32.10	22.79	175.3
T211	8.536	120.3	60.47	57.84	70.74	32.15	173.3
A212	8.683	127.1	53.37	60.43	18.51	70.66	173.7
G213	8.455	110.2	44.97*	53.07*	-	-	-
N214	7.988	119.3	52.27	45.00*	39.90	-	-
S215	-	-	-	-	-	-	-
N216	8.377	120.4	52.99*	58.93*	-	-	174.3
T217	8.399	119.5	62.63	53.00*	-	-	174.8
R218	9.069	125.7	54.87	62.80	31.16	69.52	174.2
F219	8.070	119.2	55.78	54.87	42.23	31.13	175.1
G220	8.614	108.0	45.25	55.77	-	42.16	173.5
I221	8.832	119.0	59.64	45.18	41.50	-	171.0
A222	8.917	126.4	50.28	59.70	22.02	41.46	172.2
A223	9.221	119.4	50.35	50.35	23.16	22.14	175.2
K224	8.817	122.9	56.22	50.35	35.68	23.34	-

*Appendix B Assignment tables of important hVDAC1 mutants*

	Hi	Ni	C <sub>αi</sub>	C <sub>αi-1</sub>	C <sub>βi</sub>	C <sub>βi-1</sub>	C' i-1*
Y225	9.949	130.8	55.74	56.38	42.33	35.68	-
Q226	9.007	131.4	54.76	55.69	27.18	42.24	-
I227	7.982	130.3	64.87	54.31	38.46	27.41	-
D228	8.489	118.3	52.79	64.49*	40.14	-	175.5
P229	-	-	-	-	-	-	-
D230	8.539	114.4	54.20	65.54	43.06	-	176.0
A231	7.760	124.6	51.77	54.39	22.17	-	-
S232	8.714	114.4	56.41	51.74	66.28	22.25	-
F233	9.084	122.1	55.24	56.41	44.25	66.26	-
S234	9.345	124.8	57.73	55.30	65.33	44.20	-
A235	8.067	122.6	50.87	57.87	22.93	65.39	-
K236	9.039	114.6	54.72	50.78	35.91	23.15	175.7
V237	8.974	116.9	59.62	54.67	34.07	35.62	174.1
N238	7.620	122.4	51.18	59.58	41.26	34.17	174.1
N239	8.483	113.6	55.07	41.53	36.72	-	176.4
S240	7.829	116.8	58.14*	54.80*	63.05	-	-
S241	8.162	109.5	59.62*	58.08*	-	-	172.8
L242	7.321	118.6	54.04	59.56*	40.73	-	172.3
I243	9.062	126.6	59.55	54.02	40.58	40.58	176.7
G244	9.305	114.5	44.29	59.65	-	40.45	174.8
L245	9.375	122.2	53.70	44.30	46.39	-	171.4
G246	9.188	111.1	44.81	53.66	-	46.64	-
Y247	9.091	125.6	55.55	44.89	42.92	-	170.9
T248	8.533	124.8	60.95	55.60	71.37	43.09	-
Q249	9.029	126.2	52.76	52.44*	32.82	71.52	-
T250	9.203	125.8	63.05	52.27*	68.88	68.43	-
L251	8.958	130.1	56.37	-	42.50	-	-
K252	8.794	126.7	54.69*	-	-	-	-
P253	-	-	-	-	-	-	-
G254	8.558	110.8	47.14	62.43*	-	-	-
I255	8.285	121.6	59.95	47.24	39.93	-	-
K256	8.710	127.0	55.33	59.86*	36.51	-	174.9
L257	8.828	127.7	53.24	55.07	45.78	36.00	-
T258	9.583	122.7	61.14	53.14	71.25	45.64	-
L259	9.345	129.9	53.03	61.25	45.10	71.04	-
S260	8.809	115.0	56.74	53.14	66.74	45.05	-
A261	9.549	120.6	51.26	56.95	22.59	-	172.7
L262	8.764	124.8	52.95	51.26	42.55	22.87	173.4

	Hi	Ni	C <sub>αi</sub>	C <sub>αi-1</sub>	C <sub>βi</sub>	C <sub>βi-1</sub>	C' i-1*
L263	9.516	126.2	52.56	52.86	43.14	42.59	175.2
D264	9.847	124.5	52.63	52.63	39.92	43.32	175.3
G265	8.478	112.3	46.19	52.60	-	39.93	176.8
K266	8.161	117.4	56.92	46.09	31.50	-	173.8
N267	7.460	117.6	52.07	56.91	38.15	31.43	177.0
V268	8.505	121.9	65.10	52.06	31.12	38.18	175.0
N269	8.325	117.0	54.31*	65.07*	-	-	177.4
A270	7.531	120.7	51.93	54.34*	18.30	-	175.8
G271	7.725	106.7	45.00	51.66	-	-	177.5
G272	8.516	109.1	45.44	44.97	-	-	174.0
H273	7.269	120.6	54.67	45.63	31.22	-	174.0
K274	8.863	119.4	54.88	54.88	36.27	31.10	176.2
L275	8.685	121.7	54.13	54.93	45.63	36.46	175.1
G276	9.083	110.8	45.30	54.40	-	-	-
L277	8.241	121.0	54.07	45.13	46.80	-	171.1
G278	9.597	116.0	44.49	54.21	-	46.29	175.4
L279	8.741	124.8	53.34	44.17	45.86	-	171.7
E280	8.865	125.8	54.16	53.33	32.81	45.39	174.9
F281	-	-	-	-	-	-	-
Q282	-	-	-	-	-	-	-
A283	8.246	126.6	51.37	54.19	18.88	30.90	174.5
L284	-	-	-	-	-	-	-
E285	7.764	125.9	56.97	55.58*	30.57	-	174.0

\*: Shift obtained from RGS-E73V hVDAC1

### B.3 Chemical shift assignment of Quintuple hVDAC1

**Table B.3:** Backbone chemical shifts of Quintuple hVDAC1

	Hi	Ni	C <sub>αi</sub>	C <sub>αi-1</sub>	C <sub>βi</sub>	C <sub>βi-1</sub>	C <sub>i-1</sub> *
M1	-	-	-	-	-	-	-
A2	-	-	-	-	-	-	-
V3	-	-	-	-	-	-	-
P4	-	-	-	-	-	-	-
P5	-	-	-	-	-	-	-
T6	7.406	106.1	60.25	61.12	70.08	31.55	174.3
Y7	7.683	122.7	62.09	60.22	-	70.15	176.1
A8	8.396	119.1	53.73	62.09	-	-	-
D9	7.251	115.3	53.66	53.66	40.43	-	177.7
L10	7.367	125.3	55.94	53.85	-	-	177.7
G11	-	-	-	-	-	-	-
K12	7.261	122.2	59.54	44.70	-	-	-
S13	8.089	115.4	-	-	-	-	-
A14	8.354	121.6	54.85	-	-	-	-
R15	7.346	117.5	59.49	54.80	-	-	-
D16	-	-	-	-	-	-	-
V17	7.521	121.2	66.27	-	-	-	-
F18	-	-	-	-	-	-	-
T19	-	-	-	-	-	-	-
K20	-	-	-	-	-	-	-
V21	-	-	-	-	-	-	-
Y22	-	-	-	-	-	-	-
V23	-	-	-	-	-	-	-
F24	6.960	120.9	-	-	-	-	-
G25	8.224	111.1	45.46	54.09	-	-	-
L26	7.232	117.4	53.15	45.50	-	-	173.0
I27	8.360	118.3	59.81	-	-	-	-
K28	9.043	127.8	54.74	59.92	36.24	-	175.0
L29	8.885	125.3	53.29	54.64	45.49	-	174.0
D30	8.518	123.1	53.64	53.35	45.31	45.31	174.5
L31	9.078	123.8	53.80	53.61	-	-	174.0
K32	8.532	125.1	55.12	53.88	35.83	-	-
T33	9.025	116.3	60.19	55.15	-	-	-
K34	8.207	123.7	55.31	60.19	-	-	175.2

## B.3 Chemical shift assignment of Quintuple hVDAC1

	Hi	Ni	C <sub>αi</sub>	C <sub>αi-1</sub>	C <sub>βi</sub>	C <sub>βi-1</sub>	C' i-1*
S35	8.754	119.7	56.99	55.35	64.49	34.64	172.5
E36	-	-	-	-	-	-	-
N37	-	-	-	-	-	-	-
G38	7.841	106.2	45.09	52.78	-	-	-
L39	7.803	122.6	55.06	45.03	42.94	-	-
E40	8.925	124.3	54.96	54.96	-	42.47	-
F41	8.792	124.7	56.48	54.80	-	-	174.8
T42	8.843	120.2	60.81	56.49	71.69	41.94	174.5
S43	9.208	120.6	56.58	60.82	66.43	-	173.6
S44	8.557	120.5	56.50	56.50	66.19	66.19	-
G45	9.117	111.7	45.03	56.38	-	-	173.3
S46	9.059	115.6	56.75	45.01	66.16	-	171.9
A47	9.111	124.0	49.37	56.70	-	-	-
N48	8.311	122.5	52.85	49.38	-	-	175.4
T49	8.473	118.8	52.74	52.74	-	-	-
E50	8.526	119.9	57.55	-	30.25	-	-
T51	8.099	114.2	-	-	-	-	-
T52	7.879	108.2	61.17	-	-	-	-
K53	8.523	114.0	56.37	-	-	-	-
V54	8.643	125.8	59.99	56.38	33.36	-	176.8
T55	8.715	118.7	59.73	60.01	71.69	33.32	175.3
G56	8.332	109.1	45.57	59.69	-	-	-
S57	9.082	113.0	57.26	45.63	-	-	171.4
L58	9.089	122.2	54.05	57.27	45.83	-	172.5
E59	9.044	123.6	54.75	54.07	33.18	-	174.7
T60	9.071	120.0	60.68	54.74	70.38	-	175.3
K61	8.856	126.8	54.16	60.67	36.79	-	171.8
Y62	9.227	124.2	57.11	54.12	-	-	174.6
R63	7.700	127.7	54.30	57.14	31.21	42.70	173.5
W64	9.146	128.0	53.47	54.30	30.05	-	173.6
T65	-	-	-	-	-	-	-
E66	-	-	-	-	-	-	-
Y67	6.476	114.0	55.94	59.17	39.10	-	-
G68	7.578	105.6	46.75	56.03	-	-	-
L69	7.262	120.3	53.61	46.73	45.58	-	175.5
T70	9.114	120.4	62.12	53.61	70.64	-	-
F71	9.274	127.3	55.54	62.16	42.46	-	172.5
T72	9.008	120.8	61.24	55.59	70.49	-	174.0

*Appendix B Assignment tables of important hVDAC1 mutants*

	Hi	Ni	C <sub>αi</sub>	C <sub>αi-1</sub>	C <sub>βi</sub>	C <sub>βi-1</sub>	C' i-1*
V73	8.990	125.7	60.64	61.21	33.83	-	173.7
K74	9.114	125.8	54.45	60.55	36.03	33.44	173.6
W75	9.096	124.0	55.58	54.47	-	-	-
N76	7.616	123.2	51.35	55.62	40.83	-	174.7
T77	8.047	108.2	63.76	51.37	-	-	-
D78	7.642	121.1	53.95	63.85	39.33	-	-
N79	8.509	115.2	54.01	54.01	37.16	-	-
T80	7.241	112.6	62.62	54.06	-	-	174.0
L81	8.640	128.0	53.00	62.62	44.55	-	173.4
G82	9.353	112.1	44.42	52.98	-	-	175.9
T83	8.816	114.1	59.59	44.41	71.92	-	172.5
E84	8.884	124.6	54.44	59.52	33.65	72.10	172.7
I85	9.130	126.6	60.20	54.44	41.46	-	175.3
T86	8.994	123.9	60.68	60.14	71.95	-	-
V87	9.369	125.0	59.65	60.77	33.83	71.56	173.0
E88	9.056	128.2	55.13	59.72	32.94	-	174.5
D89	8.766	118.0	57.14	55.14	38.77	33.08	175.5
Q90	7.904	117.2	55.13	57.09	-	-	175.4
L91	8.331	117.9	58.46	55.15	40.50	-	177.4
A92	7.577	116.9	51.21	58.44	-	-	176.9
R93	8.769	124.0	57.68	51.05	-	-	-
G94	9.462	115.4	45.12	57.68	-	-	177.3
L95	7.892	122.8	53.86	45.08	44.14	-	173.9
K96	9.616	131.8	55.09	53.88	35.01	-	175.5
L97	8.785	127.1	53.65	55.21	45.22	-	174.2
T98	9.169	120.3	61.21	53.65	72.11	-	174.7
F99	9.442	127.0	56.03	61.20	41.10	-	172.9
D100	8.664	129.3	52.32	56.00	44.97	41.39	173.0
S101	8.660	118.9	57.01	52.33	66.51	-	174.8
S102	8.651	118.2	56.98	56.98	66.45	66.45	-
F103	9.102	121.3	55.85	57.10	42.60	-	173.1
S104	7.975	120.3	53.26	55.88	63.59	42.66	173.6
P105	-	-	-	-	-	-	-
N106	-	-	-	-	-	-	-
T107	-	-	-	-	-	-	-
G108	8.223	111.7	-	-	-	-	-
K109	7.514	120.8	56.59	45.31	32.86	-	173.8
K110	8.435	123.7	54.73	56.59	34.82	-	176.0

	Hi	Ni	C <sub>αi</sub>	C <sub>αi-1</sub>	C <sub>βi</sub>	C <sub>βi-1</sub>	C' <sub>i-1</sub> *
N111	8.572	119.5	52.70	54.74	42.30	-	175.1
A112	8.578	124.3	51.57	52.71	22.35	-	173.5
K113	9.040	119.8	54.66	51.54	36.63	-	175.0
I114	8.768	120.6	59.29	54.67	40.34	-	175.3
K115	9.257	126.4	54.56	59.18	33.66	-	174.9
T116	8.811	116.9	59.90	54.56	-	32.92	175.8
G117	9.125	113.5	44.63	59.90	-	-	174.4
Y118	9.151	123.1	56.70	44.62	-	-	171.0
K119	7.504	124.9	52.97	56.63	34.83	-	172.2
R120	8.730	118.1	53.86	52.99	30.91	-	-
E121	8.597	120.8	59.60	53.82	-	-	174.6
H122	7.889	112.4	59.60	59.78	28.32	-	-
I123	7.943	118.0	59.95	59.59	41.95	-	-
N124	8.975	125.3	52.82	59.96	41.10	-	-
L125	9.452	128.8	53.30	52.84	45.78	-	174.4
G126	9.399	113.5	45.19	53.22	-	-	175.9
A127	8.404	122.3	51.32	45.19	20.64	-	171.2
D128	9.167	123.7	52.34	51.31	42.73	-	175.3
M129	9.423	123.3	54.43	52.30	35.41	-	174.9
D130	8.565	125.5	52.83	54.40	42.92	35.90	173.2
F131	8.515	122.9	57.80	52.98	37.90	42.74	175.9
D132	7.374	123.0	53.94	57.81	42.84	37.91	174.7
I133	8.425	123.8	63.03	54.05	37.60	-	-
A134	8.442	123.6	52.48	63.04	18.48	-	176.8
G135	7.660	106.5	44.33	52.47	-	18.33	177.8
P136	-	-	-	-	-	-	-
S137	8.809	118.5	57.35	62.87	65.53	-	177.2
I138	9.138	122.3	59.46	57.35	41.09	-	172.8
R139	8.921	128.7	53.83	59.42	31.74	-	173.4
G140	8.746	113.1	43.72	53.93	-	-	176.1
A141	9.089	123.9	51.00	43.75	23.00	-	170.8
L142	9.212	123.5	53.72	50.91	45.92	22.89	174.9
V143	9.008	125.2	60.97	53.73	34.82	-	-
L144	9.367	126.3	52.06	61.00	45.18	34.89	174.7
G145	8.189	105.5	44.73	52.05	-	-	176.2
Y146	8.491	122.7	59.00	44.74	-	-	172.9
E147	9.060	125.8	57.25	58.94	-	-	173.7
G148	8.129	109.0	45.28	57.31	-	-	-

*Appendix B Assignment tables of important hVDAC1 mutants*

	Hi	Ni	C <sub>αi</sub>	C <sub>αi-1</sub>	C <sub>βi</sub>	C <sub>βi-1</sub>	C' i-1*
W149	8.193	121.5	57.26	45.10	-	-	173.2
L150	8.968	123.5	53.56	57.27	-	-	177.3
A151	8.886	121.8	50.74	53.55	21.91	-	175.4
G152	9.640	107.8	45.29	50.76	-	-	-
Y153	9.217	122.3	55.97	45.28	43.57	-	171.2
Q154	8.243	128.0	53.12	56.02	31.55	-	172.0
M155	8.770	121.3	52.93	53.17	34.82	-	172.2
N156	7.976	119.4	52.34	52.83	42.72	-	-
F157	9.552	127.7	56.21	52.36	42.27	-	173.4
E158	8.087	127.8	54.87	56.14	29.93	41.84	173.6
T159	7.656	117.0	64.96	54.93	-	-	176.9
A160	9.112	125.0	54.48	65.01	-	-	-
K161	-	-	-	-	-	-	-
S162	7.493	114.8	58.41	54.59	61.51	-	175.2
R163	6.840	115.1	53.79	58.35	33.32	-	173.1
V164	8.822	124.5	62.02	53.78	30.87	-	174.5
T165	8.660	118.9	61.31	61.92	68.50	-	-
Q166	7.455	121.1	55.39	-	-	-	-
S167	8.355	119.1	57.16	55.40	63.83	-	174.8
N168	9.138	127.1	51.53	57.14	41.45	-	172.7
F169	8.634	118.3	56.15	51.50	43.93	-	172.0
A170	8.856	122.8	51.07	56.16	23.68	-	-
V171	8.233	116.9	60.20	51.08	-	-	176.1
G172	9.452	113.1	45.72	60.13	-	-	-
Y173	8.603	119.6	56.63	45.67	43.21	-	-
K174	8.787	129.5	55.48	56.66	-	-	-
T175	8.651	118.2	59.45	-	-	-	173.7
D176	-	-	-	-	-	-	-
E177	-	-	-	-	-	-	-
F178	7.545	120.2	56.38	-	-	-	-
Q179	8.753	122.8	53.76	56.34	-	-	174.1
L180	9.356	127.9	53.38	53.69	-	-	-
H181	9.179	128.9	-	-	-	-	-
T182	-	-	-	-	-	-	-
N183	8.874	116.5	53.05	-	-	-	-
V184	8.945	118.6	60.04	52.98	32.87	-	172.7
N185	9.319	129.2	51.34	60.03	38.43	-	175.0
D186	9.062	125.3	55.78	51.32	40.87	-	174.9

B.3 Chemical shift assignment of Quintuple hVDAC1

	Hi	Ni	C <sub>αi</sub>	C <sub>αi-1</sub>	C <sub>βi</sub>	C <sub>βi-1</sub>	C' <sub>i-1</sub> *
G187	8.659	105.4	46.03	55.85	-	-	-
T188	7.651	109.1	62.52	46.09	-	-	-
E189	7.640	122.9	54.70	62.44	-	-	-
F190	8.894	125.9	55.82	-	-	-	174.4
G191	8.541	110.4	44.54	55.86	-	41.64	175.6
G192	8.650	105.7	45.54	44.49	-	-	171.6
S193	9.777	114.8	56.26	-	67.46	-	-
I194	8.798	119.2	59.59	56.36	-	-	-
Y195	8.791	127.5	55.45	59.55	35.86	-	-
Q196	8.324	125.8	53.73	55.65	31.83	-	172.9
K197	8.884	130.1	55.71	53.68	31.46	31.46	174.5
V198	8.573	130.8	64.91	55.70	-	-	-
N199	8.177	113.7	52.12	-	-	-	-
K200	-	-	-	-	-	-	-
K201	-	-	-	-	-	-	-
L202	7.559	122.2	54.09	56.72	44.48	-	175.2
E203	8.879	125.8	54.16	-	33.71	-	174.1
T204	8.463	112.6	59.46	54.19	72.69	-	-
A205	9.061	122.7	50.71	59.47	23.57	-	171.7
V206	8.995	117.2	59.46	50.65	-	-	-
N207	8.869	122.6	51.95	59.45	41.87	-	-
L208	8.700	121.5	54.20	51.98	45.93	-	-
A209	9.208	126.5	51.94	54.25	-	-	175.3
W210	9.006	121.4	57.68	51.91	-	-	175.3
T211	8.522	120.4	60.35	57.58	-	-	173.3
A212	8.674	127.2	53.23	60.36	17.52	-	173.8
G213	8.437	110.2	-	-	-	-	-
N214	7.976	119.4	-	-	-	-	-
S215	-	-	-	-	-	-	-
N216	8.366	120.5	53.12	-	-	-	-
T217	8.385	119.5	62.63	53.06	-	-	-
R218	9.060	125.8	55.01	62.73	31.32	-	174.3
F219	8.038	119.1	55.84	54.95	42.34	-	-
G220	8.560	107.7	45.10	55.85	-	-	-
I221	8.809	118.5	59.59	45.16	41.77	-	171.0
A222	8.927	126.4	50.21	59.60	22.40	-	172.4
A223	9.123	119.3	50.33	50.20	23.06	-	-
K224	8.805	123.1	56.25	50.35	35.43	-	-

*Appendix B Assignment tables of important hVDAC1 mutants*

	Hi	Ni	C <sub>αi</sub>	C <sub>αi-1</sub>	C <sub>βi</sub>	C <sub>βi-1</sub>	C' i-1*
Y225	9.875	130.9	55.48	56.24	-	-	-
Q226	9.009	131.7	54.58	55.39	27.36	-	173.7
I227	8.009	130.3	64.59	54.52	-	-	-
D228	8.463	118.7	52.66	64.57	40.09	-	-
P229	-	-	-	-	-	-	-
D230	8.521	114.3	54.03	65.74	43.02	-	176.3
A231	7.803	124.9	51.68	53.94	-	-	-
S232	9.068	124.7	-	-	-	-	-
F233	9.289	123.3	56.07	-	-	-	-
S234	9.027	124.6	57.45	55.97	-	-	-
A235	8.060	122.8	50.73	57.43	-	-	-
K236	8.944	114.8	54.63	50.65	-	-	-
V237	8.946	117.1	59.59	54.62	-	-	173.7
N238	7.563	122.6	50.83	59.54	41.15	-	174.0
N239	8.488	113.7	54.85	50.68	-	-	176.3
S240	7.796	116.9	58.24	-	-	-	-
S241	8.149	109.6	59.69	58.18	-	-	172.8
L242	7.325	118.8	53.88	59.76	40.56	-	172.2
I243	9.078	126.7	59.39	53.82	40.29	-	176.8
G244	9.154	114.2	43.75	59.44	-	-	175.1
L245	9.349	121.4	53.39	43.66	-	-	171.7
G246	9.180	110.0	44.69	53.29	-	-	-
Y247	9.208	127.0	55.91	44.63	-	-	-
T248	8.365	125.1	54.05	56.11	-	-	-
Q249	8.894	123.5	53.10	-	-	-	171.8
T250	9.237	126.3	62.89	53.09	68.73	-	174.8
L251	9.470	128.8	61.04	-	-	-	-
K252	8.794	127.3	54.89	60.85	-	-	-
P253	-	-	-	-	-	-	-
G254	8.536	110.8	47.09	62.49	-	-	-
I255	8.268	121.7	59.90	47.07	-	-	-
K256	8.685	127.3	54.67	60.02	-	-	-
L257	8.848	128.8	53.09	54.87	45.26	-	-
T258	9.603	123.2	61.44	53.12	-	-	-
L259	9.266	127.0	53.12	61.59	-	-	-
S260	8.595	116.3	56.30	53.10	67.56	-	-
A261	9.222	123.4	50.92	56.24	-	-	-
L262	8.848	124.7	52.74	50.88	42.71	-	173.7

B.3 Chemical shift assignment of Quintuple hVDAC1

	Hi	Ni	C <sub>αi</sub>	C <sub>αi-1</sub>	C <sub>βi</sub>	C <sub>βi-1</sub>	C' <sub>i-1</sub> *
L263	9.563	126.2	52.40	52.80	43.14	-	175.2
D264	9.946	124.8	52.36	52.36	39.57	-	-
G265	8.445	112.3	46.03	52.38	-	40.10	176.7
K266	8.131	117.5	56.83	46.03	31.35	-	173.7
N267	7.431	117.6	51.82	56.72	38.20	-	177.0
V268	8.480	121.8	65.06	51.83	30.84	-	175.1
N269	8.320	117.0	54.30	-	-	-	-
A270	7.532	120.8	51.69	54.53	-	-	175.6
G271	7.700	106.9	44.83	51.89	-	-	-
G272	8.524	109.2	45.65	44.94	-	-	174.1
H273	7.197	120.8	54.54	45.58	31.34	-	174.0
K274	8.940	119.3	54.61	54.61	37.12	-	-
L275	8.673	121.4	53.40	54.54	45.75	-	175.1
G276	8.866	111.0	45.68	53.55	-	-	-
L277	8.254	120.7	53.70	45.65	-	-	-
G278	9.494	111.9	43.85	53.71	-	-	-
L279	8.761	124.3	53.45	43.83	-	-	-
E280	8.775	125.6	54.07	53.38	-	-	174.7
F281	9.068	125.8	-	54.11	-	-	-
Q282	-	-	-	-	-	-	-
A283	8.243	126.6	51.32	-	30.36	-	174.4
L284	-	-	-	-	-	-	-
E285	7.759	125.9	57.17	55.80	30.40	-	174.1



## B.4 Chemical shift assignment of hVDAC1 in liposomes

**Table B.4:** Chemical shifts of hVDAC1 in liposomes

	N	C'	C <sub>α</sub>	C <sub>β</sub>	C <sub>γ</sub>
	A2		52.164*	18.995*	
	V3		59.906*	31.682*	
	P4		61.536*	30.622*	
	P5	174.244#	61.352*	31.988*	27.093*
T6	106.236 <sup>+</sup>	176.106#	60.473*	69.720*	
Y7	122.926#		62.248*	40.038*	
A8	119.378 <sup>+</sup>		53.998*	18.596*	
D9		177.388*	53.883*	40.268*	
L10	125.543 <sup>+</sup>		56.447*	39.136*	
G11		175.187*	45.100*		
K12	122.932 <sup>+</sup>		59.807*	33.442*	
S13			59.976*	63.294*	
A14	122.001 <sup>+</sup>		55.286*	18.233*	
R15	116.568 <sup>+</sup>		60.302*	30.483*	28.04*
D16	118.244 <sup>+</sup>		57.303*	40.349*	
V17			66.58*	31.286*	
T19			66.289*	68.507*	
K32	126.050 <sup>+</sup>		55.162 <sup>+</sup>	36.847 <sup>+</sup>	
T42	120.948 <sup>+</sup>		61.712 <sup>+</sup>	71.390 <sup>+</sup>	
S46	116.408 <sup>+</sup>		56.798*	66.116*	
T55	118.003 <sup>+</sup>		59.973*	71.825*	
T72	120.761 <sup>+</sup>		61.724 <sup>+</sup>	71.327 <sup>+</sup>	
N76	122.297 <sup>+</sup>		51.999 <sup>+</sup>	39.418 <sup>+</sup>	
L81	127.813 <sup>+</sup>		53.823 <sup>+</sup>	45.911 <sup>+</sup>	
T86	123.304 <sup>+</sup>		61.008*	71.754*	
P105			65.367*	31.072*	
G108		174.616*	45.528*		
K109	119.963#				
A112	125.073 <sup>+</sup>	174.833#	51.987*	23.008*	
K113	121.372#				
K115	126.495 <sup>+</sup>	175.830#	54.875 <sup>+</sup>	33.907 <sup>+</sup>	
T116	116.602#				
I123	117.851 <sup>+</sup>		59.694*	42.49*	
S137	117.726 <sup>+</sup>		57.607 <sup>+</sup>	67.534 <sup>+</sup>	
I138		173.629#	59.861#		
R139	128.035#	175.623#	54.103#		
G140	112.480#	171.271#	44.849#		
A141	124.369 <sup>+</sup>		51.100 <sup>+</sup>	23.711 <sup>+</sup>	
L142			53.842*	47.717*	

	N	C'	C <sub>α</sub>	C <sub>β</sub>	C <sub>γ</sub>
V143		174.819 <sup>#</sup>	61.186*	35.703*	23.553* & 22.251*
L144	126.074 <sup>#</sup>	176.217 <sup>#</sup>	52.113*	46.328*	
G145	105.414 <sup>#</sup>				
A151		175.212 <sup>#</sup>	51.202*	24.078*	
G152	107.812 <sup>#</sup>				
S167	118.904 <sup>+</sup>		57.131 <sup>+</sup>	66.071 <sup>+</sup>	
F169	118.472 <sup>+</sup>	174.427 <sup>#</sup>	56.518*	42.418*	
A170	121.901 <sup>+</sup>	176.555 <sup>#</sup>	50.916*	22.615*	
V171	117.409 <sup>+</sup>	175.594 <sup>#</sup>	60.456 <sup>+</sup>	36.150 <sup>+</sup>	
G172	113.565 <sup>#</sup>				
Q179	122.615 <sup>+</sup>		53.955 <sup>+</sup>	32.274 <sup>+</sup>	
Q196	125.772 <sup>+</sup>		54.046 <sup>+</sup>	32.402 <sup>+</sup>	
S193			56.353*	67.534*	
E203		174.809 <sup>#</sup>	54.529 <sup>#</sup>		
T204	112.136 <sup>+</sup>	172.117 <sup>#</sup>	59.688*	72.539*	
A205	122.775 <sup>+</sup>	174.588 <sup>#</sup>	50.916*	23.971*	
V206	117.073 <sup>+</sup>		59.804 <sup>+</sup>	35.868 <sup>+</sup>	
W210	121.628 <sup>+</sup>		58.041 <sup>+</sup>	30.264 <sup>+</sup>	
A212	127.751 <sup>+</sup>		51.805 <sup>+</sup>	23.079 <sup>+</sup>	
A222	125.687 <sup>+</sup>	175.149 <sup>#</sup>	50.711 <sup>+</sup>	22.937 <sup>+</sup>	
A223	119.402 <sup>+</sup>	175.743 <sup>#</sup>	50.595*	23.936*	
K224	120.766 <sup>#</sup>				
Q226	130.995 <sup>+</sup>		54.306*	27.968*	
A231	124.160 <sup>+</sup>		51.855 <sup>+</sup>	22.870 <sup>+</sup>	
A235	122.972 <sup>+</sup>	175.761 <sup>#</sup>	51.100 <sup>+</sup>	23.985 <sup>+</sup>	
K236	114.518 <sup>#</sup>				
V237	117.834 <sup>+</sup>	173.995 <sup>#</sup>	59.801 <sup>+</sup>	35.869 <sup>+</sup>	
N238	122.859 <sup>#</sup>				
S240	117.716 <sup>+</sup>		58.867*	62.976*	
L245	121.688 <sup>+</sup>		53.894 <sup>+</sup>	46.875 <sup>+</sup>	
P253			65.919*	32.086*	
L257	127.823 <sup>+</sup>		53.451 <sup>+</sup>	45.928 <sup>+</sup>	
T258	123.395 <sup>+</sup>		61.686*	71.112*	

<sup>\*</sup> = from PDSD<sup>+</sup> = from NCACB<sup>#</sup> = from NCOCA

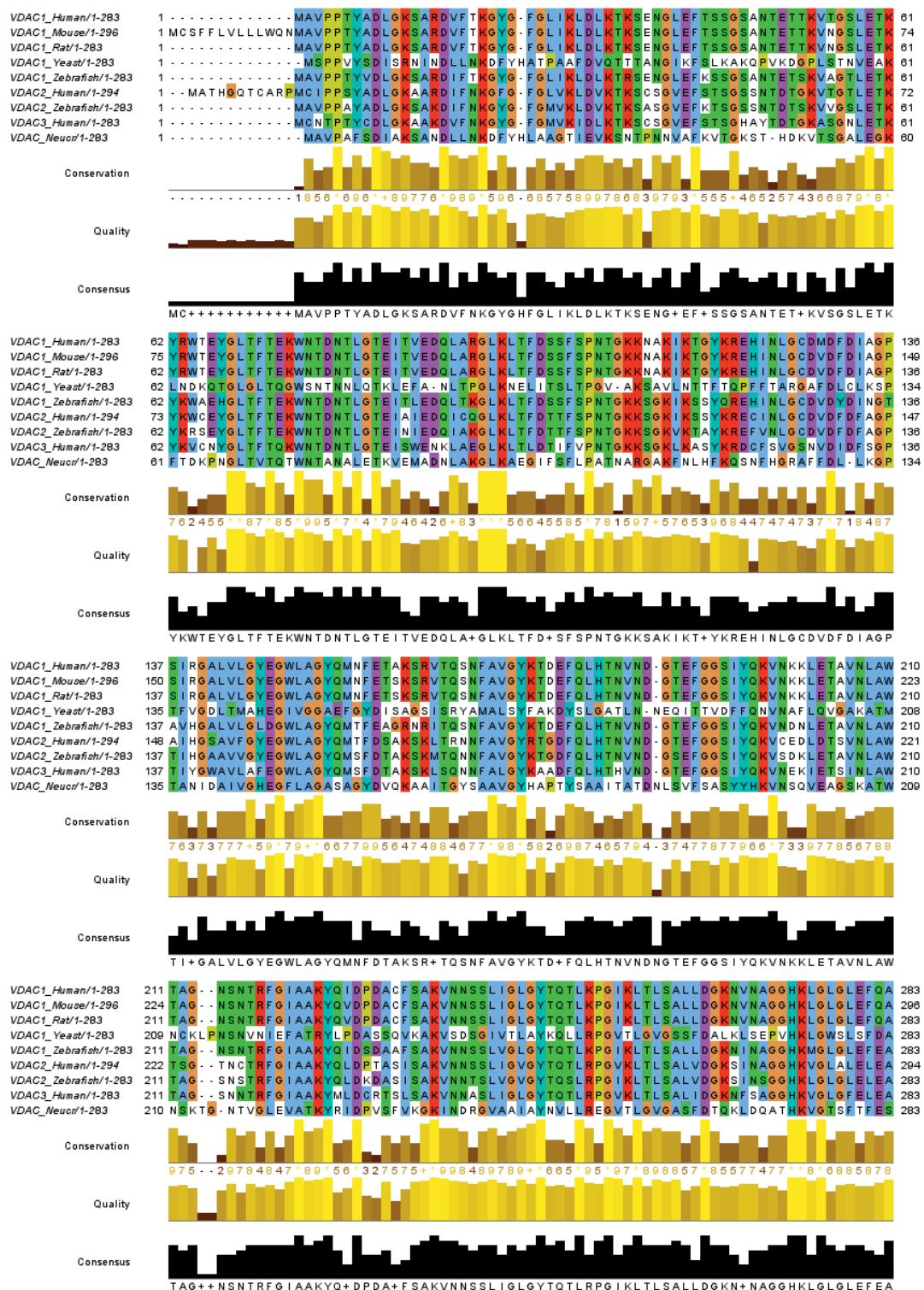


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## Appendix C Sequence alignment of mentioned VDAC isoforms

The sequences of VDAC isoforms that were mentioned in this thesis (and for completeness also including zfVDAC1) were aligned with Clustal Omega<sup>[152]</sup> and visualized with Jalview 2.10.1<sup>[161]</sup>. The figure is – due to space restrictions – following on the next page.

## Appendix C Sequence Alignment of mentioned VDAC isoforms



**Figure C.1: Alignment of VDAC sequences.** VDAC1 from *homo sapiens* (VDAC1\_Human), VDAC1 from *mus musculus* (VDAC1\_Mouse), VDAC1 from *rattus norvegicus* (VDAC1\_Rat), VDAC1 from *saccharomyces cerevisiae* (VDAC1\_Yeast), VDAC1 from *danio rerio* (VDAC1\_Zebrafish), VDAC2 from *homo sapiens* (VDAC2\_Human), VDAC2 from *danio rerio* (VDAC2\_Zebrafish), VDAC3 from *homo sapiens* (VDAC3\_Human) and VDAC from *neurospora crassa* (VDAC\_Neucr) were aligned with Clustal Omega<sup>[152]</sup> and visualized with Jalview 2.10.1<sup>[161]</sup>.

# Appendix D PDB files of the models for the closed state of VDAC

In the following, the PDB files for the two models of the closed state of VDAC are given. Due to space restrictions, only the coordinates of the N, C<sub>α</sub>, C' and O atoms are printed.

## D.1 Model 1

```

HEADER 5jdp_close1c
ATOM    1  N   SER A  1   -14.364 12.935 18.289 1.00  0.00      N
ATOM    2  CA  SER A  1   -13.382 12.614 19.359 1.00  0.00      C
ATOM    3  C   SER A  1   -12.511 11.470 18.871 1.00  0.00      C
ATOM    4  O   SER A  1   -11.378 11.296 19.322 1.00  0.00      O
ATOM   14  N   ALA A  2   -13.035 10.720 17.906 1.00  0.00      N
ATOM   15  CA  ALA A  2   -12.280 9.632 17.327 1.00  0.00      C
ATOM   16  C   ALA A  2   -11.054 10.245 16.708 1.00  0.00      C
ATOM   17  O   ALA A  2   -9.969 9.672 16.746 1.00  0.00      O
ATOM   24  N   VAL A  3   -11.246 11.482 16.142 1.00  0.00      N
ATOM   25  CA  VAL A  3   -10.246 12.451 15.751 1.00  0.00      C
ATOM   26  C   VAL A  3   -9.285 11.629 14.953 1.00  0.00      C
ATOM   27  O   VAL A  3   -8.195 11.578 15.345 1.00  0.00      O
ATOM   40  N   PRO A  4   -9.733 10.754 14.006 1.00  0.00      N
ATOM   41  CA  PRO A  4   -8.708 9.843 13.446 1.00  0.00      C
ATOM   42  C   PRO A  4   -8.119 10.817 12.434 1.00  0.00      C
ATOM   43  O   PRO A  4   -8.676 11.217 11.434 1.00  0.00      O
ATOM   68  N   THR A  6   -6.338 10.524 9.580 1.00  0.00      N
ATOM   69  CA  THR A  6   -6.698 9.087 9.832 1.00  0.00      C
ATOM   70  C   THR A  6   -5.339 8.387 9.774 1.00  0.00      C
ATOM   71  O   THR A  6   -4.786 8.801 8.794 1.00  0.00      O
ATOM   82  N   TYR A  7   -4.919 7.511 10.625 1.00  0.00      N
ATOM   83  CA  TYR A  7   -3.508 7.048 10.816 1.00  0.00      C
ATOM   84  C   TYR A  7   -2.748 6.624 9.522 1.00  0.00      C
ATOM   85  O   TYR A  7   -1.538 6.836 9.500 1.00  0.00      O
ATOM  103  N   ALA A  8   -3.421 6.151 8.483 1.00  0.00      N
ATOM  104  CA  ALA A  8   -2.824 6.080 7.126 1.00  0.00      C
ATOM  105  C   ALA A  8   -2.643 7.473 6.461 1.00  0.00      C
ATOM  106  O   ALA A  8   -1.539 7.729 5.967 1.00  0.00      O
ATOM  113  N   ASP A  9   -3.662 8.293 6.473 1.00  0.00      N
ATOM  114  CA  ASP A  9   -3.627 9.719 5.994 1.00  0.00      C
ATOM  115  C   ASP A  9   -3.067 10.745 7.042 1.00  0.00      C
ATOM  116  O   ASP A  9   -3.452 11.957 6.905 1.00  0.00      O
ATOM  125  N   LEU A 10  -2.172 10.278 7.872 1.00  0.00      N
ATOM  126  CA  LEU A 10  -0.914 11.326 9.072 1.00  0.00      C
ATOM  127  C   LEU A 10  -0.192 12.645 8.838 1.00  0.00      C
ATOM  128  O   LEU A 10  0.962 12.879 9.158 1.00  0.00      O
ATOM  144  N   GLY A 11  -0.981 13.515 8.164 1.00  0.00      N
ATOM  145  CA  GLY A 11  -0.479 14.706 7.492 1.00  0.00      C
ATOM  146  C   GLY A 11  0.171 14.327 6.158 1.00  0.00      C
ATOM  147  O   GLY A 11  1.338 13.881 6.267 1.00  0.00      O
ATOM  151  N   LYS A 12  -0.551 14.453 5.006 1.00  0.00      N
ATOM  152  CA  LYS A 12  -0.341 13.553 3.898 1.00  0.00      C
ATOM  153  C   LYS A 12  0.328 14.278 2.704 1.00  0.00      C
ATOM  154  O   LYS A 12  1.107 13.480 2.207 1.00  0.00      O
ATOM  173  N   SER A 13  -0.006 15.493 2.046 1.00  0.00      N

```



## Appendix D PDB files of the models for the closed state of VDAC

---

ATOM	174	CA	SER	A	13	0.805	16.230	1.059	1.00	0.00	C
ATOM	175	C	SER	A	13	2.295	16.178	1.388	1.00	0.00	C
ATOM	176	O	SER	A	13	3.112	15.906	0.507	1.00	0.00	O
ATOM	184	N	ALA	A	14	2.720	15.961	2.668	1.00	0.00	N
ATOM	185	CA	ALA	A	14	4.006	15.350	3.089	1.00	0.00	C
ATOM	186	C	ALA	A	14	4.201	13.881	3.046	1.00	0.00	C
ATOM	187	O	ALA	A	14	5.462	13.624	2.774	1.00	0.00	O
ATOM	194	N	ARG	A	15	3.401	12.885	3.356	1.00	0.00	N
ATOM	195	CA	ARG	A	15	3.675	11.441	3.141	1.00	0.00	C
ATOM	196	C	ARG	A	15	3.561	11.102	1.670	1.00	0.00	C
ATOM	197	O	ARG	A	15	4.594	10.514	1.223	1.00	0.00	O
ATOM	218	N	ASP	A	16	2.564	11.327	0.852	1.00	0.00	N
ATOM	219	CA	ASP	A	16	2.511	10.713	-0.505	1.00	0.00	C
ATOM	220	C	ASP	A	16	3.436	11.440	-1.522	1.00	0.00	C
ATOM	221	O	ASP	A	16	3.246	11.059	-2.713	1.00	0.00	O
ATOM	230	N	VAL	A	17	4.412	12.288	-1.175	1.00	0.00	N
ATOM	231	CA	VAL	A	17	5.665	12.460	-1.962	1.00	0.00	C
ATOM	232	C	VAL	A	17	6.855	11.931	-1.111	1.00	0.00	C
ATOM	233	O	VAL	A	17	7.776	11.424	-1.843	1.00	0.00	O
ATOM	246	N	PHE	A	18	7.001	11.933	0.194	1.00	0.00	N
ATOM	247	CA	PHE	A	18	8.004	11.132	0.980	1.00	0.00	C
ATOM	248	C	PHE	A	18	8.011	9.610	0.654	1.00	0.00	C
ATOM	249	O	PHE	A	18	9.103	9.033	0.755	1.00	0.00	O
ATOM	266	N	THR	A	19	6.892	9.052	0.195	1.00	0.00	N
ATOM	267	CA	THR	A	19	6.846	7.851	-0.662	1.00	0.00	C
ATOM	268	C	THR	A	19	6.692	8.245	-2.147	1.00	0.00	C
ATOM	269	O	THR	A	19	5.653	7.848	-2.733	1.00	0.00	O
ATOM	280	N	LYS	A	20	7.664	8.948	-2.736	1.00	0.00	N
ATOM	281	CA	LYS	A	20	7.858	8.920	-4.173	1.00	0.00	C
ATOM	282	C	LYS	A	20	9.370	8.734	-4.532	1.00	0.00	C
ATOM	283	O	LYS	A	20	10.101	9.713	-5.161	1.00	0.00	O
ATOM	302	N	GLY	A	21	9.692	7.196	-4.225	1.00	0.00	N
ATOM	303	CA	GLY	A	21	10.538	6.404	-5.261	1.00	0.00	C
ATOM	304	C	GLY	A	21	10.900	6.726	-3.925	1.00	0.00	C
ATOM	305	O	GLY	A	21	10.796	7.849	-2.757	1.00	0.00	O
ATOM	309	N	TYR	A	22	11.792	5.344	-3.756	1.00	0.00	N
ATOM	310	CA	TYR	A	22	11.018	4.066	-3.460	1.00	0.00	C
ATOM	311	C	TYR	A	22	10.610	3.896	-2.028	1.00	0.00	C
ATOM	312	O	TYR	A	22	9.375	3.504	-1.956	1.00	0.00	O
ATOM	330	N	GLY	A	23	11.277	4.096	-0.925	1.00	0.00	N
ATOM	331	CA	GLY	A	23	10.618	3.943	0.391	1.00	0.00	C
ATOM	332	C	GLY	A	23	11.512	4.381	1.560	1.00	0.00	C
ATOM	333	O	GLY	A	23	12.660	3.898	1.541	1.00	0.00	O
ATOM	337	N	PHE	A	24	11.024	5.208	2.469	1.00	0.00	N
ATOM	338	CA	PHE	A	24	11.687	5.640	3.729	1.00	0.00	C
ATOM	339	C	PHE	A	24	10.838	5.329	4.963	1.00	0.00	C
ATOM	340	O	PHE	A	24	9.661	4.961	4.792	1.00	0.00	O
ATOM	357	N	GLY	A	25	11.360	5.464	6.194	1.00	0.00	N
ATOM	358	CA	GLY	A	25	10.497	5.273	7.358	1.00	0.00	C
ATOM	359	C	GLY	A	25	11.142	5.449	8.734	1.00	0.00	C
ATOM	360	O	GLY	A	25	11.423	6.586	9.133	1.00	0.00	O
ATOM	364	N	LEU	A	26	11.337	4.376	9.499	1.00	0.00	N
ATOM	365	CA	LEU	A	26	11.549	4.437	10.965	1.00	0.00	C
ATOM	366	C	LEU	A	26	13.031	4.421	11.326	1.00	0.00	C
ATOM	367	O	LEU	A	26	13.767	4.834	10.333	1.00	0.00	O
ATOM	383	N	ILE	A	27	13.700	4.059	12.425	1.00	0.00	N
ATOM	384	CA	ILE	A	27	15.140	3.722	12.459	1.00	0.00	C
ATOM	385	C	ILE	A	27	15.102	2.375	11.698	1.00	0.00	C
ATOM	386	O	ILE	A	27	14.864	1.430	12.374	1.00	0.00	O
ATOM	402	N	LYS	A	28	15.300	2.436	10.392	1.00	0.00	N
ATOM	403	CA	LYS	A	28	15.102	1.346	9.426	1.00	0.00	C
ATOM	404	C	LYS	A	28	16.442	0.930	8.812	1.00	0.00	C
ATOM	405	O	LYS	A	28	16.976	1.919	8.229	1.00	0.00	O
ATOM	424	N	LEU	A	29	16.887	-0.330	8.942	1.00	0.00	N
ATOM	425	CA	LEU	A	29	18.105	-0.820	8.307	1.00	0.00	C
ATOM	426	C	LEU	A	29	17.664	-1.567	7.045	1.00	0.00	C
ATOM	427	O	LEU	A	29	16.868	-2.499	7.308	1.00	0.00	O
ATOM	443	N	ASP	A	30	18.081	-1.162	5.831	1.00	0.00	N
ATOM	444	CA	ASP	A	30	17.722	-1.827	4.577	1.00	0.00	C
ATOM	445	C	ASP	A	30	18.957	-2.329	3.841	1.00	0.00	C
ATOM	446	O	ASP	A	30	19.923	-1.590	3.639	1.00	0.00	O
ATOM	455	N	LEU	A	31	18.905	-3.598	3.442	1.00	0.00	N
ATOM	456	CA	LEU	A	31	20.002	-4.237	2.720	1.00	0.00	C
ATOM	457	C	LEU	A	31	19.539	-4.554	1.294	1.00	0.00	C
ATOM	458	O	LEU	A	31	18.509	-5.217	1.117	1.00	0.00	O
ATOM	474	N	LYS	A	32	20.244	-4.052	0.271	1.00	0.00	N
ATOM	475	CA	LYS	A	32	20.037	-4.365	-1.166	1.00	0.00	C
ATOM	476	C	LYS	A	32	21.195	-5.208	-1.686	1.00	0.00	C
ATOM	477	O	LYS	A	32	22.322	-5.085	-1.222	1.00	0.00	O
ATOM	496	N	THR	A	33	20.137	-6.301	-2.446	1.00	0.00	N
ATOM	497	CA	THR	A	33	20.856	-7.299	-3.242	1.00	0.00	C
ATOM	498	C	THR	A	33	20.336	-7.259	-4.682	1.00	0.00	C
ATOM	499	O	THR	A	33	19.157	-7.538	-4.943	1.00	0.00	O
ATOM	510	N	LYS	A	34	21.202	-6.890	-5.613	1.00	0.00	N
ATOM	511	CA	LYS	A	34	20.783	-6.792	-7.003	1.00	0.00	C
ATOM	512	C	LYS	A	34	20.787	-8.141	-7.700	1.00	0.00	C
ATOM	513	O	LYS	A	34	21.833	-8.770	-7.853	1.00	0.00	O
ATOM	532	N	SER	A	35	19.611	-8.544	-8.175	1.00	0.00	N
ATOM	533	CA	SER	A	35	19.497	-9.784	-8.919	1.00	0.00	C
ATOM	534	C	SER	A	35	19.856	-9.500	-10.372	1.00	0.00	C
ATOM	535	O	SER	A	35	19.687	-8.379	-10.848	1.00	0.00	O
ATOM	543	N	GLU	A	36	20.354	-10.499	-11.081	1.00	0.00	N
ATOM	544	CA	GLU	A	36	20.718	-10.287	-12.468	1.00	0.00	C
ATOM	545	C	GLU	A	36	19.486	-10.408	-13.361	1.00	0.00	C

ATOM	546	O	GLU	A	36	19.375	-9.715	-14.371	1.00	0.00	O
ATOM	558	N	ASN	A	37	18.538	-11.259	-12.959	1.00	0.00	N
ATOM	559	CA	ASN	A	37	17.303	-11.410	-13.721	1.00	0.00	C
ATOM	560	C	ASN	A	37	16.525	-10.091	-13.693	1.00	0.00	C
ATOM	561	O	ASN	A	37	15.965	-9.659	-14.703	1.00	0.00	O
ATOM	572	N	GLY	A	38	16.501	-9.463	-12.519	1.00	0.00	N
ATOM	573	CA	GLY	A	38	15.805	-8.204	-12.341	1.00	0.00	C
ATOM	574	C	GLY	A	38	14.847	-8.283	-11.162	1.00	0.00	C
ATOM	575	O	GLY	A	38	13.966	-7.442	-11.043	1.00	0.00	O
ATOM	579	N	LEU	A	39	15.022	-9.307	-10.315	1.00	0.00	N
ATOM	580	CA	LEU	A	39	14.155	-9.514	-9.152	1.00	0.00	C
ATOM	581	C	LEU	A	39	14.271	-8.372	-8.147	1.00	0.00	C
ATOM	582	O	LEU	A	39	13.277	-7.945	-7.565	1.00	0.00	O
ATOM	598	N	GLU	A	40	15.491	-7.891	-7.944	1.00	0.00	N
ATOM	599	CA	GLU	A	40	15.736	-6.791	-7.012	1.00	0.00	C
ATOM	600	C	GLU	A	40	15.237	-7.132	-5.605	1.00	0.00	C
ATOM	601	O	GLU	A	40	14.173	-6.663	-5.204	1.00	0.00	O
ATOM	613	N	PHE	A	41	16.008	-7.911	-4.845	1.00	0.00	N
ATOM	614	CA	PHE	A	41	15.609	-8.250	-3.478	1.00	0.00	C
ATOM	615	C	PHE	A	41	16.264	-7.355	-2.450	1.00	0.00	C
ATOM	616	O	PHE	A	41	17.471	-7.412	-2.225	1.00	0.00	O
ATOM	633	N	THR	A	42	15.424	-6.563	-1.796	1.00	0.00	N
ATOM	634	CA	THR	A	42	15.877	-5.676	-0.737	1.00	0.00	C
ATOM	635	C	THR	A	42	15.088	-5.958	0.519	1.00	0.00	C
ATOM	636	O	THR	A	42	13.912	-5.602	0.609	1.00	0.00	O
ATOM	647	N	SER	A	43	15.720	-6.589	1.490	1.00	0.00	N
ATOM	648	CA	SER	A	43	15.028	-6.884	2.729	1.00	0.00	C
ATOM	649	C	SER	A	43	15.353	-5.808	3.748	1.00	0.00	C
ATOM	650	O	SER	A	43	16.338	-5.084	3.601	1.00	0.00	O
ATOM	669	N	GLY	A	45	14.987	-4.409	8.149	1.00	0.00	N
ATOM	670	CA	GLY	A	45	14.796	-4.663	9.568	1.00	0.00	C
ATOM	671	C	GLY	A	45	14.814	-3.327	10.288	1.00	0.00	C
ATOM	672	O	GLY	A	45	15.755	-2.546	10.146	1.00	0.00	O
ATOM	676	N	SER	A	46	13.786	-2.952	11.038	1.00	0.00	N
ATOM	677	CA	SER	A	46	13.645	-1.601	11.631	1.00	0.00	C
ATOM	678	C	SER	A	46	13.457	-1.647	13.118	1.00	0.00	C
ATOM	679	O	SER	A	46	13.432	-2.690	13.677	1.00	0.00	O
ATOM	687	N	ALA	A	47	13.324	-0.451	13.682	1.00	0.00	N
ATOM	688	CA	ALA	A	47	12.782	-0.283	14.998	1.00	0.00	C
ATOM	689	C	ALA	A	47	12.286	1.140	15.244	1.00	0.00	C
ATOM	690	O	ALA	A	47	12.754	2.069	14.588	1.00	0.00	O
ATOM	697	N	ASN	A	48	11.361	1.236	16.164	1.00	0.00	N
ATOM	698	CA	ASN	A	48	10.609	2.450	16.469	1.00	0.00	C
ATOM	699	C	ASN	A	48	11.291	3.068	17.698	1.00	0.00	C
ATOM	700	O	ASN	A	48	11.594	2.322	18.637	1.00	0.00	O
ATOM	711	N	THR	A	49	11.513	4.374	17.707	1.00	0.00	N
ATOM	712	CA	THR	A	49	12.094	5.041	18.884	1.00	0.00	C
ATOM	713	C	THR	A	49	11.155	5.040	20.099	1.00	0.00	C
ATOM	714	O	THR	A	49	11.757	5.016	21.197	1.00	0.00	O
ATOM	725	N	GLU	A	50	9.840	5.078	19.980	1.00	0.00	N
ATOM	726	CA	GLU	A	50	8.980	5.021	21.170	1.00	0.00	C
ATOM	727	C	GLU	A	50	9.033	3.622	21.797	1.00	0.00	C
ATOM	728	O	GLU	A	50	9.510	3.554	22.947	1.00	0.00	O
ATOM	740	N	THR	A	51	8.584	2.592	21.078	1.00	0.00	N
ATOM	741	CA	THR	A	51	8.203	1.320	21.705	1.00	0.00	C
ATOM	742	C	THR	A	51	9.335	0.290	21.744	1.00	0.00	C
ATOM	743	O	THR	A	51	9.236	-0.698	22.469	1.00	0.00	O
ATOM	768	N	LYS	A	53	10.025	-1.751	19.449	1.00	0.00	N
ATOM	769	CA	LYS	A	53	9.562	-2.900	18.615	1.00	0.00	C
ATOM	770	C	LYS	A	53	10.487	-3.066	17.417	1.00	0.00	C
ATOM	771	O	LYS	A	53	11.299	-2.202	17.174	1.00	0.00	O
ATOM	790	N	VAL	A	54	10.318	-4.205	16.720	1.00	0.00	N
ATOM	791	CA	VAL	A	54	11.083	-4.624	15.566	1.00	0.00	C
ATOM	792	C	VAL	A	54	10.076	-4.999	14.440	1.00	0.00	C
ATOM	793	O	VAL	A	54	9.541	-6.114	14.523	1.00	0.00	O
ATOM	806	N	THR	A	55	9.879	-4.126	13.468	1.00	0.00	N
ATOM	807	CA	THR	A	55	9.179	-4.407	12.182	1.00	0.00	C
ATOM	808	C	THR	A	55	10.222	-4.552	11.078	1.00	0.00	C
ATOM	809	O	THR	A	55	11.385	-4.234	11.258	1.00	0.00	O
ATOM	820	N	GLY	A	56	9.799	-5.060	9.929	1.00	0.00	N
ATOM	821	CA	GLY	A	56	10.638	-5.110	8.725	1.00	0.00	C
ATOM	822	C	GLY	A	56	9.790	-5.256	7.470	1.00	0.00	C
ATOM	823	O	GLY	A	56	8.567	-5.053	7.540	1.00	0.00	O
ATOM	827	N	SER	A	57	10.433	-5.606	6.377	1.00	0.00	N
ATOM	828	CA	SER	A	57	9.766	-5.690	5.077	1.00	0.00	C
ATOM	829	C	SER	A	57	10.641	-6.441	4.076	1.00	0.00	C
ATOM	830	O	SER	A	57	11.850	-6.578	4.333	1.00	0.00	O
ATOM	838	N	LEU	A	58	9.999	-6.897	3.013	1.00	0.00	N
ATOM	839	CA	LEU	A	58	10.553	-7.605	1.877	1.00	0.00	C
ATOM	840	C	LEU	A	58	10.174	-6.880	0.595	1.00	0.00	C
ATOM	841	O	LEU	A	58	9.018	-6.948	0.167	1.00	0.00	O
ATOM	857	N	GLU	A	59	11.079	-6.189	-0.084	1.00	0.00	N
ATOM	858	CA	GLU	A	59	10.819	-5.321	-1.251	1.00	0.00	C
ATOM	859	C	GLU	A	59	11.427	-6.015	-2.465	1.00	0.00	C
ATOM	860	O	GLU	A	59	12.675	-5.938	-2.516	1.00	0.00	O
ATOM	872	N	THR	A	60	10.655	-6.736	-3.321	1.00	0.00	N
ATOM	873	CA	THR	A	60	11.098	-7.532	-4.463	1.00	0.00	C
ATOM	874	C	THR	A	60	10.548	-6.957	-5.763	1.00	0.00	C
ATOM	875	O	THR	A	60	9.336	-6.976	-5.995	1.00	0.00	O
ATOM	886	N	LYS	A	61	11.358	-6.575	-6.761	1.00	0.00	N
ATOM	887	CA	LYS	A	61	10.948	-5.697	-7.876	1.00	0.00	C
ATOM	888	C	LYS	A	61	11.752	-6.040	-9.135	1.00	0.00	C
ATOM	889	O	LYS	A	61	12.454	-7.041	-9.203	1.00	0.00	O
ATOM	908	N	TYR	A	62	10.516	-7.160	-9.805	1.00	0.00	N

## Appendix D PDB files of the models for the closed state of VDAC

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ATOM	909	CA	TYR A	62	10.885	-7.913	-11.012	1.00	0.00	C
ATOM	910	C	TYR A	62	10.745	-7.022	-12.247	1.00	0.00	C
ATOM	911	O	TYR A	62	9.680	-6.425	-12.453	1.00	0.00	O
ATOM	929	N	ARG A	63	11.821	-6.947	-13.059	1.00	0.00	N
ATOM	930	CA	ARG A	63	11.816	-6.138	-14.300	1.00	0.00	C
ATOM	931	C	ARG A	63	12.026	-4.641	-14.027	1.00	0.00	C
ATOM	932	O	ARG A	63	13.099	-4.269	-13.579	1.00	0.00	O
ATOM	953	N	TRP A	64	10.981	-3.821	-14.286	1.00	0.00	N
ATOM	954	CA	TRP A	64	10.996	-2.358	-14.062	1.00	0.00	C
ATOM	955	C	TRP A	64	10.096	-1.711	-15.115	1.00	0.00	C
ATOM	956	O	TRP A	64	9.313	-2.399	-15.768	1.00	0.00	O
ATOM	977	N	THR A	65	10.194	-0.393	-15.270	1.00	0.00	N
ATOM	978	CA	THR A	65	9.369	0.308	-16.242	1.00	0.00	C
ATOM	979	C	THR A	65	9.944	0.183	-17.650	1.00	0.00	C
ATOM	980	O	THR A	65	9.561	0.939	-18.539	1.00	0.00	O
ATOM	991	N	GLU A	66	10.858	-0.775	-17.858	1.00	0.00	N
ATOM	992	CA	GLU A	66	11.448	-0.968	-19.185	1.00	0.00	C
ATOM	993	C	GLU A	66	10.358	-0.953	-20.244	1.00	0.00	C
ATOM	994	O	GLU A	66	9.932	0.116	-20.688	1.00	0.00	O
ATOM	1006	N	TYR A	67	9.900	-2.134	-20.664	1.00	0.00	N
ATOM	1007	CA	TYR A	67	8.863	-2.207	-21.667	1.00	0.00	C
ATOM	1008	C	TYR A	67	7.491	-2.169	-20.999	1.00	0.00	C
ATOM	1009	O	TYR A	67	6.502	-2.640	-21.561	1.00	0.00	O
ATOM	1027	N	GLY A	68	7.441	-1.541	-19.819	1.00	0.00	N
ATOM	1028	CA	GLY A	68	6.181	-1.370	-19.103	1.00	0.00	C
ATOM	1029	C	GLY A	68	5.722	-2.639	-18.396	1.00	0.00	C
ATOM	1030	O	GLY A	68	4.555	-2.740	-18.028	1.00	0.00	O
ATOM	1034	N	LEU A	69	6.632	-3.601	-18.218	1.00	0.00	N
ATOM	1035	CA	LEU A	69	6.289	-4.856	-17.554	1.00	0.00	C
ATOM	1036	C	LEU A	69	6.868	-4.911	-16.149	1.00	0.00	C
ATOM	1037	O	LEU A	69	7.736	-5.732	-15.857	1.00	0.00	O
ATOM	1053	N	THR A	70	6.351	-4.055	-15.282	1.00	0.00	N
ATOM	1054	CA	THR A	70	6.804	-4.032	-13.896	1.00	0.00	C
ATOM	1055	C	THR A	70	5.946	-4.942	-13.042	1.00	0.00	C
ATOM	1056	O	THR A	70	4.762	-4.655	-12.831	1.00	0.00	O
ATOM	1067	N	PHE A	71	6.541	-6.023	-12.525	1.00	0.00	N
ATOM	1068	CA	PHE A	71	5.800	-6.919	-11.655	1.00	0.00	C
ATOM	1069	C	PHE A	71	6.374	-6.745	-10.251	1.00	0.00	C
ATOM	1070	O	PHE A	71	7.412	-7.309	-9.904	1.00	0.00	O
ATOM	1087	N	THR A	72	5.711	-5.910	-9.473	1.00	0.00	N
ATOM	1088	CA	THR A	72	6.176	-5.596	-8.126	1.00	0.00	C
ATOM	1089	C	THR A	72	5.631	-6.586	-7.113	1.00	0.00	C
ATOM	1090	O	THR A	72	4.460	-6.947	-7.154	1.00	0.00	O
ATOM	1101	N	VAL A	73	6.496	-6.991	-6.181	1.00	0.00	N
ATOM	1102	CA	VAL A	73	6.092	-7.912	-5.130	1.00	0.00	C
ATOM	1103	C	VAL A	73	6.496	-7.371	-3.757	1.00	0.00	C
ATOM	1104	O	VAL A	73	7.673	-7.134	-3.485	1.00	0.00	O
ATOM	1117	N	LYS A	74	5.498	-7.170	-2.902	1.00	0.00	N
ATOM	1118	CA	LYS A	74	5.724	-6.649	-1.557	1.00	0.00	C
ATOM	1119	C	LYS A	74	5.228	-7.626	-0.494	1.00	0.00	C
ATOM	1120	O	LYS A	74	4.025	-7.842	-0.358	1.00	0.00	O
ATOM	1139	N	TRP A	75	6.154	-8.181	0.275	1.00	0.00	N
ATOM	1140	CA	TRP A	75	5.798	-9.101	1.353	1.00	0.00	C
ATOM	1141	C	TRP A	75	6.213	-8.478	2.690	1.00	0.00	C
ATOM	1142	O	TRP A	75	7.258	-7.831	2.778	1.00	0.00	O
ATOM	1163	N	ASN A	76	5.458	-8.662	3.785	1.00	0.00	N
ATOM	1164	CA	ASN A	76	5.759	-8.127	5.145	1.00	0.00	C
ATOM	1165	C	ASN A	76	5.519	-9.172	6.240	1.00	0.00	C
ATOM	1166	O	ASN A	76	4.864	-10.191	6.044	1.00	0.00	O
ATOM	1177	N	THR A	77	6.076	-8.879	7.422	1.00	0.00	N
ATOM	1178	CA	THR A	77	6.041	-9.728	8.618	1.00	0.00	C
ATOM	1179	C	THR A	77	4.711	-9.747	9.368	1.00	0.00	C
ATOM	1180	O	THR A	77	4.326	-10.792	9.907	1.00	0.00	O
ATOM	1191	N	ASP A	78	3.878	-8.697	9.226	1.00	0.00	N
ATOM	1192	CA	ASP A	78	2.422	-8.821	9.369	1.00	0.00	C
ATOM	1193	C	ASP A	78	1.698	-9.755	8.387	1.00	0.00	C
ATOM	1194	O	ASP A	78	0.461	-9.693	8.598	1.00	0.00	O
ATOM	1203	N	ASN A	79	2.252	-10.492	7.388	1.00	0.00	N
ATOM	1204	CA	ASN A	79	1.687	-11.485	6.467	1.00	0.00	C
ATOM	1205	C	ASN A	79	1.082	-10.859	5.206	1.00	0.00	C
ATOM	1206	O	ASN A	79	0.648	-11.589	4.318	1.00	0.00	O
ATOM	1217	N	THR A	80	1.064	-9.531	5.096	1.00	0.00	N
ATOM	1218	CA	THR A	80	0.509	-8.915	3.883	1.00	0.00	C
ATOM	1219	C	THR A	80	1.360	-9.293	2.679	1.00	0.00	C
ATOM	1220	O	THR A	80	2.579	-9.106	2.688	1.00	0.00	O
ATOM	1231	N	LEU A	81	0.696	-9.780	1.632	1.00	0.00	N
ATOM	1232	CA	LEU A	81	1.378	-10.147	0.395	1.00	0.00	C
ATOM	1233	C	LEU A	81	0.797	-9.312	-0.740	1.00	0.00	C
ATOM	1234	O	LEU A	81	-0.424	-9.226	-0.853	1.00	0.00	O
ATOM	1250	N	GLY A	82	1.632	-8.689	-1.572	1.00	0.00	N
ATOM	1251	CA	GLY A	82	1.100	-7.874	-2.662	1.00	0.00	C
ATOM	1252	C	GLY A	82	1.882	-8.065	-3.954	1.00	0.00	C
ATOM	1253	O	GLY A	82	3.078	-7.830	-3.989	1.00	0.00	O
ATOM	1257	N	THR A	83	1.182	-8.441	-5.024	1.00	0.00	N
ATOM	1258	CA	THR A	83	1.818	-8.606	-6.334	1.00	0.00	C
ATOM	1259	C	THR A	83	1.081	-7.764	-7.376	1.00	0.00	C
ATOM	1260	O	THR A	83	-0.126	-7.921	-7.572	1.00	0.00	O
ATOM	1271	N	GLU A	84	1.805	-6.855	-8.023	1.00	0.00	N
ATOM	1272	CA	GLU A	84	1.195	-5.977	-9.028	1.00	0.00	C
ATOM	1273	C	GLU A	84	1.831	-6.154	-10.404	1.00	0.00	C
ATOM	1274	O	GLU A	84	2.965	-5.734	-10.612	1.00	0.00	O
ATOM	1286	N	ILE A	85	1.089	-6.722	-11.363	1.00	0.00	N
ATOM	1287	CA	ILE A	85	1.623	-6.878	-12.721	1.00	0.00	C
ATOM	1288	C	ILE A	85	1.031	-5.801	-13.621	1.00	0.00	C

ATOM	1289	O	ILE	A	85	-0.178	-5.758	-13.817	1.00	0.00	O
ATOM	1305	N	THR	A	86	1.893	-4.924	-14.138	1.00	0.00	N
ATOM	1306	CA	THR	A	86	1.435	-3.823	-14.987	1.00	0.00	C
ATOM	1307	C	THR	A	86	2.006	-3.871	-16.401	1.00	0.00	C
ATOM	1308	O	THR	A	86	3.218	-3.999	-16.582	1.00	0.00	O
ATOM	1319	N	VAL	A	87	1.124	-3.693	-17.397	1.00	0.00	N
ATOM	1320	CA	VAL	A	87	1.555	-3.637	-18.796	1.00	0.00	C
ATOM	1321	C	VAL	A	87	1.166	-2.281	-19.380	1.00	0.00	C
ATOM	1322	O	VAL	A	87	0.004	-2.055	-19.718	1.00	0.00	O
ATOM	1335	N	GLU	A	88	2.134	-1.383	-19.492	1.00	0.00	N
ATOM	1336	CA	GLU	A	88	1.869	-0.046	-20.032	1.00	0.00	C
ATOM	1337	C	GLU	A	88	2.183	0.025	-21.535	1.00	0.00	C
ATOM	1338	O	GLU	A	88	3.017	-0.731	-22.026	1.00	0.00	O
ATOM	1350	N	ASP	A	89	1.523	0.952	-22.247	1.00	0.00	N
ATOM	1351	CA	ASP	A	89	1.761	1.143	-23.690	1.00	0.00	C
ATOM	1352	C	ASP	A	89	1.411	-0.105	-24.504	1.00	0.00	C
ATOM	1353	O	ASP	A	89	2.148	-0.467	-25.422	1.00	0.00	O
ATOM	1362	N	GLN	A	90	0.422	-0.933	-24.148	1.00	0.00	N
ATOM	1363	CA	GLN	A	90	0.066	-2.096	-24.985	1.00	0.00	C
ATOM	1364	C	GLN	A	90	-0.423	-1.682	-26.380	1.00	0.00	C
ATOM	1365	O	GLN	A	90	0.192	-2.366	-27.254	1.00	0.00	O
ATOM	1379	N	LEU	A	91	-1.250	-0.668	-26.572	1.00	0.00	N
ATOM	1380	CA	LEU	A	91	-1.692	-0.366	-27.935	1.00	0.00	C
ATOM	1381	C	LEU	A	91	-0.657	0.532	-28.609	1.00	0.00	C
ATOM	1382	O	LEU	A	91	0.030	0.121	-29.552	1.00	0.00	O
ATOM	1398	N	ALA	A	92	-0.542	1.757	-28.106	1.00	0.00	N
ATOM	1399	CA	ALA	A	92	0.420	2.711	-28.648	1.00	0.00	C
ATOM	1400	C	ALA	A	92	0.314	4.082	-27.964	1.00	0.00	C
ATOM	1401	O	ALA	A	92	1.334	4.699	-27.652	1.00	0.00	O
ATOM	1408	N	ARG	A	93	-0.907	4.556	-27.704	1.00	0.00	N
ATOM	1409	CA	ARG	A	93	-1.069	5.842	-27.045	1.00	0.00	C
ATOM	1410	C	ARG	A	93	-0.781	5.684	-25.562	1.00	0.00	C
ATOM	1411	O	ARG	A	93	0.309	5.995	-25.081	1.00	0.00	O
ATOM	1432	N	GLY	A	94	-1.627	4.998	-24.825	1.00	0.00	N
ATOM	1433	CA	GLY	A	94	-1.593	4.980	-23.376	1.00	0.00	C
ATOM	1434	C	GLY	A	94	-2.581	3.906	-22.925	1.00	0.00	C
ATOM	1435	O	GLY	A	94	-3.736	4.276	-22.718	1.00	0.00	O
ATOM	1439	N	LEU	A	95	-2.119	2.673	-22.903	1.00	0.00	N
ATOM	1440	CA	LEU	A	95	-2.896	1.511	-22.475	1.00	0.00	C
ATOM	1441	C	LEU	A	95	-2.174	0.836	-21.302	1.00	0.00	C
ATOM	1442	O	LEU	A	95	-1.011	0.440	-21.427	1.00	0.00	O
ATOM	1458	N	LYS	A	96	-2.792	0.799	-20.112	1.00	0.00	N
ATOM	1459	CA	LYS	A	96	-2.234	0.286	-18.830	1.00	0.00	C
ATOM	1460	C	LYS	A	96	-3.108	-0.898	-18.402	1.00	0.00	C
ATOM	1461	O	LYS	A	96	-4.262	-0.518	-18.357	1.00	0.00	O
ATOM	1480	N	LEU	A	97	-2.597	-2.126	-18.149	1.00	0.00	N
ATOM	1481	CA	LEU	A	97	-3.371	-3.257	-17.629	1.00	0.00	C
ATOM	1482	C	LEU	A	97	-2.724	-3.752	-16.354	1.00	0.00	C
ATOM	1483	O	LEU	A	97	-1.656	-4.362	-16.418	1.00	0.00	O
ATOM	1499	N	THR	A	98	-3.348	-3.467	-15.199	1.00	0.00	N
ATOM	1500	CA	THR	A	98	-2.760	-3.885	-13.927	1.00	0.00	C
ATOM	1501	C	THR	A	98	-3.513	-4.986	-13.189	1.00	0.00	C
ATOM	1502	O	THR	A	98	-4.470	-4.709	-12.456	1.00	0.00	O
ATOM	1513	N	PHE	A	99	-3.036	-6.224	-13.307	1.00	0.00	N
ATOM	1514	CA	PHE	A	99	-3.646	-7.290	-12.541	1.00	0.00	C
ATOM	1515	C	PHE	A	99	-2.992	-7.208	-11.182	1.00	0.00	C
ATOM	1516	O	PHE	A	99	-1.764	-7.140	-11.091	1.00	0.00	O
ATOM	1533	N	ASP	A	100	-3.753	-7.049	-10.117	1.00	0.00	N
ATOM	1534	CA	ASP	A	100	-3.269	-6.903	-8.743	1.00	0.00	C
ATOM	1535	C	ASP	A	100	-3.550	-8.215	-8.014	1.00	0.00	C
ATOM	1536	O	ASP	A	100	-4.451	-8.909	-8.415	1.00	0.00	O
ATOM	1545	N	SER	A	101	-2.801	-8.486	-6.963	1.00	0.00	N
ATOM	1546	CA	SER	A	101	-3.055	-9.578	-6.024	1.00	0.00	C
ATOM	1547	C	SER	A	101	-2.602	-9.188	-4.631	1.00	0.00	C
ATOM	1548	O	SER	A	101	-1.406	-9.081	-4.370	1.00	0.00	O
ATOM	1567	N	PHE	A	103	-3.136	-10.341	-0.727	1.00	0.00	N
ATOM	1568	CA	PHE	A	103	-3.606	-11.319	0.252	1.00	0.00	C
ATOM	1569	C	PHE	A	103	-3.261	-10.843	1.671	1.00	0.00	C
ATOM	1570	O	PHE	A	103	-2.094	-10.894	2.059	1.00	0.00	O
ATOM	1587	N	SER	A	104	-4.261	-10.386	2.442	1.00	0.00	N
ATOM	1588	CA	SER	A	104	-3.998	-9.909	3.810	1.00	0.00	C
ATOM	1589	C	SER	A	104	-4.601	-10.835	4.889	1.00	0.00	C
ATOM	1590	O	SER	A	104	-5.524	-11.618	4.622	1.00	0.00	O
ATOM	1598	N	PRO	A	105	-4.087	-10.740	6.105	1.00	0.00	N
ATOM	1599	CA	PRO	A	105	-4.544	-11.555	7.275	1.00	0.00	C
ATOM	1600	C	PRO	A	105	-6.056	-11.771	7.353	1.00	0.00	C
ATOM	1601	O	PRO	A	105	-6.508	-12.898	7.550	1.00	0.00	O
ATOM	1612	N	ASN	A	106	-6.830	-10.693	7.233	1.00	0.00	N
ATOM	1613	CA	ASN	A	106	-8.288	-10.794	7.328	1.00	0.00	C
ATOM	1614	C	ASN	A	106	-8.798	-12.090	6.693	1.00	0.00	C
ATOM	1615	O	ASN	A	106	-9.049	-12.139	5.494	1.00	0.00	O
ATOM	1626	N	THR	A	107	-8.955	-13.126	7.520	1.00	0.00	N
ATOM	1627	CA	THR	A	107	-9.437	-14.432	7.057	1.00	0.00	C
ATOM	1628	C	THR	A	107	-8.949	-14.734	5.644	1.00	0.00	C
ATOM	1629	O	THR	A	107	-9.727	-15.174	4.798	1.00	0.00	O
ATOM	1640	N	GLY	A	108	-7.665	-14.497	5.391	1.00	0.00	N
ATOM	1641	CA	GLY	A	108	-7.104	-14.754	4.067	1.00	0.00	C
ATOM	1642	C	GLY	A	108	-7.839	-13.941	3.007	1.00	0.00	C
ATOM	1643	O	GLY	A	108	-8.243	-14.474	1.975	1.00	0.00	O
ATOM	1647	N	LYS	A	109	-8.015	-12.652	3.278	1.00	0.00	N
ATOM	1648	CA	LYS	A	109	-8.717	-11.772	2.343	1.00	0.00	C
ATOM	1649	C	LYS	A	109	-7.932	-11.634	1.043	1.00	0.00	C
ATOM	1650	O	LYS	A	109	-6.829	-11.086	1.028	1.00	0.00	O
ATOM	1669	N	LYS	A	110	-8.512	-12.148	-0.040	1.00	0.00	N

## Appendix D PDB files of the models for the closed state of VDAC

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ATOM	1670	CA	LYS A	110	-7.874	-12.104	-1.356	1.00	0.00	C
ATOM	1671	C	LYS A	110	-8.410	-10.945	-2.189	1.00	0.00	C
ATOM	1672	O	LYS A	110	-9.612	-10.855	-2.447	1.00	0.00	O
ATOM	1691	N	ASN A	111	-7.604	-10.019	-2.719	1.00	0.00	N
ATOM	1692	CA	ASN A	111	-8.048	-8.885	-3.578	1.00	0.00	C
ATOM	1693	C	ASN A	111	-7.439	-9.049	-4.973	1.00	0.00	C
ATOM	1694	O	ASN A	111	-6.286	-8.669	-5.139	1.00	0.00	O
ATOM	1705	N	ALA A	112	-8.215	-9.517	-5.939	1.00	0.00	N
ATOM	1706	CA	ALA A	112	-7.803	-9.787	-7.320	1.00	0.00	C
ATOM	1707	C	ALA A	112	-8.438	-8.753	-8.256	1.00	0.00	C
ATOM	1708	O	ALA A	112	-9.521	-9.047	-8.786	1.00	0.00	O
ATOM	1715	N	LYS A	113	-7.779	-7.608	-8.445	1.00	0.00	N
ATOM	1716	CA	LYS A	113	-8.255	-6.489	-9.309	1.00	0.00	C
ATOM	1717	C	LYS A	113	-7.586	-6.523	-10.685	1.00	0.00	C
ATOM	1718	O	LYS A	113	-6.438	-6.817	-10.598	1.00	0.00	O
ATOM	1737	N	ILE A	114	-8.281	-6.227	-11.754	1.00	0.00	N
ATOM	1738	CA	ILE A	114	-7.775	-6.106	-13.146	1.00	0.00	C
ATOM	1739	C	ILE A	114	-8.047	-4.648	-13.545	1.00	0.00	C
ATOM	1740	O	ILE A	114	-9.186	-4.551	-13.943	1.00	0.00	O
ATOM	1756	N	LYS A	115	-7.147	-3.692	-13.579	1.00	0.00	N
ATOM	1757	CA	LYS A	115	-7.446	-2.275	-13.912	1.00	0.00	C
ATOM	1758	C	LYS A	115	-7.098	-2.102	-15.385	1.00	0.00	C
ATOM	1759	O	LYS A	115	-5.984	-2.493	-15.542	1.00	0.00	O
ATOM	1778	N	THR A	116	-7.935	-1.631	-16.299	1.00	0.00	N
ATOM	1779	CA	THR A	116	-7.637	-1.391	-17.698	1.00	0.00	C
ATOM	1780	C	THR A	116	-7.941	0.060	-17.985	1.00	0.00	C
ATOM	1781	O	THR A	116	-9.099	0.446	-18.024	1.00	0.00	O
ATOM	1792	N	GLY A	117	-6.909	0.873	-18.218	1.00	0.00	N
ATOM	1793	CA	GLY A	117	-6.942	2.315	-18.399	1.00	0.00	C
ATOM	1794	C	GLY A	117	-6.491	2.691	-19.793	1.00	0.00	C
ATOM	1795	O	GLY A	117	-5.315	3.041	-19.959	1.00	0.00	O
ATOM	1796	H	GLY A	117	-5.985	0.603	-18.291	1.00	0.00	H
ATOM	1797	HA2	GLY A	117	-7.890	2.732	-18.254	1.00	0.00	H
ATOM	1798	HA3	GLY A	117	-6.281	2.759	-17.664	1.00	0.00	H
ATOM	1799	N	TYR A	118	-7.408	2.637	-20.743	1.00	0.00	N
ATOM	1800	CA	TYR A	118	-7.181	3.165	-22.086	1.00	0.00	C
ATOM	1801	C	TYR A	118	-7.174	4.682	-22.007	1.00	0.00	C
ATOM	1802	O	TYR A	118	-8.085	5.143	-21.301	1.00	0.00	O
ATOM	1820	N	LYS A	119	-6.270	5.326	-22.699	1.00	0.00	N
ATOM	1821	CA	LYS A	119	-6.076	6.771	-22.735	1.00	0.00	C
ATOM	1822	C	LYS A	119	-6.100	7.214	-24.205	1.00	0.00	C
ATOM	1823	O	LYS A	119	-5.654	6.458	-25.074	1.00	0.00	O
ATOM	1842	N	ARG A	120	-6.555	8.422	-24.454	1.00	0.00	N
ATOM	1843	CA	ARG A	120	-6.633	9.001	-25.795	1.00	0.00	C
ATOM	1844	C	ARG A	120	-6.914	10.482	-25.649	1.00	0.00	C
ATOM	1845	O	ARG A	120	-8.066	10.889	-25.650	1.00	0.00	O
ATOM	1866	N	GLU A	121	-5.850	11.275	-25.486	1.00	0.00	N
ATOM	1867	CA	GLU A	121	-5.970	12.718	-25.319	1.00	0.00	C
ATOM	1868	C	GLU A	121	-7.232	13.115	-24.536	1.00	0.00	C
ATOM	1869	O	GLU A	121	-8.362	12.908	-24.971	1.00	0.00	O
ATOM	1881	N	HIS A	122	-7.004	13.757	-23.405	1.00	0.00	N
ATOM	1882	CA	HIS A	122	-8.092	14.268	-22.560	1.00	0.00	C
ATOM	1883	C	HIS A	122	-9.091	13.192	-22.121	1.00	0.00	C
ATOM	1884	O	HIS A	122	-9.897	13.463	-21.222	1.00	0.00	O
ATOM	1898	N	ILE A	123	-9.078	12.003	-22.754	1.00	0.00	N
ATOM	1899	CA	ILE A	123	-10.031	10.950	-22.387	1.00	0.00	C
ATOM	1900	C	ILE A	123	-9.355	9.732	-21.772	1.00	0.00	C
ATOM	1901	O	ILE A	123	-8.661	8.993	-22.476	1.00	0.00	O
ATOM	1917	N	ASN A	124	-9.582	9.427	-20.481	1.00	0.00	N
ATOM	1918	CA	ASN A	124	-9.011	8.240	-19.787	1.00	0.00	C
ATOM	1919	C	ASN A	124	-10.197	7.406	-19.241	1.00	0.00	C
ATOM	1920	O	ASN A	124	-11.081	8.043	-18.665	1.00	0.00	O
ATOM	1931	N	LEU A	125	-10.182	6.084	-19.420	1.00	0.00	N
ATOM	1932	CA	LEU A	125	-11.295	5.165	-19.113	1.00	0.00	C
ATOM	1933	C	LEU A	125	-10.772	3.948	-18.346	1.00	0.00	C
ATOM	1934	O	LEU A	125	-10.441	2.901	-18.877	1.00	0.00	O
ATOM	1950	N	GLY A	126	-10.689	4.046	-17.000	1.00	0.00	N
ATOM	1951	CA	GLY A	126	-10.302	2.934	-16.136	1.00	0.00	C
ATOM	1952	C	GLY A	126	-11.398	1.883	-16.057	1.00	0.00	C
ATOM	1953	O	GLY A	126	-12.543	2.370	-16.032	1.00	0.00	O
ATOM	1957	N	CYS A	127	-11.072	0.576	-15.989	1.00	0.00	N
ATOM	1958	CA	CYS A	127	-11.994	-0.473	-15.555	1.00	0.00	C
ATOM	1959	C	CYS A	127	-11.319	-1.420	-14.547	1.00	0.00	C
ATOM	1960	O	CYS A	127	-10.488	-2.243	-14.916	1.00	0.00	O
ATOM	1968	N	ASP A	128	-11.561	-1.342	-13.263	1.00	0.00	N
ATOM	1969	CA	ASP A	128	-11.003	-2.148	-12.160	1.00	0.00	C
ATOM	1970	C	ASP A	128	-12.071	-3.167	-11.698	1.00	0.00	C
ATOM	1971	O	ASP A	128	-13.019	-2.710	-11.038	1.00	0.00	O
ATOM	1980	N	MET A	129	-11.888	-4.450	-11.985	1.00	0.00	N
ATOM	1981	CA	MET A	129	-12.812	-5.569	-11.686	1.00	0.00	C
ATOM	1982	C	MET A	129	-12.252	-6.366	-10.509	1.00	0.00	C
ATOM	1983	O	MET A	129	-11.429	-7.255	-10.738	1.00	0.00	O
ATOM	1997	N	ASP A	130	-12.716	-6.132	-9.290	1.00	0.00	N
ATOM	1998	CA	ASP A	130	-12.163	-6.764	-8.084	1.00	0.00	C
ATOM	1999	C	ASP A	130	-12.831	-8.110	-7.817	1.00	0.00	C
ATOM	2000	O	ASP A	130	-13.810	-8.174	-7.112	1.00	0.00	O
ATOM	2009	N	PHE A	131	-12.261	-9.211	-8.334	1.00	0.00	N
ATOM	2010	CA	PHE A	131	-12.789	-10.553	-8.062	1.00	0.00	C
ATOM	2011	C	PHE A	131	-12.511	-10.953	-6.612	1.00	0.00	C
ATOM	2012	O	PHE A	131	-11.570	-11.684	-6.299	1.00	0.00	O
ATOM	2029	N	ASP A	132	-13.375	-10.485	-5.740	1.00	0.00	N
ATOM	2030	CA	ASP A	132	-13.435	-10.796	-4.304	1.00	0.00	C
ATOM	2031	C	ASP A	132	-14.744	-11.555	-4.050	1.00	0.00	C
ATOM	2032	O	ASP A	132	-15.478	-11.725	-5.013	1.00	0.00	O

ATOM	2041	N	ILE	A	133	-15.043	-12.056	-2.863	1.00	0.00		N
ATOM	2042	CA	ILE	A	133	-16.299	-12.692	-2.516	1.00	0.00		C
ATOM	2043	C	ILE	A	133	-17.402	-11.644	-2.470	1.00	0.00		C
ATOM	2044	O	ILE	A	133	-18.483	-11.868	-3.016	1.00	0.00		O
ATOM	2060	N	ALA	A	134	-17.110	-10.468	-1.907	1.00	0.00		N
ATOM	2061	CA	ALA	A	134	-18.094	-9.393	-1.920	1.00	0.00		C
ATOM	2062	C	ALA	A	134	-18.226	-8.921	-3.367	1.00	0.00		C
ATOM	2063	O	ALA	A	134	-19.309	-8.586	-3.839	1.00	0.00		O
ATOM	2070	N	GLY	A	135	-17.098	-8.971	-4.081	1.00	0.00		N
ATOM	2071	CA	GLY	A	135	-17.067	-8.636	-5.502	1.00	0.00		C
ATOM	2072	C	GLY	A	135	-17.647	-9.834	-6.247	1.00	0.00		C
ATOM	2073	O	GLY	A	135	-18.151	-10.759	-5.603	1.00	0.00		O
ATOM	2077	N	PRO	A	136	-17.595	-9.903	-7.553	1.00	0.00		N
ATOM	2078	CA	PRO	A	136	-17.014	-8.867	-8.485	1.00	0.00		C
ATOM	2079	C	PRO	A	136	-17.568	-7.455	-8.318	1.00	0.00		C
ATOM	2080	O	PRO	A	136	-18.766	-7.235	-8.520	1.00	0.00		O
ATOM	2091	N	SER	A	137	-16.694	-6.473	-8.014	1.00	0.00		N
ATOM	2092	CA	SER	A	137	-17.022	-5.037	-7.872	1.00	0.00		C
ATOM	2093	C	SER	A	137	-16.499	-4.360	-9.145	1.00	0.00		C
ATOM	2094	O	SER	A	137	-15.285	-4.699	-9.289	1.00	0.00		O
ATOM	2102	N	ILE	A	138	-17.200	-3.514	-9.916	1.00	0.00		N
ATOM	2103	CA	ILE	A	138	-16.637	-2.818	-11.074	1.00	0.00		C
ATOM	2104	C	ILE	A	138	-16.393	-1.347	-10.754	1.00	0.00		C
ATOM	2105	O	ILE	A	138	-17.325	-0.557	-10.736	1.00	0.00		O
ATOM	2121	N	ARG	A	139	-15.186	-1.097	-10.306	1.00	0.00		N
ATOM	2122	CA	ARG	A	139	-14.719	0.254	-10.160	1.00	0.00		C
ATOM	2123	C	ARG	A	139	-14.212	0.641	-11.571	1.00	0.00		C
ATOM	2124	O	ARG	A	139	-13.466	-0.007	-12.069	1.00	0.00		O
ATOM	2145	N	GLY	A	140	-14.620	1.883	-12.091	1.00	0.00		N
ATOM	2146	CA	GLY	A	140	-13.940	2.514	-13.242	1.00	0.00		C
ATOM	2147	C	GLY	A	140	-13.930	4.028	-13.091	1.00	0.00		C
ATOM	2148	O	GLY	A	140	-14.709	4.519	-12.309	1.00	0.00		O
ATOM	2152	N	ALA	A	141	-13.049	4.768	-13.800	1.00	0.00		N
ATOM	2153	CA	ALA	A	141	-12.895	6.221	-13.753	1.00	0.00		C
ATOM	2154	C	ALA	A	141	-13.105	6.715	-15.175	1.00	0.00		C
ATOM	2155	O	ALA	A	141	-12.284	6.127	-15.929	1.00	0.00		O
ATOM	2162	N	LEU	A	142	-14.022	7.609	-15.511	1.00	0.00		N
ATOM	2163	CA	LEU	A	142	-14.108	8.188	-16.852	1.00	0.00		C
ATOM	2164	C	LEU	A	142	-13.750	9.650	-16.770	1.00	0.00		C
ATOM	2165	O	LEU	A	142	-14.527	10.417	-16.215	1.00	0.00		O
ATOM	2181	N	VAL	A	143	-12.576	10.009	-17.326	1.00	0.00		N
ATOM	2182	CA	VAL	A	143	-11.990	11.357	-17.218	1.00	0.00		C
ATOM	2183	C	VAL	A	143	-12.166	12.067	-18.555	1.00	0.00		C
ATOM	2184	O	VAL	A	143	-11.721	11.324	-19.364	1.00	0.00		O
ATOM	2197	N	LEU	A	144	-12.676	13.293	-18.654	1.00	0.00		N
ATOM	2198	CA	LEU	A	144	-12.748	14.088	-19.869	1.00	0.00		C
ATOM	2199	C	LEU	A	144	-11.959	15.356	-19.647	1.00	0.00		C
ATOM	2200	O	LEU	A	144	-12.057	15.957	-18.577	1.00	0.00		O
ATOM	2216	N	GLY	A	145	-11.182	15.774	-20.645	1.00	0.00		N
ATOM	2217	CA	GLY	A	145	-10.380	16.985	-20.507	1.00	0.00		C
ATOM	2218	C	GLY	A	145	-10.952	18.142	-21.324	1.00	0.00		C
ATOM	2219	O	GLY	A	145	-11.528	17.930	-22.392	1.00	0.00		O
ATOM	2223	N	TYR	A	146	-10.757	19.373	-20.847	1.00	0.00		N
ATOM	2224	CA	TYR	A	146	-11.228	20.544	-21.588	1.00	0.00		C
ATOM	2225	C	TYR	A	146	-10.260	21.693	-21.365	1.00	0.00		C
ATOM	2226	O	TYR	A	146	-9.130	21.437	-20.972	1.00	0.00		O
ATOM	2244	N	GLU	A	147	-10.726	22.939	-21.619	1.00	0.00		N
ATOM	2245	CA	GLU	A	147	-9.927	24.177	-21.462	1.00	0.00		C
ATOM	2246	C	GLU	A	147	-9.150	24.237	-20.137	1.00	0.00		C
ATOM	2247	O	GLU	A	147	-9.262	25.213	-19.398	1.00	0.00		O
ATOM	2259	N	GLY	A	148	-8.384	23.207	-19.836	1.00	0.00		N
ATOM	2260	CA	GLY	A	148	-7.633	23.156	-18.593	1.00	0.00		C
ATOM	2261	C	GLY	A	148	-8.480	22.539	-17.474	1.00	0.00		C
ATOM	2262	O	GLY	A	148	-8.370	22.937	-16.319	1.00	0.00		O
ATOM	2266	N	TRP	A	149	-9.328	21.571	-17.824	1.00	0.00		N
ATOM	2267	CA	TRP	A	149	-10.173	20.925	-16.809	1.00	0.00		C
ATOM	2268	C	TRP	A	149	-10.142	19.418	-16.943	1.00	0.00		C
ATOM	2269	O	TRP	A	149	-10.162	18.887	-18.038	1.00	0.00		O
ATOM	2290	N	LEU	A	150	-10.144	18.762	-15.792	1.00	0.00		N
ATOM	2291	CA	LEU	A	150	-10.181	17.306	-15.686	1.00	0.00		C
ATOM	2292	C	LEU	A	150	-11.414	16.903	-14.857	1.00	0.00		C
ATOM	2293	O	LEU	A	150	-11.424	17.122	-13.647	1.00	0.00		O
ATOM	2309	N	ALA	A	151	-12.414	16.280	-15.466	1.00	0.00		N
ATOM	2310	CA	ALA	A	151	-13.589	15.824	-14.700	1.00	0.00		C
ATOM	2311	C	ALA	A	151	-13.598	14.307	-14.712	1.00	0.00		C
ATOM	2312	O	ALA	A	151	-13.263	13.742	-15.735	1.00	0.00		O
ATOM	2319	N	GLY	A	152	-13.940	13.632	-13.597	1.00	0.00		N
ATOM	2320	CA	GLY	A	152	-13.921	12.170	-13.582	1.00	0.00		C
ATOM	2321	C	GLY	A	152	-15.195	11.583	-12.990	1.00	0.00		C
ATOM	2322	O	GLY	A	152	-15.832	12.195	-12.125	1.00	0.00		O
ATOM	2326	N	TYR	A	153	-15.550	10.379	-13.446	1.00	0.00		N
ATOM	2327	CA	TYR	A	153	-16.744	9.702	-12.954	1.00	0.00		C
ATOM	2328	C	TYR	A	153	-16.344	8.315	-12.535	1.00	0.00		C
ATOM	2329	O	TYR	A	153	-15.985	7.507	-13.393	1.00	0.00		O
ATOM	2347	N	GLN	A	154	-16.392	8.016	-11.234	1.00	0.00		N
ATOM	2348	CA	GLN	A	154	-16.002	6.713	-10.769	1.00	0.00		C
ATOM	2349	C	GLN	A	154	-17.232	5.893	-10.423	1.00	0.00		C
ATOM	2350	O	GLN	A	154	-17.926	6.238	-9.485	1.00	0.00		O
ATOM	2364	N	MET	A	155	-17.488	4.822	-11.178	1.00	0.00		N
ATOM	2365	CA	MET	A	155	-18.659	3.976	-10.918	1.00	0.00		C
ATOM	2366	C	MET	A	155	-18.271	2.637	-10.289	1.00	0.00		C
ATOM	2367	O	MET	A	155	-17.598	1.814	-10.909	1.00	0.00		O
ATOM	2381	N	ASN	A	156	-18.754	2.421	-9.070	1.00	0.00		N
ATOM	2382	CA	ASN	A	156	-18.521	1.172	-8.362	1.00	0.00		C

## Appendix D PDB files of the models for the closed state of VDAC

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ATOM	2383	C	ASN A	156	-19.795	0.338	-8.427	1.00	0.00	C
ATOM	2384	O	ASN A	156	-20.830	0.712	-7.868	1.00	0.00	O
ATOM	2395	N	PHE A	157	-19.707	-0.784	-9.115	1.00	0.00	N
ATOM	2396	CA	PHE A	157	-20.857	-1.669	-9.266	1.00	0.00	C
ATOM	2397	C	PHE A	157	-20.680	-2.945	-8.461	1.00	0.00	C
ATOM	2398	O	PHE A	157	-20.048	-3.892	-8.914	1.00	0.00	O
ATOM	2415	N	GLU A	158	-21.234	-2.958	-7.262	1.00	0.00	N
ATOM	2416	CA	GLU A	158	-21.142	-4.142	-6.408	1.00	0.00	C
ATOM	2417	C	GLU A	158	-22.094	-5.200	-6.963	1.00	0.00	C
ATOM	2418	O	GLU A	158	-23.299	-5.136	-6.753	1.00	0.00	O
ATOM	2430	N	THR A	159	-21.554	-6.143	-7.732	1.00	0.00	N
ATOM	2431	CA	THR A	159	-22.391	-7.164	-8.378	1.00	0.00	C
ATOM	2432	C	THR A	159	-23.040	-8.143	-7.392	1.00	0.00	C
ATOM	2433	O	THR A	159	-24.148	-8.617	-7.644	1.00	0.00	O
ATOM	2444	N	ALA A	160	-22.372	-8.464	-6.285	1.00	0.00	N
ATOM	2445	CA	ALA A	160	-22.960	-9.409	-5.334	1.00	0.00	C
ATOM	2446	C	ALA A	160	-24.291	-8.869	-4.830	1.00	0.00	C
ATOM	2447	O	ALA A	160	-25.270	-9.603	-4.708	1.00	0.00	O
ATOM	2454	N	LYS A	161	-24.323	-7.567	-4.576	1.00	0.00	N
ATOM	2455	CA	LYS A	161	-25.545	-6.912	-4.126	1.00	0.00	C
ATOM	2456	C	LYS A	161	-26.203	-6.244	-5.329	1.00	0.00	C
ATOM	2457	O	LYS A	161	-27.352	-5.813	-5.273	1.00	0.00	O
ATOM	2476	N	SER A	162	-25.431	-6.144	-6.403	1.00	0.00	N
ATOM	2477	CA	SER A	162	-25.890	-5.497	-7.629	1.00	0.00	C
ATOM	2478	C	SER A	162	-26.352	-4.084	-7.302	1.00	0.00	C
ATOM	2479	O	SER A	162	-27.461	-3.681	-7.650	1.00	0.00	O
ATOM	2487	N	ARG A	163	-25.483	-3.343	-6.612	1.00	0.00	N
ATOM	2488	CA	ARG A	163	-25.789	-1.975	-6.216	1.00	0.00	C
ATOM	2489	C	ARG A	163	-24.685	-1.009	-6.628	1.00	0.00	C
ATOM	2490	O	ARG A	163	-23.497	-1.287	-6.447	1.00	0.00	O
ATOM	2511	N	VAL A	164	-25.096	0.161	-7.103	1.00	0.00	N
ATOM	2512	CA	VAL A	164	-24.166	1.203	-7.445	1.00	0.00	C
ATOM	2513	C	VAL A	164	-24.270	2.226	-6.335	1.00	0.00	C
ATOM	2514	O	VAL A	164	-25.227	3.001	-6.275	1.00	0.00	O
ATOM	2527	N	THR A	165	-23.311	2.193	-5.447	1.00	0.00	N
ATOM	2528	CA	THR A	165	-23.309	3.102	-4.297	1.00	0.00	C
ATOM	2529	C	THR A	165	-22.067	3.967	-4.298	1.00	0.00	C
ATOM	2530	O	THR A	165	-22.120	5.198	-4.286	1.00	0.00	O
ATOM	2541	N	GLN A	166	-20.955	3.281	-4.297	1.00	0.00	N
ATOM	2542	CA	GLN A	166	-19.664	3.944	-4.286	1.00	0.00	C
ATOM	2543	C	GLN A	166	-19.401	4.641	-5.620	1.00	0.00	C
ATOM	2544	O	GLN A	166	-19.147	3.996	-6.639	1.00	0.00	O
ATOM	2558	N	SER A	167	-19.446	5.969	-5.594	1.00	0.00	N
ATOM	2559	CA	SER A	167	-19.190	6.762	-6.791	1.00	0.00	C
ATOM	2560	C	SER A	167	-18.358	7.989	-6.426	1.00	0.00	C
ATOM	2561	O	SER A	167	-18.558	8.585	-5.364	1.00	0.00	O
ATOM	2569	N	ASN A	168	-17.400	8.340	-7.288	1.00	0.00	N
ATOM	2570	CA	ASN A	168	-16.529	9.475	-7.019	1.00	0.00	C
ATOM	2571	C	ASN A	168	-16.478	10.455	-8.198	1.00	0.00	C
ATOM	2572	O	ASN A	168	-16.063	10.094	-9.300	1.00	0.00	O
ATOM	2583	N	PHE A	169	-16.869	11.706	-7.944	1.00	0.00	N
ATOM	2584	CA	PHE A	169	-16.834	12.749	-8.978	1.00	0.00	C
ATOM	2585	C	PHE A	169	-15.509	13.497	-8.891	1.00	0.00	C
ATOM	2586	O	PHE A	169	-15.266	14.195	-7.911	1.00	0.00	O
ATOM	2603	N	ALA A	170	-14.671	13.390	-9.909	1.00	0.00	N
ATOM	2604	CA	ALA A	170	-13.405	14.110	-9.895	1.00	0.00	C
ATOM	2605	C	ALA A	170	-13.570	15.450	-10.582	1.00	0.00	C
ATOM	2606	O	ALA A	170	-14.085	15.501	-11.675	1.00	0.00	O
ATOM	2613	N	VAL A	171	-13.104	16.522	-9.958	1.00	0.00	N
ATOM	2614	CA	VAL A	171	-13.182	17.844	-10.575	1.00	0.00	C
ATOM	2615	C	VAL A	171	-11.817	18.518	-10.462	1.00	0.00	C
ATOM	2616	O	VAL A	171	-11.252	18.607	-9.370	1.00	0.00	O
ATOM	2629	N	GLY A	172	-11.286	18.962	-11.599	1.00	0.00	N
ATOM	2630	CA	GLY A	172	-9.995	19.595	-11.597	1.00	0.00	C
ATOM	2631	C	GLY A	172	-9.946	20.835	-12.465	1.00	0.00	C
ATOM	2632	O	GLY A	172	-10.577	20.915	-13.520	1.00	0.00	O
ATOM	2636	N	TYR A	173	-9.147	21.780	-12.002	1.00	0.00	N
ATOM	2637	CA	TYR A	173	-8.937	23.034	-12.703	1.00	0.00	C
ATOM	2638	C	TYR A	173	-7.445	23.256	-12.853	1.00	0.00	C
ATOM	2639	O	TYR A	173	-6.695	23.128	-11.886	1.00	0.00	O
ATOM	2657	N	LYS A	174	-7.001	23.528	-14.063	1.00	0.00	N
ATOM	2658	CA	LYS A	174	-5.575	23.696	-14.321	1.00	0.00	C
ATOM	2659	C	LYS A	174	-5.077	25.142	-14.207	1.00	0.00	C
ATOM	2660	O	LYS A	174	-5.749	26.103	-14.571	1.00	0.00	O
ATOM	2679	N	THR A	175	-3.846	25.234	-13.746	1.00	0.00	N
ATOM	2680	CA	THR A	175	-3.129	26.494	-13.600	1.00	0.00	C
ATOM	2681	C	THR A	175	-1.683	26.253	-14.049	1.00	0.00	C
ATOM	2682	O	THR A	175	-1.205	25.121	-13.979	1.00	0.00	O
ATOM	2693	N	ASP A	176	-1.003	27.283	-14.549	1.00	0.00	N
ATOM	2694	CA	ASP A	176	0.368	27.105	-15.045	1.00	0.00	C
ATOM	2695	C	ASP A	176	1.166	26.140	-14.172	1.00	0.00	C
ATOM	2696	O	ASP A	176	1.327	26.372	-12.974	1.00	0.00	O
ATOM	2705	N	GLU A	177	1.653	25.050	-14.788	1.00	0.00	N
ATOM	2706	CA	GLU A	177	2.442	24.032	-14.075	1.00	0.00	C
ATOM	2707	C	GLU A	177	1.672	23.394	-12.916	1.00	0.00	C
ATOM	2708	O	GLU A	177	2.084	22.353	-12.385	1.00	0.00	O
ATOM	2720	N	PHE A	178	0.624	24.070	-12.458	1.00	0.00	N
ATOM	2721	CA	PHE A	178	-0.128	23.606	-11.299	1.00	0.00	C
ATOM	2722	C	PHE A	178	-1.471	22.963	-11.691	1.00	0.00	C
ATOM	2723	O	PHE A	178	-2.206	23.515	-12.501	1.00	0.00	O
ATOM	2740	N	GLN A	179	-1.812	21.818	-11.062	1.00	0.00	N
ATOM	2741	CA	GLN A	179	-3.104	21.147	-11.306	1.00	0.00	C
ATOM	2742	C	GLN A	179	-3.947	21.119	-10.031	1.00	0.00	C
ATOM	2743	O	GLN A	179	-3.421	20.817	-8.954	1.00	0.00	O

ATOM	2757	N	LEU	A	180	-5.260	21.335	-10.175	1.00	0.00		N
ATOM	2758	CA	LEU	A	180	-6.175	21.258	-9.032	1.00	0.00		C
ATOM	2759	C	LEU	A	180	-7.005	19.991	-9.167	1.00	0.00		C
ATOM	2760	O	LEU	A	180	-7.822	19.893	-10.060	1.00	0.00		O
ATOM	2776	N	HIS	A	181	-6.754	19.022	-8.294	1.00	0.00		N
ATOM	2777	CA	HIS	A	181	-7.475	17.748	-8.327	1.00	0.00		C
ATOM	2778	C	HIS	A	181	-8.258	17.496	-7.027	1.00	0.00		C
ATOM	2779	O	HIS	A	181	-7.664	17.357	-5.955	1.00	0.00		O
ATOM	2793	N	THR	A	182	-9.583	17.396	-7.136	1.00	0.00		N
ATOM	2794	CA	THR	A	182	-10.447	17.111	-5.974	1.00	0.00		C
ATOM	2795	C	THR	A	182	-11.452	16.034	-6.349	1.00	0.00		C
ATOM	2796	O	THR	A	182	-11.788	15.924	-7.534	1.00	0.00		O
ATOM	2807	N	ASN	A	183	-11.976	15.239	-5.384	1.00	0.00		N
ATOM	2808	CA	ASN	A	183	-12.956	14.238	-5.737	1.00	0.00		C
ATOM	2809	C	ASN	A	183	-14.044	14.194	-4.675	1.00	0.00		C
ATOM	2810	O	ASN	A	183	-13.754	14.206	-3.480	1.00	0.00		O
ATOM	2821	N	VAL	A	184	-15.290	14.112	-5.111	1.00	0.00		N
ATOM	2822	CA	VAL	A	184	-16.392	14.037	-4.169	1.00	0.00		C
ATOM	2823	C	VAL	A	184	-16.645	12.582	-3.918	1.00	0.00		C
ATOM	2824	O	VAL	A	184	-16.972	11.847	-4.845	1.00	0.00		O
ATOM	2837	N	ASN	A	185	-16.479	12.152	-2.679	1.00	0.00		N
ATOM	2838	CA	ASN	A	185	-16.674	10.770	-2.360	1.00	0.00		C
ATOM	2839	C	ASN	A	185	-18.056	10.558	-1.800	1.00	0.00		C
ATOM	2840	O	ASN	A	185	-18.358	11.042	-0.709	1.00	0.00		O
ATOM	2851	N	ASP	A	186	-18.864	9.860	-2.589	1.00	0.00		N
ATOM	2852	CA	ASP	A	186	-20.247	9.528	-2.228	1.00	0.00		C
ATOM	2853	C	ASP	A	186	-21.070	10.751	-1.835	1.00	0.00		C
ATOM	2854	O	ASP	A	186	-22.064	10.620	-1.117	1.00	0.00		O
ATOM	2863	N	GLY	A	187	-20.679	11.931	-2.304	1.00	0.00		N
ATOM	2864	CA	GLY	A	187	-21.432	13.134	-1.969	1.00	0.00		C
ATOM	2865	C	GLY	A	187	-21.267	13.467	-0.493	1.00	0.00		C
ATOM	2866	O	GLY	A	187	-22.082	14.191	0.078	1.00	0.00		O
ATOM	2870	N	THR	A	188	-20.243	12.881	0.130	1.00	0.00		N
ATOM	2871	CA	THR	A	188	-20.019	13.071	1.558	1.00	0.00		C
ATOM	2872	C	THR	A	188	-18.633	13.652	1.888	1.00	0.00		C
ATOM	2873	O	THR	A	188	-18.538	14.775	2.383	1.00	0.00		O
ATOM	2884	N	GLU	A	189	-17.562	12.894	1.657	1.00	0.00		N
ATOM	2885	CA	GLU	A	189	-16.213	13.404	1.994	1.00	0.00		C
ATOM	2886	C	GLU	A	189	-15.380	13.741	0.757	1.00	0.00		C
ATOM	2887	O	GLU	A	189	-15.173	12.888	-0.104	1.00	0.00		O
ATOM	2899	N	PHE	A	190	-14.880	14.985	0.674	1.00	0.00		N
ATOM	2900	CA	PHE	A	190	-14.059	15.407	-0.455	1.00	0.00		C
ATOM	2901	C	PHE	A	190	-12.584	15.143	-0.157	1.00	0.00		C
ATOM	2902	O	PHE	A	190	-12.148	15.187	0.986	1.00	0.00		O
ATOM	2919	N	GLY	A	191	-11.815	14.896	-1.221	1.00	0.00		N
ATOM	2920	CA	GLY	A	191	-10.384	14.651	-1.064	1.00	0.00		C
ATOM	2921	C	GLY	A	191	-9.672	14.565	-2.403	1.00	0.00		C
ATOM	2922	O	GLY	A	191	-10.234	14.067	-3.382	1.00	0.00		O
ATOM	2926	N	GLY	A	192	-8.445	15.094	-2.469	1.00	0.00		N
ATOM	2927	CA	GLY	A	192	-7.689	15.072	-3.717	1.00	0.00		C
ATOM	2928	C	GLY	A	192	-6.308	15.682	-3.516	1.00	0.00		C
ATOM	2929	O	GLY	A	192	-5.829	15.794	-2.384	1.00	0.00		O
ATOM	2933	N	SER	A	193	-5.669	16.072	-4.618	1.00	0.00		N
ATOM	2934	CA	SER	A	193	-4.329	16.647	-4.541	1.00	0.00		C
ATOM	2935	C	SER	A	193	-4.161	17.859	-5.443	1.00	0.00		C
ATOM	2936	O	SER	A	193	-5.013	18.168	-6.277	1.00	0.00		O
ATOM	2944	N	ILE	A	194	-3.010	18.499	-5.314	1.00	0.00		N
ATOM	2945	CA	ILE	A	194	-2.667	19.632	-6.145	1.00	0.00		C
ATOM	2946	C	ILE	A	194	-1.263	19.384	-6.674	1.00	0.00		C
ATOM	2947	O	ILE	A	194	-0.313	19.382	-5.899	1.00	0.00		O
ATOM	2963	N	TYR	A	195	-1.129	19.165	-7.971	1.00	0.00		N
ATOM	2964	CA	TYR	A	195	0.198	18.903	-8.544	1.00	0.00		C
ATOM	2965	C	TYR	A	195	0.872	20.222	-8.844	1.00	0.00		C
ATOM	2966	O	TYR	A	195	0.342	21.005	-9.621	1.00	0.00		O
ATOM	2984	N	GLN	A	196	2.017	20.482	-8.211	1.00	0.00		N
ATOM	2985	CA	GLN	A	196	2.702	21.745	-8.429	1.00	0.00		C
ATOM	2986	C	GLN	A	196	4.137	21.564	-8.912	1.00	0.00		C
ATOM	2987	O	GLN	A	196	5.038	21.347	-8.100	1.00	0.00		O
ATOM	3001	N	LYS	A	197	4.376	21.757	-10.212	1.00	0.00		N
ATOM	3002	CA	LYS	A	197	5.749	21.704	-10.699	1.00	0.00		C
ATOM	3003	C	LYS	A	197	6.293	23.109	-10.493	1.00	0.00		C
ATOM	3004	O	LYS	A	197	5.990	24.026	-11.258	1.00	0.00		O
ATOM	3023	N	VAL	A	198	7.027	23.285	-9.401	1.00	0.00		N
ATOM	3024	CA	VAL	A	198	7.543	24.590	-9.022	1.00	0.00		C
ATOM	3025	C	VAL	A	198	8.727	25.010	-9.871	1.00	0.00		C
ATOM	3026	O	VAL	A	198	8.834	26.166	-10.280	1.00	0.00		O
ATOM	3039	N	ASN	A	199	9.605	24.069	-10.137	1.00	0.00		N
ATOM	3040	CA	ASN	A	199	10.780	24.342	-10.954	1.00	0.00		C
ATOM	3041	C	ASN	A	199	11.446	23.022	-11.316	1.00	0.00		C
ATOM	3042	O	ASN	A	199	10.865	21.961	-11.108	1.00	0.00		O
ATOM	3053	N	LYS	A	200	12.641	23.084	-11.876	1.00	0.00		N
ATOM	3054	CA	LYS	A	200	13.337	21.864	-12.278	1.00	0.00		C
ATOM	3055	C	LYS	A	200	13.635	20.957	-11.081	1.00	0.00		C
ATOM	3056	O	LYS	A	200	13.506	19.739	-11.178	1.00	0.00		O
ATOM	3075	N	LYS	A	201	14.069	21.543	-9.968	1.00	0.00		N
ATOM	3076	CA	LYS	A	201	14.418	20.744	-8.791	1.00	0.00		C
ATOM	3077	C	LYS	A	201	13.303	20.658	-7.738	1.00	0.00		C
ATOM	3078	O	LYS	A	201	13.479	19.964	-6.737	1.00	0.00		O
ATOM	3097	N	LEU	A	202	12.194	21.397	-7.805	1.00	0.00		N
ATOM	3098	CA	LEU	A	202	11.032	21.429	-6.854	1.00	0.00		C
ATOM	3099	C	LEU	A	202	9.756	20.801	-7.456	1.00	0.00		C
ATOM	3100	O	LEU	A	202	9.147	21.460	-8.335	1.00	0.00		O
ATOM	3116	N	GLU	A	203	9.380	19.609	-6.992	1.00	0.00		N
ATOM	3117	CA	GLU	A	203	8.077	18.975	-7.322	1.00	0.00		C



## Appendix D PDB files of the models for the closed state of VDAC

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ATOM	3118	C	GLU A	203	7.319	18.795	-6.019	1.00	0.00	C
ATOM	3119	O	GLU A	203	7.774	17.877	-5.301	1.00	0.00	O
ATOM	3131	N	THR A	204	6.301	19.643	-5.686	1.00	0.00	N
ATOM	3132	CA	THR A	204	5.548	19.562	-4.443	1.00	0.00	C
ATOM	3133	C	THR A	204	4.092	19.199	-4.719	1.00	0.00	C
ATOM	3134	O	THR A	204	3.613	19.364	-5.842	1.00	0.00	O
ATOM	3145	N	ALA A	205	3.377	18.708	-3.701	1.00	0.00	N
ATOM	3146	CA	ALA A	205	1.983	18.346	-3.876	1.00	0.00	C
ATOM	3147	C	ALA A	205	1.184	18.695	-2.627	1.00	0.00	C
ATOM	3148	O	ALA A	205	1.529	18.277	-1.520	1.00	0.00	O
ATOM	3155	N	VAL A	206	0.100	19.446	-2.815	1.00	0.00	N
ATOM	3156	CA	VAL A	206	-0.759	19.831	-1.687	1.00	0.00	C
ATOM	3157	C	VAL A	206	-2.005	18.939	-1.667	1.00	0.00	C
ATOM	3158	O	VAL A	206	-2.719	18.832	-2.664	1.00	0.00	O
ATOM	3171	N	ASN A	207	-2.268	18.299	-0.532	1.00	0.00	N
ATOM	3172	CA	ASN A	207	-3.435	17.434	-0.428	1.00	0.00	C
ATOM	3173	C	ASN A	207	-4.575	18.175	0.240	1.00	0.00	C
ATOM	3174	O	ASN A	207	-4.382	18.730	1.309	1.00	0.00	O
ATOM	3185	N	LEU A	208	-5.754	18.179	-0.401	1.00	0.00	N
ATOM	3186	CA	LEU A	208	-6.923	18.875	0.166	1.00	0.00	C
ATOM	3187	C	LEU A	208	-8.004	17.881	0.574	1.00	0.00	C
ATOM	3188	O	LEU A	208	-8.645	17.285	-0.289	1.00	0.00	O
ATOM	3204	N	ALA A	209	-8.160	17.620	1.869	1.00	0.00	N
ATOM	3205	CA	ALA A	209	-9.215	16.783	2.467	1.00	0.00	C
ATOM	3206	C	ALA A	209	-10.227	17.690	3.147	1.00	0.00	C
ATOM	3207	O	ALA A	209	-9.711	18.531	3.912	1.00	0.00	O
ATOM	3214	N	TRP A	210	-11.539	17.546	2.906	1.00	0.00	N
ATOM	3215	CA	TRP A	210	-12.539	18.206	3.720	1.00	0.00	C
ATOM	3216	C	TRP A	210	-13.776	17.328	3.832	1.00	0.00	C
ATOM	3217	O	TRP A	210	-13.881	16.293	3.177	1.00	0.00	O
ATOM	3238	N	THR A	211	-14.693	17.716	4.686	1.00	0.00	N
ATOM	3239	CA	THR A	211	-15.890	16.906	4.904	1.00	0.00	C
ATOM	3240	C	THR A	211	-17.154	17.730	4.796	1.00	0.00	C
ATOM	3241	O	THR A	211	-17.374	18.632	5.602	1.00	0.00	O
ATOM	3252	N	ALA A	212	-17.990	17.428	3.793	1.00	0.00	N
ATOM	3253	CA	ALA A	212	-19.223	18.192	3.629	1.00	0.00	C
ATOM	3254	C	ALA A	212	-19.878	18.446	4.971	1.00	0.00	C
ATOM	3255	O	ALA A	212	-20.550	17.579	5.535	1.00	0.00	O
ATOM	3262	N	GLY A	213	-19.660	19.655	5.457	1.00	0.00	N
ATOM	3263	CA	GLY A	213	-20.190	20.102	6.724	1.00	0.00	C
ATOM	3264	C	GLY A	213	-19.617	21.482	7.035	1.00	0.00	C
ATOM	3265	O	GLY A	213	-19.153	22.185	6.137	1.00	0.00	O
ATOM	3269	N	ASN A	214	-19.625	21.867	8.295	1.00	0.00	N
ATOM	3270	CA	ASN A	214	-19.081	23.162	8.665	1.00	0.00	C
ATOM	3271	C	ASN A	214	-17.667	22.973	9.206	1.00	0.00	C
ATOM	3272	O	ASN A	214	-17.083	23.889	9.776	1.00	0.00	O
ATOM	3283	N	SER A	215	-16.966	21.852	8.992	1.00	0.00	N
ATOM	3284	CA	SER A	215	-15.632	21.562	9.564	1.00	0.00	C
ATOM	3285	C	SER A	215	-14.536	22.132	8.643	1.00	0.00	C
ATOM	3286	O	SER A	215	-14.805	22.476	7.490	1.00	0.00	O
ATOM	3294	N	ASN A	216	-13.302	22.216	9.147	1.00	0.00	N
ATOM	3295	CA	ASN A	216	-12.212	22.805	8.353	1.00	0.00	C
ATOM	3296	C	ASN A	216	-11.758	21.909	7.201	1.00	0.00	C
ATOM	3297	O	ASN A	216	-12.174	20.734	7.187	1.00	0.00	O
ATOM	3308	N	THR A	217	-10.947	22.493	6.313	1.00	0.00	N
ATOM	3309	CA	THR A	217	-10.249	21.852	5.190	1.00	0.00	C
ATOM	3310	C	THR A	217	-8.874	21.906	5.561	1.00	0.00	C
ATOM	3311	O	THR A	217	-8.330	22.938	5.837	1.00	0.00	O
ATOM	3322	N	ARG A	218	-8.703	21.292	5.320	1.00	0.00	N
ATOM	3323	CA	ARG A	218	-7.551	20.824	5.995	1.00	0.00	C
ATOM	3324	C	ARG A	218	-6.626	20.324	4.929	1.00	0.00	C
ATOM	3325	O	ARG A	218	-7.160	19.810	3.932	1.00	0.00	O
ATOM	3346	N	PHE A	219	-5.310	20.462	5.073	1.00	0.00	N
ATOM	3347	CA	PHE A	219	-4.434	20.241	3.944	1.00	0.00	C
ATOM	3348	C	PHE A	219	-2.962	20.124	4.190	1.00	0.00	C
ATOM	3349	O	PHE A	219	-2.624	21.000	4.984	1.00	0.00	O
ATOM	3366	N	GLY A	220	-2.245	19.210	3.484	1.00	0.00	N
ATOM	3367	CA	GLY A	220	-0.808	19.110	3.667	1.00	0.00	C
ATOM	3368	C	GLY A	220	-0.056	19.725	2.518	1.00	0.00	C
ATOM	3369	O	GLY A	220	-0.594	19.819	1.420	1.00	0.00	O
ATOM	3373	N	ILE A	221	1.201	20.118	2.769	1.00	0.00	N
ATOM	3374	CA	ILE A	221	2.026	20.686	1.709	1.00	0.00	C
ATOM	3375	C	ILE A	221	3.298	19.888	1.670	1.00	0.00	C
ATOM	3376	O	ILE A	221	4.139	20.059	2.546	1.00	0.00	O
ATOM	3392	N	ALA A	222	3.436	19.013	0.680	1.00	0.00	N
ATOM	3393	CA	ALA A	222	4.627	18.192	0.597	1.00	0.00	C
ATOM	3394	C	ALA A	222	5.554	18.681	-0.495	1.00	0.00	C
ATOM	3395	O	ALA A	222	5.179	18.721	-1.661	1.00	0.00	O
ATOM	3402	N	ALA A	223	6.763	19.047	-0.091	1.00	0.00	N
ATOM	3403	CA	ALA A	223	7.755	19.542	-1.030	1.00	0.00	C
ATOM	3404	C	ALA A	223	8.952	18.608	-1.116	1.00	0.00	C
ATOM	3405	O	ALA A	223	9.615	18.347	-0.113	1.00	0.00	O
ATOM	3412	N	LYS A	224	9.237	17.885	-2.241	1.00	0.00	N
ATOM	3413	CA	LYS A	224	10.476	17.126	-2.508	1.00	0.00	C
ATOM	3414	C	LYS A	224	11.381	18.162	-3.162	1.00	0.00	C
ATOM	3415	O	LYS A	224	10.995	18.451	-4.341	1.00	0.00	O
ATOM	3434	N	TYR A	225	12.433	18.609	-2.460	1.00	0.00	N
ATOM	3435	CA	TYR A	225	13.468	19.481	-2.997	1.00	0.00	C
ATOM	3436	C	TYR A	225	14.764	18.755	-3.274	1.00	0.00	C
ATOM	3437	O	TYR A	225	15.591	18.552	-2.385	1.00	0.00	O
ATOM	3455	N	GLN A	226	14.996	18.424	-4.534	1.00	0.00	N
ATOM	3456	CA	GLN A	226	16.175	17.704	-4.978	1.00	0.00	C
ATOM	3457	C	GLN A	226	17.290	18.718	-4.932	1.00	0.00	C
ATOM	3458	O	GLN A	226	17.333	19.594	-5.622	1.00	0.00	O

ATOM	3472	N	ILE A	227	18.238	18.567	-4.034	1.00	0.00		N
ATOM	3473	CA	ILE A	227	19.236	19.514	-3.808	1.00	0.00		C
ATOM	3474	C	ILE A	227	20.307	19.534	-4.903	1.00	0.00		C
ATOM	3475	O	ILE A	227	20.528	20.532	-5.607	1.00	0.00		O
ATOM	3491	N	ASP A	228	20.912	18.354	-5.016	1.00	0.00		N
ATOM	3492	CA	ASP A	228	21.953	18.039	-5.993	1.00	0.00		C
ATOM	3493	C	ASP A	228	21.675	16.592	-6.336	1.00	0.00		C
ATOM	3494	O	ASP A	228	20.741	16.023	-5.777	1.00	0.00		O
ATOM	3503	N	PRO A	229	22.366	15.978	-7.229	1.00	0.00		N
ATOM	3504	CA	PRO A	229	22.032	14.583	-7.603	1.00	0.00		C
ATOM	3505	C	PRO A	229	22.214	13.598	-6.445	1.00	0.00		C
ATOM	3506	O	PRO A	229	21.736	12.467	-6.520	1.00	0.00		O
ATOM	3517	N	ASP A	230	22.943	14.012	-5.400	1.00	0.00		N
ATOM	3518	CA	ASP A	230	23.222	13.133	-4.272	1.00	0.00		C
ATOM	3519	C	ASP A	230	22.447	13.502	-2.996	1.00	0.00		C
ATOM	3520	O	ASP A	230	22.665	12.875	-1.963	1.00	0.00		O
ATOM	3529	N	ALA A	231	21.558	14.504	-3.035	1.00	0.00		N
ATOM	3530	CA	ALA A	231	20.817	14.867	-1.813	1.00	0.00		C
ATOM	3531	C	ALA A	231	19.461	15.517	-2.120	1.00	0.00		C
ATOM	3532	O	ALA A	231	19.319	16.238	-3.109	1.00	0.00		O
ATOM	3539	N	CYS A	232	18.447	15.236	-1.265	1.00	0.00		N
ATOM	3540	CA	CYS A	232	17.018	15.602	-1.466	1.00	0.00		C
ATOM	3541	C	CYS A	232	16.344	15.976	-0.121	1.00	0.00		C
ATOM	3542	O	CYS A	232	16.525	15.076	0.717	1.00	0.00		O
ATOM	3550	N	PHE A	233	15.618	17.110	0.038	1.00	0.00		N
ATOM	3551	CA	PHE A	233	14.903	17.441	1.286	1.00	0.00		C
ATOM	3552	C	PHE A	233	13.384	17.326	1.122	1.00	0.00		C
ATOM	3553	O	PHE A	233	12.782	18.039	0.320	1.00	0.00		O
ATOM	3570	N	SER A	234	12.772	16.379	1.841	1.00	0.00		N
ATOM	3571	CA	SER A	234	11.317	16.142	1.862	1.00	0.00		C
ATOM	3572	C	SER A	234	10.736	16.901	3.065	1.00	0.00		C
ATOM	3573	O	SER A	234	11.392	16.769	4.107	1.00	0.00		O
ATOM	3581	N	ALA A	235	9.643	17.651	2.934	1.00	0.00		N
ATOM	3582	CA	ALA A	235	8.936	18.249	4.071	1.00	0.00		C
ATOM	3583	C	ALA A	235	7.445	18.215	3.813	1.00	0.00		C
ATOM	3584	O	ALA A	235	7.035	18.328	2.668	1.00	0.00		O
ATOM	3591	N	LYS A	236	6.961	18.140	5.172	1.00	0.00		N
ATOM	3592	CA	LYS A	236	5.659	18.392	5.652	1.00	0.00		C
ATOM	3593	C	LYS A	236	5.048	19.617	5.870	1.00	0.00		C
ATOM	3594	O	LYS A	236	5.937	20.492	6.092	1.00	0.00		O
ATOM	3613	N	VAL A	237	3.711	19.742	5.640	1.00	0.00		N
ATOM	3614	CA	VAL A	237	2.990	20.415	6.745	1.00	0.00		C
ATOM	3615	C	VAL A	237	1.489	20.259	6.622	1.00	0.00		C
ATOM	3616	O	VAL A	237	0.905	20.730	5.657	1.00	0.00		O
ATOM	3629	N	ASN A	238	0.690	19.804	7.655	1.00	0.00		N
ATOM	3630	CA	ASN A	238	-0.786	19.868	7.627	1.00	0.00		C
ATOM	3631	C	ASN A	238	-1.270	21.223	8.200	1.00	0.00		C
ATOM	3632	O	ASN A	238	-0.675	21.888	9.006	1.00	0.00		O
ATOM	3643	N	ASN A	239	-2.468	21.641	7.785	1.00	0.00		N
ATOM	3644	CA	ASN A	239	-3.218	22.781	8.335	1.00	0.00		C
ATOM	3645	C	ASN A	239	-3.620	22.666	9.806	1.00	0.00		C
ATOM	3646	O	ASN A	239	-3.590	23.698	10.491	1.00	0.00		O
ATOM	3657	N	SER A	240	-3.916	21.445	10.293	1.00	0.00		N
ATOM	3658	CA	SER A	240	-4.038	21.139	11.737	1.00	0.00		C
ATOM	3659	C	SER A	240	-2.745	21.045	12.525	1.00	0.00		C
ATOM	3660	O	SER A	240	-2.901	20.586	13.701	1.00	0.00		O
ATOM	3668	N	SER A	241	-1.508	21.425	12.100	1.00	0.00		N
ATOM	3669	CA	SER A	241	-0.296	21.982	12.696	1.00	0.00		C
ATOM	3670	C	SER A	241	0.835	20.960	12.845	1.00	0.00		C
ATOM	3671	O	SER A	241	1.595	21.043	13.801	1.00	0.00		O
ATOM	3679	N	LEU A	242	1.001	19.950	11.966	1.00	0.00		N
ATOM	3680	CA	LEU A	242	2.066	18.892	12.036	1.00	0.00		C
ATOM	3681	C	LEU A	242	3.091	19.116	10.916	1.00	0.00		C
ATOM	3682	O	LEU A	242	2.845	18.633	9.826	1.00	0.00		O
ATOM	3698	N	ILE A	243	4.259	19.717	11.224	1.00	0.00		N
ATOM	3699	CA	ILE A	243	5.343	19.951	10.246	1.00	0.00		C
ATOM	3700	C	ILE A	243	6.270	18.754	10.210	1.00	0.00		C
ATOM	3701	O	ILE A	243	6.742	18.362	11.278	1.00	0.00		O
ATOM	3717	N	GLY A	244	7.192	18.282	9.452	1.00	0.00		N
ATOM	3718	CA	GLY A	244	7.763	17.049	9.504	1.00	0.00		C
ATOM	3719	C	GLY A	244	8.755	17.415	8.444	1.00	0.00		C
ATOM	3720	O	GLY A	244	8.521	17.833	7.358	1.00	0.00		O
ATOM	3724	N	LEU A	245	9.843	17.548	8.981	1.00	0.00		N
ATOM	3725	CA	LEU A	245	11.196	17.674	8.409	1.00	0.00		C
ATOM	3726	C	LEU A	245	11.731	16.273	8.154	1.00	0.00		C
ATOM	3727	O	LEU A	245	11.545	15.424	9.048	1.00	0.00		O
ATOM	3743	N	GLY A	246	12.376	16.006	7.041	1.00	0.00		N
ATOM	3744	CA	GLY A	246	13.129	14.760	6.940	1.00	0.00		C
ATOM	3745	C	GLY A	246	14.042	14.708	5.708	1.00	0.00		C
ATOM	3746	O	GLY A	246	13.490	14.826	4.600	1.00	0.00		O
ATOM	3750	N	TYR A	247	15.353	14.581	5.909	1.00	0.00		N
ATOM	3751	CA	TYR A	247	16.373	15.057	4.951	1.00	0.00		C
ATOM	3752	C	TYR A	247	17.103	13.829	4.408	1.00	0.00		C
ATOM	3753	O	TYR A	247	17.687	13.185	5.274	1.00	0.00		O
ATOM	3771	N	THR A	248	16.984	13.522	3.125	1.00	0.00		N
ATOM	3772	CA	THR A	248	17.515	12.290	2.508	1.00	0.00		C
ATOM	3773	C	THR A	248	18.890	12.609	1.923	1.00	0.00		C
ATOM	3774	O	THR A	248	18.933	13.714	1.347	1.00	0.00		O
ATOM	3785	N	GLN A	249	19.891	11.722	2.070	1.00	0.00		N
ATOM	3786	CA	GLN A	249	21.195	11.871	1.452	1.00	0.00		C
ATOM	3787	C	GLN A	249	21.595	10.606	0.690	1.00	0.00		C
ATOM	3788	O	GLN A	249	21.900	9.582	1.298	1.00	0.00		O
ATOM	3802	N	THR A	250	21.649	10.637	-0.641	1.00	0.00		N
ATOM	3803	CA	THR A	250	21.845	9.464	-1.524	1.00	0.00		C



## Appendix D PDB files of the models for the closed state of VDAC

ATOM	3804	C	THR A 250	23.290	9.559	-2.041	1.00	0.00	C
ATOM	3805	O	THR A 250	23.530	10.120	-3.073	1.00	0.00	O
ATOM	3816	N	LEU A 251	24.260	8.948	-1.366	1.00	0.00	N
ATOM	3817	CA	LEU A 251	25.667	9.150	-1.745	1.00	0.00	C
ATOM	3818	C	LEU A 251	26.020	8.287	-2.981	1.00	0.00	C
ATOM	3819	O	LEU A 251	25.316	7.270	-3.090	1.00	0.00	O
ATOM	3835	N	LYS A 252	26.975	8.679	-3.876	1.00	0.00	N
ATOM	3836	CA	LYS A 252	27.457	7.928	-5.059	1.00	0.00	C
ATOM	3837	C	LYS A 252	27.893	6.543	-4.578	1.00	0.00	C
ATOM	3838	O	LYS A 252	27.417	5.523	-5.082	1.00	0.00	O
ATOM	3857	N	PRO A 253	28.775	6.484	-3.602	1.00	0.00	N
ATOM	3858	CA	PRO A 253	29.241	5.197	-3.022	1.00	0.00	C
ATOM	3859	C	PRO A 253	28.112	4.161	-2.895	1.00	0.00	C
ATOM	3860	O	PRO A 253	28.376	2.973	-2.731	1.00	0.00	O
ATOM	3871	N	GLY A 254	26.858	4.612	-2.989	1.00	0.00	N
ATOM	3872	CA	GLY A 254	25.719	3.697	-2.905	1.00	0.00	C
ATOM	3873	C	GLY A 254	25.147	3.603	-1.492	1.00	0.00	C
ATOM	3874	O	GLY A 254	24.672	2.542	-1.084	1.00	0.00	O
ATOM	3878	N	ILE A 255	25.191	4.708	-0.755	1.00	0.00	N
ATOM	3879	CA	ILE A 255	24.666	4.714	0.617	1.00	0.00	C
ATOM	3880	C	ILE A 255	23.586	5.784	0.816	1.00	0.00	C
ATOM	3881	O	ILE A 255	23.786	6.951	0.475	1.00	0.00	O
ATOM	3897	N	LYS A 256	22.363	5.388	1.258	1.00	0.00	N
ATOM	3898	CA	LYS A 256	21.252	6.331	1.556	1.00	0.00	C
ATOM	3899	C	LYS A 256	21.290	6.566	3.073	1.00	0.00	C
ATOM	3900	O	LYS A 256	21.392	5.534	3.737	1.00	0.00	O
ATOM	3919	N	LEU A 257	21.258	7.813	3.536	1.00	0.00	N
ATOM	3920	CA	LEU A 257	21.127	8.208	4.937	1.00	0.00	C
ATOM	3921	C	LEU A 257	19.991	9.219	5.092	1.00	0.00	C
ATOM	3922	O	LEU A 257	20.150	10.375	4.708	1.00	0.00	O
ATOM	3938	N	THR A 258	18.820	8.777	5.549	1.00	0.00	N
ATOM	3939	CA	THR A 258	17.678	9.665	5.839	1.00	0.00	C
ATOM	3940	C	THR A 258	17.703	10.046	7.319	1.00	0.00	C
ATOM	3941	O	THR A 258	18.196	9.234	8.082	1.00	0.00	O
ATOM	3952	N	LEU A 259	17.223	11.223	7.658	1.00	0.00	N
ATOM	3953	CA	LEU A 259	16.703	11.600	8.984	1.00	0.00	C
ATOM	3954	C	LEU A 259	15.213	11.844	8.783	1.00	0.00	C
ATOM	3955	O	LEU A 259	14.869	12.292	7.691	1.00	0.00	O
ATOM	3971	N	SER A 260	14.408	11.596	9.811	1.00	0.00	N
ATOM	3972	CA	SER A 260	13.017	12.076	9.892	1.00	0.00	C
ATOM	3973	C	SER A 260	12.684	12.530	11.324	1.00	0.00	C
ATOM	3974	O	SER A 260	13.405	12.211	12.266	1.00	0.00	O
ATOM	3982	N	ALA A 261	11.590	13.263	11.401	1.00	0.00	N
ATOM	3983	CA	ALA A 261	10.973	13.774	12.624	1.00	0.00	C
ATOM	3984	C	ALA A 261	9.584	14.330	12.293	1.00	0.00	C
ATOM	3985	O	ALA A 261	9.441	14.558	11.067	1.00	0.00	O
ATOM	3992	N	LEU A 262	8.591	14.425	13.143	1.00	0.00	N
ATOM	3993	CA	LEU A 262	7.152	14.653	12.845	1.00	0.00	C
ATOM	3994	C	LEU A 262	6.591	15.622	13.895	1.00	0.00	C
ATOM	3995	O	LEU A 262	5.908	15.079	14.792	1.00	0.00	O
ATOM	4011	N	LEU A 263	6.859	16.945	13.882	1.00	0.00	N
ATOM	4012	CA	LEU A 263	6.686	17.746	15.126	1.00	0.00	C
ATOM	4013	C	LEU A 263	5.276	18.390	15.131	1.00	0.00	C
ATOM	4014	O	LEU A 263	5.075	19.197	14.210	1.00	0.00	O
ATOM	4030	N	ASP A 264	4.397	18.085	16.123	1.00	0.00	N
ATOM	4031	CA	ASP A 264	3.024	18.590	16.153	1.00	0.00	C
ATOM	4032	C	ASP A 264	2.911	19.887	16.934	1.00	0.00	C
ATOM	4033	O	ASP A 264	2.945	19.900	18.147	1.00	0.00	O
ATOM	4042	N	GLY A 265	2.781	20.963	16.196	1.00	0.00	N
ATOM	4043	CA	GLY A 265	2.671	22.307	16.742	1.00	0.00	C
ATOM	4044	C	GLY A 265	1.571	22.464	17.781	1.00	0.00	C
ATOM	4045	O	GLY A 265	1.643	23.366	18.619	1.00	0.00	O
ATOM	4049	N	LYS A 266	0.608	21.546	17.767	1.00	0.00	N
ATOM	4050	CA	LYS A 266	-0.468	21.516	18.753	1.00	0.00	C
ATOM	4051	C	LYS A 266	-0.062	20.555	19.853	1.00	0.00	C
ATOM	4052	O	LYS A 266	-0.646	20.563	20.935	1.00	0.00	O
ATOM	4071	N	ASN A 267	1.001	19.771	19.593	1.00	0.00	N
ATOM	4072	CA	ASN A 267	1.490	18.877	20.569	1.00	0.00	C
ATOM	4073	C	ASN A 267	3.027	18.905	20.594	1.00	0.00	C
ATOM	4074	O	ASN A 267	3.619	17.924	20.985	1.00	0.00	O
ATOM	4085	N	VAL A 268	3.692	20.018	20.218	1.00	0.00	N
ATOM	4086	CA	VAL A 268	5.165	20.039	20.320	1.00	0.00	C
ATOM	4087	C	VAL A 268	5.513	19.947	21.742	1.00	0.00	C
ATOM	4088	O	VAL A 268	6.570	19.441	22.123	1.00	0.00	O
ATOM	4101	N	ASN A 269	4.566	20.412	22.510	1.00	0.00	N
ATOM	4102	CA	ASN A 269	4.657	20.356	23.957	1.00	0.00	C
ATOM	4103	C	ASN A 269	3.978	19.101	24.484	1.00	0.00	C
ATOM	4104	O	ASN A 269	4.092	18.804	25.659	1.00	0.00	O
ATOM	4115	N	ALA A 270	3.348	18.309	23.603	1.00	0.00	N
ATOM	4116	CA	ALA A 270	2.801	17.040	24.039	1.00	0.00	C
ATOM	4117	C	ALA A 270	3.723	15.944	23.484	1.00	0.00	C
ATOM	4118	O	ALA A 270	3.963	14.948	24.169	1.00	0.00	O
ATOM	4125	N	GLY A 271	4.293	16.134	22.258	1.00	0.00	N
ATOM	4126	CA	GLY A 271	5.204	15.176	21.726	1.00	0.00	C
ATOM	4127	C	GLY A 271	4.633	14.438	20.546	1.00	0.00	C
ATOM	4128	O	GLY A 271	4.268	15.010	19.520	1.00	0.00	O
ATOM	4132	N	GLY A 272	4.601	13.146	20.707	1.00	0.00	N
ATOM	4133	CA	GLY A 272	4.132	12.265	19.663	1.00	0.00	C
ATOM	4134	C	GLY A 272	4.790	12.671	18.351	1.00	0.00	C
ATOM	4135	O	GLY A 272	4.100	12.926	17.367	1.00	0.00	O
ATOM	4139	N	HIS A 273	6.122	12.623	18.406	1.00	0.00	N
ATOM	4140	CA	HIS A 273	7.147	12.713	17.364	1.00	0.00	C
ATOM	4141	C	HIS A 273	7.661	11.353	16.991	1.00	0.00	C
ATOM	4142	O	HIS A 273	7.522	10.426	17.812	1.00	0.00	O

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ATOM 4156 N LYS A 274      8.198 11.212 15.819 1.00 0.00      N
ATOM 4157 CA LYS A 274     8.685 9.958 15.253 1.00 0.00      C
ATOM 4158 C LYS A 274     10.042 10.256 14.617 1.00 0.00      C
ATOM 4159 O LYS A 274     10.208 10.439 13.419 1.00 0.00      O
ATOM 4178 N LEU A 275     11.009 10.307 15.524 1.00 0.00      N
ATOM 4179 CA LEU A 275    12.448 10.366 15.195 1.00 0.00      C
ATOM 4180 C LEU A 275    12.822 9.083 14.447 1.00 0.00      C
ATOM 4181 O LEU A 275    12.320 8.017 14.847 1.00 0.00      O
ATOM 4197 N GLY A 276     13.628 9.178 13.407 1.00 0.00      N
ATOM 4198 CA GLY A 276    13.847 8.054 12.486 1.00 0.00      C
ATOM 4199 C GLY A 276    15.052 8.280 11.582 1.00 0.00      C
ATOM 4200 O GLY A 276    15.489 9.421 11.434 1.00 0.00      O
ATOM 4204 N LEU A 277    15.563 7.165 11.054 1.00 0.00      N
ATOM 4205 CA LEU A 277   16.916 7.019 10.498 1.00 0.00      C
ATOM 4206 C LEU A 277   16.889 5.895 9.435 1.00 0.00      C
ATOM 4207 O LEU A 277   16.984 4.708 9.730 1.00 0.00      O
ATOM 4223 N GLY A 278   16.736 6.245 8.163 1.00 0.00      N
ATOM 4224 CA GLY A 278   16.513 5.313 7.047 1.00 0.00      C
ATOM 4225 C GLY A 278   17.850 4.961 6.401 1.00 0.00      C
ATOM 4226 O GLY A 278   18.311 5.942 5.761 1.00 0.00      O
ATOM 4230 N LEU A 279   18.404 3.754 6.561 1.00 0.00      N
ATOM 4231 CA LEU A 279  19.786 3.403 6.250 1.00 0.00      C
ATOM 4232 C LEU A 279  19.802 2.302 5.216 1.00 0.00      C
ATOM 4233 O LEU A 279  19.646 1.125 5.610 1.00 0.00      O
ATOM 4249 N GLU A 280   19.961 2.675 3.903 1.00 0.00      N
ATOM 4250 CA GLU A 280  19.841 1.723 2.791 1.00 0.00      C
ATOM 4251 C GLU A 280  21.224 1.552 2.178 1.00 0.00      C
ATOM 4252 O GLU A 280  21.644 2.649 1.713 1.00 0.00      O
ATOM 4264 N PHE A 281  21.834 0.353 2.209 1.00 0.00      N
ATOM 4265 CA PHE A 281 23.152 0.120 1.641 1.00 0.00      C
ATOM 4266 C PHE A 281  23.054 -0.705 0.361 1.00 0.00      C
ATOM 4267 O PHE A 281  22.318 -1.701 0.305 1.00 0.00      O
ATOM 4284 N GLN A 282  23.792 -0.273 -0.664 1.00 0.00      N
ATOM 4285 CA GLN A 282 23.792 -0.958 -1.950 1.00 0.00      C
ATOM 4286 C GLN A 282  24.829 -2.079 -1.958 1.00 0.00      C
ATOM 4287 O GLN A 282  26.031 -1.828 -1.872 1.00 0.00      O
ATOM 4301 N ALA A 283  24.351 -3.317 -2.054 1.00 0.00      N
ATOM 4302 CA ALA A 283 25.236 -4.479 -2.068 1.00 0.00      C
ATOM 4303 C ALA A 283  26.272 -4.394 -0.951 1.00 0.00      C
ATOM 4304 O ALA A 283  26.243 -3.479 -0.128 1.00 0.00      O
ATOM 4311 N ARG A 284  27.187 -5.362 -0.928 1.00 0.00      N
ATOM 4312 CA ARG A 284 28.230 -5.397 0.092 1.00 0.00      C
ATOM 4313 C ARG A 284  29.386 -4.481 -0.296 1.00 0.00      C
ATOM 4314 O ARG A 284  29.687 -4.313 -1.478 1.00 0.00      O
ATOM 4335 N SER A 285  30.030 -3.891 0.710 1.00 0.00      N
ATOM 4336 CA SER A 285 31.155 -2.989 0.468 1.00 0.00      C
ATOM 4337 C SER A 285  32.466 -3.628 0.921 1.00 0.00      C
ATOM 4338 O SER A 285  33.335 -2.895 1.361 1.00 0.00      O
END
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## D.2 Model 2

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HEADER 5jdp
ATOM   1 N  SER A  1    17.791 -1.898 -4.458  1.00  0.00      N
ATOM   2 CA SER A  1    16.470 -2.608 -4.563  1.00  0.00      C
ATOM   3 C  SER A  1    16.126 -3.317 -3.256  1.00  0.00      C
ATOM   4 O  SER A  1    14.899 -3.689 -3.218  1.00  0.00      O
ATOM   14 N  ALA A  2    16.953 -3.488 -2.270  1.00  0.00      N
ATOM   15 CA ALA A  2    16.543 -3.683 -0.880  1.00  0.00      C
ATOM   16 C  ALA A  2    15.605 -2.561 -0.458  1.00  0.00      C
ATOM   17 O  ALA A  2    14.473 -2.972 -0.103  1.00  0.00      O
ATOM   24 N  VAL A  3    16.030 -1.363 -0.592  1.00  0.00      N
ATOM   25 CA VAL A  3    15.407 -0.057 -0.468  1.00  0.00      C
ATOM   26 C  VAL A  3    14.550 -0.074  0.838  1.00  0.00      C
ATOM   27 O  VAL A  3    13.383 -0.346  0.760  1.00  0.00      O
ATOM   40 N  PRO A  4    15.099  0.241  2.018  1.00  0.00      N
ATOM   41 CA PRO A  4    14.364  0.444  3.339  1.00  0.00      C
ATOM   42 C  PRO A  4    13.618  1.786  3.677  1.00  0.00      C
ATOM   43 O  PRO A  4    14.423  2.795  3.699  1.00  0.00      O
ATOM   68 N  THR A  6    11.057  4.212  6.513  1.00  0.00      N
ATOM   69 CA THR A  6    11.127  2.935  7.255  1.00  0.00      C
ATOM   70 C  THR A  6    9.849  2.930  8.114  1.00  0.00      C
ATOM   71 O  THR A  6    9.240  4.004  8.294  1.00  0.00      O
ATOM   82 N  TYR A  7    9.513  1.781  8.606  1.00  0.00      N
ATOM   83 CA TYR A  7    8.274  1.668  9.380  1.00  0.00      C
ATOM   84 C  TYR A  7    8.355  2.358 10.761  1.00  0.00      C
ATOM   85 O  TYR A  7    7.313  2.883 11.190  1.00  0.00      O
ATOM   103 N  ALA A  8    9.461  2.332 11.463  1.00  0.00      N
ATOM   104 CA ALA A  8    9.402  2.347 12.935  1.00  0.00      C
ATOM   105 C  ALA A  8    9.106  3.734 13.567  1.00  0.00      C
ATOM   106 O  ALA A  8    8.146  3.715 14.378  1.00  0.00      O
ATOM   113 N  ASP A  9    9.821  4.736 13.198  1.00  0.00      N
ATOM   114 CA ASP A  9    9.618  6.112 13.718  1.00  0.00      C
ATOM   115 C  ASP A  9    8.675  6.925 12.802  1.00  0.00      C
ATOM   116 O  ASP A  9    9.035  7.992 12.264  1.00  0.00      O
ATOM   125 N  LEU A 10   7.428  6.409 12.653  1.00  0.00      N
ATOM   126 CA LEU A 10   6.482  7.002 11.713  1.00  0.00      C
ATOM   127 C  LEU A 10   5.948  8.271 12.356  1.00  0.00      C
ATOM   128 O  LEU A 10   5.037  8.172 13.195  1.00  0.00      O
ATOM   144 N  GLY A 11   6.517  9.404 11.997  1.00  0.00      N
ATOM   145 CA GLY A 11   6.073 10.713 12.464  1.00  0.00      C
ATOM   146 C  GLY A 11   6.774 11.072 13.782  1.00  0.00      C
ATOM   147 O  GLY A 11   6.043 11.476 14.699  1.00  0.00      O
ATOM   151 N  LYS A 12   8.103 10.968 13.842  1.00  0.00      N
ATOM   152 CA LYS A 12   8.883 11.402 15.023  1.00  0.00      C
ATOM   153 C  LYS A 12   9.327 12.885 14.911  1.00  0.00      C
ATOM   154 O  LYS A 12   10.288 13.385 15.492  1.00  0.00      O
ATOM   173 N  SER A 13   8.546 13.791 14.250  1.00  0.00      N
ATOM   174 CA SER A 13   8.292 15.139 14.830  1.00  0.00      C
ATOM   175 C  SER A 13   6.824 15.596 14.720  1.00  0.00      C
ATOM   176 O  SER A 13   6.566 16.808 14.933  1.00  0.00      O
ATOM   184 N  ALA A 14   5.821 14.741 14.635  1.00  0.00      N
ATOM   185 CA ALA A 14   4.652 14.768 15.513  1.00  0.00      C
ATOM   186 C  ALA A 14   5.021 14.222 16.888  1.00  0.00      C
ATOM   187 O  ALA A 14   4.343 14.749 17.811  1.00  0.00      O
ATOM   194 N  ARG A 15   5.945 13.314 17.053  1.00  0.00      N
ATOM   195 CA ARG A 15   6.135 12.496 18.250  1.00  0.00      C
ATOM   196 C  ARG A 15   7.641 12.366 18.497  1.00  0.00      C
ATOM   197 O  ARG A 15   8.121 11.193 18.568  1.00  0.00      O
ATOM   218 N  ASP A 16   8.554 13.315 18.444  1.00  0.00      N
ATOM   219 CA ASP A 16   8.566 14.630 19.133  1.00  0.00      C
ATOM   220 C  ASP A 16   7.406 15.514 18.782  1.00  0.00      C
ATOM   221 O  ASP A 16   7.195 16.186 17.799  1.00  0.00      O
ATOM   230 N  VAL A 17   6.409 15.927 19.551  1.00  0.00      N
ATOM   231 CA VAL A 17   6.634 16.331 20.927  1.00  0.00      C
ATOM   232 C  VAL A 17   6.495 15.117 21.861  1.00  0.00      C
ATOM   233 O  VAL A 17   7.548 14.935 22.464  1.00  0.00      O
ATOM   246 N  PHE A 18   5.415 14.421 21.946  1.00  0.00      N
ATOM   247 CA PHE A 18   4.799 13.560 22.985  1.00  0.00      C
ATOM   248 C  PHE A 18   5.666 13.239 24.233  1.00  0.00      C
ATOM   249 O  PHE A 18   5.178 13.367 25.351  1.00  0.00      O
ATOM   266 N  THR A 19   6.944 12.941 23.997  1.00  0.00      N
ATOM   267 CA THR A 19   8.003 13.043 25.015  1.00  0.00      C
ATOM   268 C  THR A 19   8.558 14.481 25.152  1.00  0.00      C
ATOM   269 O  THR A 19   7.642 15.331 25.299  1.00  0.00      O
ATOM   280 N  LYS A 20   9.788 14.860 25.180  1.00  0.00      N
ATOM   281 CA LYS A 20   10.372 15.983 26.023  1.00  0.00      C
ATOM   282 C  LYS A 20   9.715 16.230 27.416  1.00  0.00      C
ATOM   283 O  LYS A 20   9.284 17.221 27.996  1.00  0.00      O
ATOM   302 N  GLY A 21   10.408 15.220 27.805  1.00  0.00      N
ATOM   303 CA GLY A 21   11.228 15.266 29.039  1.00  0.00      C
ATOM   304 C  GLY A 21   10.820 14.100 30.039  1.00  0.00      C
ATOM   305 O  GLY A 21   9.786 13.915 29.567  1.00  0.00      O
ATOM   309 N  TYR A 22   11.954 14.182 30.622  1.00  0.00      N
ATOM   310 CA TYR A 22   12.815 13.027 30.944  1.00  0.00      C
ATOM   311 C  TYR A 22   13.534 12.575 29.673  1.00  0.00      C
ATOM   312 O  TYR A 22   12.872 12.057 28.765  1.00  0.00      O
ATOM   330 N  GLY A 23   14.846 12.772 29.595  1.00  0.00      N
ATOM   331 CA GLY A 23   15.630 12.296 28.454  1.00  0.00      C
ATOM   332 C  GLY A 23   15.433 13.203 27.243  1.00  0.00      C
ATOM   333 O  GLY A 23   14.557 12.825 26.429  1.00  0.00      O
ATOM   337 N  PHE A 24   16.234 14.279 27.226  1.00  0.00      N

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ATOM	338	CA	PHE	A	24	16.407	14.790	25.841	1.00	0.00	C
ATOM	339	C	PHE	A	24	17.202	13.765	24.981	1.00	0.00	C
ATOM	340	O	PHE	A	24	17.943	12.891	25.403	1.00	0.00	O
ATOM	357	N	GLY	A	25	16.746	14.138	23.806	1.00	0.00	N
ATOM	358	CA	GLY	A	25	17.212	12.519	23.025	1.00	0.00	C
ATOM	359	C	GLY	A	25	17.044	11.000	23.083	1.00	0.00	C
ATOM	360	O	GLY	A	25	17.924	11.018	22.224	1.00	0.00	O
ATOM	364	N	LEU	A	26	16.763	9.273	23.204	1.00	0.00	N
ATOM	365	CA	LEU	A	26	17.860	8.427	22.757	1.00	0.00	C
ATOM	366	C	LEU	A	26	17.591	8.024	21.319	1.00	0.00	C
ATOM	367	O	LEU	A	26	16.449	7.781	20.933	1.00	0.00	O
ATOM	383	N	ILE	A	27	18.561	7.547	21.007	1.00	0.00	N
ATOM	384	CA	ILE	A	27	18.970	7.585	19.726	1.00	0.00	C
ATOM	385	C	ILE	A	27	18.893	6.062	19.439	1.00	0.00	C
ATOM	386	O	ILE	A	27	18.980	5.299	20.455	1.00	0.00	O
ATOM	402	N	LYS	A	28	18.590	5.516	18.084	1.00	0.00	N
ATOM	403	CA	LYS	A	28	18.557	4.074	17.867	1.00	0.00	C
ATOM	404	C	LYS	A	28	20.005	4.089	18.345	1.00	0.00	C
ATOM	405	O	LYS	A	28	20.990	4.283	17.619	1.00	0.00	O
ATOM	424	N	LEU	A	29	20.865	3.186	18.841	1.00	0.00	N
ATOM	425	CA	LEU	A	29	22.279	2.759	18.950	1.00	0.00	C
ATOM	426	C	LEU	A	29	22.419	1.295	18.544	1.00	0.00	C
ATOM	427	O	LEU	A	29	21.423	0.604	18.519	1.00	0.00	O
ATOM	443	N	ASP	A	30	23.661	0.848	18.297	1.00	0.00	N
ATOM	444	CA	ASP	A	30	23.944	-0.578	18.097	1.00	0.00	C
ATOM	445	C	ASP	A	30	25.432	-0.914	18.286	1.00	0.00	C
ATOM	446	O	ASP	A	30	26.226	-0.017	17.988	1.00	0.00	O
ATOM	455	N	LEU	A	31	25.748	-2.124	18.725	1.00	0.00	N
ATOM	456	CA	LEU	A	31	27.106	-2.612	19.046	1.00	0.00	C
ATOM	457	C	LEU	A	31	27.312	-3.990	18.380	1.00	0.00	C
ATOM	458	O	LEU	A	31	26.491	-4.848	18.731	1.00	0.00	O
ATOM	474	N	LYS	A	32	28.315	-4.182	17.517	1.00	0.00	N
ATOM	475	CA	LYS	A	32	28.615	-5.471	16.840	1.00	0.00	C
ATOM	476	C	LYS	A	32	29.758	-6.200	17.563	1.00	0.00	C
ATOM	477	O	LYS	A	32	30.699	-5.466	17.907	1.00	0.00	O
ATOM	496	N	THR	A	33	29.660	-7.536	17.731	1.00	0.00	N
ATOM	497	CA	THR	A	33	30.732	-8.419	18.194	1.00	0.00	C
ATOM	498	C	THR	A	33	30.867	-9.594	17.221	1.00	0.00	C
ATOM	499	O	THR	A	33	29.939	-10.398	17.059	1.00	0.00	O
ATOM	510	N	LYS	A	34	32.009	-9.683	16.557	1.00	0.00	N
ATOM	511	CA	LYS	A	34	32.209	-10.750	15.587	1.00	0.00	C
ATOM	512	C	LYS	A	34	32.626	-12.053	16.246	1.00	0.00	C
ATOM	513	O	LYS	A	34	33.691	-12.139	16.856	1.00	0.00	O
ATOM	532	N	SER	A	35	31.802	-13.080	16.057	1.00	0.00	N
ATOM	533	CA	SER	A	35	32.123	-14.395	16.579	1.00	0.00	C
ATOM	534	C	SER	A	35	33.035	-15.093	15.577	1.00	0.00	C
ATOM	535	O	SER	A	35	32.984	-14.806	14.382	1.00	0.00	O
ATOM	543	N	GLU	A	36	33.872	-16.001	16.049	1.00	0.00	N
ATOM	544	CA	GLU	A	36	34.767	-16.695	15.144	1.00	0.00	C
ATOM	545	C	GLU	A	36	34.043	-17.862	14.479	1.00	0.00	C
ATOM	546	O	GLU	A	36	34.308	-18.184	13.321	1.00	0.00	O
ATOM	558	N	ASN	A	37	33.094	-18.466	15.199	1.00	0.00	N
ATOM	559	CA	ASN	A	37	32.314	-19.561	14.632	1.00	0.00	C
ATOM	560	C	ASN	A	37	31.483	-19.039	13.457	1.00	0.00	C
ATOM	561	O	ASN	A	37	31.366	-19.692	12.417	1.00	0.00	O
ATOM	572	N	GLY	A	38	30.910	-17.850	13.642	1.00	0.00	N
ATOM	573	CA	GLY	A	38	30.096	-17.228	12.617	1.00	0.00	C
ATOM	574	C	GLY	A	38	28.741	-16.827	13.179	1.00	0.00	C
ATOM	575	O	GLY	A	38	27.821	-16.571	12.414	1.00	0.00	O
ATOM	579	N	LEU	A	39	28.632	-16.794	14.514	1.00	0.00	N
ATOM	580	CA	LEU	A	39	27.377	-16.447	15.187	1.00	0.00	C
ATOM	581	C	LEU	A	39	26.957	-15.009	14.895	1.00	0.00	C
ATOM	582	O	LEU	A	39	25.776	-14.728	14.703	1.00	0.00	O
ATOM	598	N	GLU	A	40	28.044	-14.015	15.029	1.00	0.00	N
ATOM	599	CA	GLU	A	40	27.785	-12.625	14.649	1.00	0.00	C
ATOM	600	C	GLU	A	40	26.710	-12.006	15.558	1.00	0.00	C
ATOM	601	O	GLU	A	40	25.688	-11.481	15.104	1.00	0.00	O
ATOM	613	N	PHE	A	41	26.937	-12.054	16.879	1.00	0.00	N
ATOM	614	CA	PHE	A	41	26.085	-11.343	17.840	1.00	0.00	C
ATOM	615	C	PHE	A	41	26.219	-9.836	17.696	1.00	0.00	C
ATOM	616	O	PHE	A	41	27.334	-9.340	17.864	1.00	0.00	O
ATOM	633	N	THR	A	42	25.119	-9.168	17.440	1.00	0.00	N
ATOM	634	CA	THR	A	42	24.988	-7.714	17.392	1.00	0.00	C
ATOM	635	C	THR	A	42	23.785	-7.341	18.258	1.00	0.00	C
ATOM	636	O	THR	A	42	22.693	-7.710	17.757	1.00	0.00	O
ATOM	647	N	SER	A	43	23.959	-6.690	19.376	1.00	0.00	N
ATOM	648	CA	SER	A	43	22.905	-6.274	20.314	1.00	0.00	C
ATOM	649	C	SER	A	43	22.777	-4.758	20.236	1.00	0.00	C
ATOM	650	O	SER	A	43	23.826	-4.125	20.013	1.00	0.00	O
ATOM	658	N	SER	A	44	21.595	-4.255	20.476	1.00	0.00	N
ATOM	659	CA	SER	A	44	21.214	-2.830	20.411	1.00	0.00	C
ATOM	660	C	SER	A	44	20.483	-2.370	21.672	1.00	0.00	C
ATOM	661	O	SER	A	44	19.964	-3.181	22.441	1.00	0.00	O
ATOM	669	N	GLY	A	45	20.506	-1.051	21.836	1.00	0.00	N
ATOM	670	CA	GLY	A	45	20.188	-0.332	23.066	1.00	0.00	C
ATOM	671	C	GLY	A	45	19.537	0.987	22.671	1.00	0.00	C
ATOM	672	O	GLY	A	45	20.142	2.054	22.803	1.00	0.00	O
ATOM	676	N	SER	A	46	18.343	0.863	22.144	1.00	0.00	N
ATOM	677	CA	SER	A	46	17.763	1.826	21.219	1.00	0.00	C
ATOM	678	C	SER	A	46	16.679	2.644	21.892	1.00	0.00	C
ATOM	679	O	SER	A	46	16.568	2.575	23.108	1.00	0.00	O
ATOM	687	N	ALA	A	47	15.875	3.328	21.120	1.00	0.00	N
ATOM	688	CA	ALA	A	47	14.531	3.693	21.494	1.00	0.00	C
ATOM	689	C	ALA	A	47	13.614	3.651	20.261	1.00	0.00	C

## Appendix D PDB files of the models for the closed state of VDAC

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ATOM	690	O	ALA A	47	13.954	4.381	19.314	1.00	0.00		O
ATOM	697	N	ASN A	48	12.562	2.887	20.275	1.00	0.00		N
ATOM	698	CA	ASN A	48	11.478	2.851	19.283	1.00	0.00		C
ATOM	699	C	ASN A	48	10.320	3.620	19.907	1.00	0.00		C
ATOM	700	O	ASN A	48	9.362	3.017	20.415	1.00	0.00		O
ATOM	711	N	THR A	49	10.430	4.958	19.928	1.00	0.00		N
ATOM	712	CA	THR A	49	9.577	5.752	20.809	1.00	0.00		C
ATOM	713	C	THR A	49	8.137	5.852	20.294	1.00	0.00		C
ATOM	714	O	THR A	49	7.778	6.758	19.541	1.00	0.00		O
ATOM	725	N	GLU A	50	7.295	4.921	20.761	1.00	0.00		N
ATOM	726	CA	GLU A	50	5.867	5.224	20.920	1.00	0.00		C
ATOM	727	C	GLU A	50	5.333	4.499	22.164	1.00	0.00		C
ATOM	728	O	GLU A	50	4.368	3.747	22.184	1.00	0.00		O
ATOM	740	N	THR A	51	6.082	4.656	23.267	1.00	0.00		N
ATOM	741	CA	THR A	51	5.981	3.901	24.535	1.00	0.00		C
ATOM	742	C	THR A	51	6.597	4.695	25.706	1.00	0.00		C
ATOM	743	O	THR A	51	5.786	5.171	26.508	1.00	0.00		O
ATOM	754	N	THR A	52	7.892	4.857	25.798	1.00	0.00		N
ATOM	755	CA	THR A	52	8.741	5.535	26.819	1.00	0.00		C
ATOM	756	C	THR A	52	10.194	5.358	26.366	1.00	0.00		C
ATOM	757	O	THR A	52	10.624	6.393	25.729	1.00	0.00		O
ATOM	768	N	LYS A	53	11.013	4.357	26.356	1.00	0.00		N
ATOM	769	CA	LYS A	53	11.603	3.174	26.995	1.00	0.00		C
ATOM	770	C	LYS A	53	13.128	3.141	26.744	1.00	0.00		C
ATOM	771	O	LYS A	53	13.624	4.064	26.082	1.00	0.00		O
ATOM	790	N	VAL A	54	13.823	2.073	27.158	1.00	0.00		N
ATOM	791	CA	VAL A	54	14.957	1.481	26.480	1.00	0.00		C
ATOM	792	C	VAL A	54	14.471	0.123	25.895	1.00	0.00		C
ATOM	793	O	VAL A	54	14.666	-0.924	26.531	1.00	0.00		O
ATOM	806	N	THR A	55	13.877	0.121	24.696	1.00	0.00		N
ATOM	807	CA	THR A	55	13.765	-1.107	23.861	1.00	0.00		C
ATOM	808	C	THR A	55	15.127	-1.398	23.225	1.00	0.00		C
ATOM	809	O	THR A	55	16.016	-0.562	23.285	1.00	0.00		O
ATOM	820	N	GLY A	56	15.266	-2.567	22.609	1.00	0.00		N
ATOM	821	CA	GLY A	56	16.510	-2.995	21.953	1.00	0.00		C
ATOM	822	C	GLY A	56	16.244	-4.015	20.854	1.00	0.00		C
ATOM	823	O	GLY A	56	15.139	-4.107	20.317	1.00	0.00		O
ATOM	827	N	SER A	57	17.270	-4.779	20.551	1.00	0.00		N
ATOM	828	CA	SER A	57	17.164	-5.986	19.726	1.00	0.00		C
ATOM	829	C	SER A	57	18.392	-6.865	19.935	1.00	0.00		C
ATOM	830	O	SER A	57	19.370	-6.383	20.536	1.00	0.00		O
ATOM	838	N	LEU A	58	18.292	-8.072	19.447	1.00	0.00		N
ATOM	839	CA	LEU A	58	19.362	-9.062	19.416	1.00	0.00		C
ATOM	840	C	LEU A	58	19.400	-9.672	18.008	1.00	0.00		C
ATOM	841	O	LEU A	58	18.470	-10.458	17.717	1.00	0.00		O
ATOM	857	N	GLU A	59	20.364	-9.326	17.192	1.00	0.00		N
ATOM	858	CA	GLU A	59	20.591	-9.839	15.837	1.00	0.00		C
ATOM	859	C	GLU A	59	21.734	-10.851	15.933	1.00	0.00		C
ATOM	860	O	GLU A	59	22.783	-10.429	16.469	1.00	0.00		O
ATOM	872	N	THR A	60	21.544	-12.074	15.504	1.00	0.00		N
ATOM	873	CA	THR A	60	22.458	-13.224	15.691	1.00	0.00		C
ATOM	874	C	THR A	60	22.415	-14.075	14.414	1.00	0.00		C
ATOM	875	O	THR A	60	21.344	-14.636	14.208	1.00	0.00		O
ATOM	886	N	LYS A	61	23.474	-14.142	13.616	1.00	0.00		N
ATOM	887	CA	LYS A	61	23.368	-14.509	12.197	1.00	0.00		C
ATOM	888	C	LYS A	61	24.329	-15.650	11.852	1.00	0.00		C
ATOM	889	O	LYS A	61	25.402	-15.743	12.437	1.00	0.00		O
ATOM	908	N	TYR A	62	24.161	-16.867	12.240	1.00	0.00		N
ATOM	909	CA	TYR A	62	25.070	-18.020	12.201	1.00	0.00		C
ATOM	910	C	TYR A	62	25.386	-18.390	10.751	1.00	0.00		C
ATOM	911	O	TYR A	62	24.459	-18.579	9.953	1.00	0.00		O
ATOM	929	N	ARG A	63	26.693	-18.494	10.429	1.00	0.00		N
ATOM	930	CA	ARG A	63	27.141	-18.866	9.066	1.00	0.00		C
ATOM	931	C	ARG A	63	27.082	-17.684	8.086	1.00	0.00		C
ATOM	932	O	ARG A	63	27.828	-16.734	8.262	1.00	0.00		O
ATOM	953	N	TRP A	64	26.175	-17.767	7.084	1.00	0.00		N
ATOM	954	CA	TRP A	64	25.965	-16.715	6.064	1.00	0.00		C
ATOM	955	C	TRP A	64	25.540	-17.399	4.764	1.00	0.00		C
ATOM	956	O	TRP A	64	25.170	-18.572	4.773	1.00	0.00		O
ATOM	977	N	THR A	65	25.577	-16.667	3.653	1.00	0.00		N
ATOM	978	CA	THR A	65	25.181	-17.232	2.373	1.00	0.00		C
ATOM	979	C	THR A	65	26.305	-18.065	1.762	1.00	0.00		C
ATOM	980	O	THR A	65	26.267	-18.369	0.573	1.00	0.00		O
ATOM	991	N	GLU A	66	27.302	-18.440	2.575	1.00	0.00		N
ATOM	992	CA	GLU A	66	28.412	-19.252	2.069	1.00	0.00		C
ATOM	993	C	GLU A	66	27.874	-20.397	1.227	1.00	0.00		C
ATOM	994	O	GLU A	66	27.582	-20.219	0.042	1.00	0.00		O
ATOM	1006	N	TYR A	67	27.742	-21.584	1.824	1.00	0.00		N
ATOM	1007	CA	TYR A	67	27.237	-22.724	1.094	1.00	0.00		C
ATOM	1008	C	TYR A	67	25.714	-22.764	1.186	1.00	0.00		C
ATOM	1009	O	TYR A	67	25.100	-23.821	1.037	1.00	0.00		O
ATOM	1027	N	GLY A	68	25.115	-21.582	1.369	1.00	0.00		N
ATOM	1028	CA	GLY A	68	23.660	-21.465	1.405	1.00	0.00		C
ATOM	1029	C	GLY A	68	23.060	-21.923	2.729	1.00	0.00		C
ATOM	1030	O	GLY A	68	21.861	-22.178	2.795	1.00	0.00		O
ATOM	1034	N	LEU A	69	23.891	-22.035	3.769	1.00	0.00		N
ATOM	1035	CA	LEU A	69	23.413	-22.469	5.079	1.00	0.00		C
ATOM	1036	C	LEU A	69	23.347	-21.305	6.054	1.00	0.00		C
ATOM	1037	O	LEU A	69	24.080	-21.267	7.042	1.00	0.00		O
ATOM	1053	N	THR A	70	22.439	-20.382	5.784	1.00	0.00		N
ATOM	1054	CA	THR A	70	22.261	-19.231	6.663	1.00	0.00		C
ATOM	1055	C	THR A	70	21.207	-19.521	7.710	1.00	0.00		C
ATOM	1056	O	THR A	70	20.025	-19.656	7.376	1.00	0.00		O
ATOM	1067	N	PHE A	71	21.622	-19.591	8.981	1.00	0.00		N

ATOM	1068	CA	PHE	A	71	20.665	-19.818	10.050	1.00	0.00	C
ATOM	1069	C	PHE	A	71	20.575	-18.516	10.843	1.00	0.00	C
ATOM	1070	O	PHE	A	71	21.415	-18.220	11.693	1.00	0.00	O
ATOM	1087	N	THR	A	72	19.575	-17.723	10.509	1.00	0.00	N
ATOM	1088	CA	THR	A	72	19.399	-16.419	11.138	1.00	0.00	C
ATOM	1089	C	THR	A	72	18.567	-16.527	12.403	1.00	0.00	C
ATOM	1090	O	THR	A	72	17.559	-17.225	12.435	1.00	0.00	O
ATOM	1101	N	VAL	A	73	18.990	-15.794	13.434	1.00	0.00	N
ATOM	1102	CA	VAL	A	73	18.263	-15.780	14.695	1.00	0.00	C
ATOM	1103	C	VAL	A	73	18.001	-14.343	15.147	1.00	0.00	C
ATOM	1104	O	VAL	A	73	18.926	-13.557	15.348	1.00	0.00	O
ATOM	1117	N	LYS	A	74	16.734	-13.951	15.304	1.00	0.00	N
ATOM	1118	CA	LYS	A	74	16.244	-12.610	15.694	1.00	0.00	C
ATOM	1119	C	LYS	A	74	15.360	-12.709	16.945	1.00	0.00	C
ATOM	1120	O	LYS	A	74	14.184	-13.100	16.826	1.00	0.00	O
ATOM	1139	N	TRP	A	75	15.912	-12.406	18.113	1.00	0.00	N
ATOM	1140	CA	TRP	A	75	15.183	-12.186	19.380	1.00	0.00	C
ATOM	1141	C	TRP	A	75	15.048	-10.664	19.599	1.00	0.00	C
ATOM	1142	O	TRP	A	75	15.890	-9.912	19.106	1.00	0.00	O
ATOM	1163	N	ASN	A	76	14.008	-10.258	20.316	1.00	0.00	N
ATOM	1164	CA	ASN	A	76	13.713	-8.871	20.745	1.00	0.00	C
ATOM	1165	C	ASN	A	76	13.146	-8.848	22.164	1.00	0.00	C
ATOM	1166	O	ASN	A	76	12.726	-9.862	22.713	1.00	0.00	O
ATOM	1177	N	THR	A	77	13.145	-7.647	22.757	1.00	0.00	N
ATOM	1178	CA	THR	A	77	12.639	-7.444	24.105	1.00	0.00	C
ATOM	1179	C	THR	A	77	11.120	-7.639	24.219	1.00	0.00	C
ATOM	1180	O	THR	A	77	10.718	-8.115	25.299	1.00	0.00	O
ATOM	1191	N	ASP	A	78	10.308	-7.325	23.211	1.00	0.00	N
ATOM	1192	CA	ASP	A	78	8.891	-7.730	23.166	1.00	0.00	C
ATOM	1193	C	ASP	A	78	8.641	-9.253	23.291	1.00	0.00	C
ATOM	1194	O	ASP	A	78	7.425	-9.489	23.540	1.00	0.00	O
ATOM	1203	N	ASN	A	79	9.547	-10.155	23.166	1.00	0.00	N
ATOM	1204	CA	ASN	A	79	9.635	-11.624	23.357	1.00	0.00	C
ATOM	1205	C	ASN	A	79	9.497	-12.392	22.020	1.00	0.00	C
ATOM	1206	O	ASN	A	79	9.229	-13.605	22.042	1.00	0.00	O
ATOM	1217	N	THR	A	80	9.707	-11.805	20.853	1.00	0.00	N
ATOM	1218	CA	THR	A	80	9.439	-12.480	19.555	1.00	0.00	C
ATOM	1219	C	THR	A	80	10.621	-13.406	19.294	1.00	0.00	C
ATOM	1220	O	THR	A	80	11.704	-13.273	19.832	1.00	0.00	O
ATOM	1231	N	LEU	A	81	10.707	-14.298	19.057	1.00	0.00	N
ATOM	1232	CA	LEU	A	81	11.877	-15.117	18.758	1.00	0.00	C
ATOM	1233	C	LEU	A	81	11.757	-15.621	17.324	1.00	0.00	C
ATOM	1234	O	LEU	A	81	10.696	-16.116	16.952	1.00	0.00	O
ATOM	1250	N	GLY	A	82	12.807	-15.498	16.511	1.00	0.00	N
ATOM	1251	CA	GLY	A	82	12.714	-15.963	15.129	1.00	0.00	C
ATOM	1252	C	GLY	A	82	13.982	-16.676	14.679	1.00	0.00	C
ATOM	1253	O	GLY	A	82	15.055	-16.097	14.707	1.00	0.00	O
ATOM	1257	N	THR	A	83	13.836	-17.917	14.215	1.00	0.00	N
ATOM	1258	CA	THR	A	83	14.977	-18.682	13.705	1.00	0.00	C
ATOM	1259	C	THR	A	83	14.677	-19.176	12.290	1.00	0.00	C
ATOM	1260	O	THR	A	83	13.685	-19.872	12.063	1.00	0.00	O
ATOM	1271	N	GLU	A	84	15.523	-18.794	11.337	1.00	0.00	N
ATOM	1272	CA	GLU	A	84	15.319	-19.192	9.940	1.00	0.00	C
ATOM	1273	C	GLU	A	84	16.489	-20.010	9.401	1.00	0.00	C
ATOM	1274	O	GLU	A	84	17.563	-19.463	9.170	1.00	0.00	O
ATOM	1286	N	ILE	A	85	16.275	-21.307	9.145	1.00	0.00	N
ATOM	1287	CA	ILE	A	85	17.343	-22.140	8.580	1.00	0.00	C
ATOM	1288	C	ILE	A	85	17.093	-22.337	7.091	1.00	0.00	C
ATOM	1289	O	ILE	A	85	16.081	-22.912	6.707	1.00	0.00	O
ATOM	1305	N	THR	A	86	18.011	-21.830	6.266	1.00	0.00	N
ATOM	1306	CA	THR	A	86	17.859	-21.926	4.814	1.00	0.00	C
ATOM	1307	C	THR	A	86	18.975	-22.717	4.138	1.00	0.00	C
ATOM	1308	O	THR	A	86	20.156	-22.454	4.371	1.00	0.00	O
ATOM	1319	N	VAL	A	87	18.584	-23.639	3.245	1.00	0.00	N
ATOM	1320	CA	VAL	A	87	19.559	-24.410	2.469	1.00	0.00	C
ATOM	1321	C	VAL	A	87	19.334	-24.141	0.984	1.00	0.00	C
ATOM	1322	O	VAL	A	87	18.408	-24.684	0.381	1.00	0.00	O
ATOM	1335	N	GLU	A	88	20.174	-23.299	0.399	1.00	0.00	N
ATOM	1336	CA	GLU	A	88	20.045	-22.963	-1.022	1.00	0.00	C
ATOM	1337	C	GLU	A	88	20.956	-23.842	-1.893	1.00	0.00	C
ATOM	1338	O	GLU	A	88	21.983	-24.323	-1.420	1.00	0.00	O
ATOM	1350	N	ASP	A	89	20.579	-24.026	-3.168	1.00	0.00	N
ATOM	1351	CA	ASP	A	89	21.386	-24.821	-4.114	1.00	0.00	C
ATOM	1352	C	ASP	A	89	21.525	-26.280	-3.673	1.00	0.00	C
ATOM	1353	O	ASP	A	89	22.609	-26.854	-3.778	1.00	0.00	O
ATOM	1362	N	GLN	A	90	20.462	-26.878	-3.155	1.00	0.00	N
ATOM	1363	CA	GLN	A	90	20.569	-28.261	-2.693	1.00	0.00	C
ATOM	1364	C	GLN	A	90	20.738	-29.271	-3.835	1.00	0.00	C
ATOM	1365	O	GLN	A	90	21.512	-30.218	-3.701	1.00	0.00	O
ATOM	1379	N	LEU	A	91	20.046	-29.081	-4.953	1.00	0.00	N
ATOM	1380	CA	LEU	A	91	20.196	-30.014	-6.071	1.00	0.00	C
ATOM	1381	C	LEU	A	91	21.335	-29.537	-6.972	1.00	0.00	C
ATOM	1382	O	LEU	A	91	22.388	-30.177	-7.064	1.00	0.00	O
ATOM	1398	N	ALA	A	92	21.117	-28.398	-7.622	1.00	0.00	N
ATOM	1399	CA	ALA	A	92	22.129	-27.824	-8.503	1.00	0.00	C
ATOM	1400	C	ALA	A	92	21.624	-26.555	-9.206	1.00	0.00	C
ATOM	1401	O	ALA	A	92	22.357	-25.571	-9.311	1.00	0.00	O
ATOM	1408	N	ARG	A	93	20.370	-26.556	-9.667	1.00	0.00	N
ATOM	1409	CA	ARG	A	93	19.831	-25.378	-10.328	1.00	0.00	C
ATOM	1410	C	ARG	A	93	19.479	-24.334	-9.283	1.00	0.00	C
ATOM	1411	O	ARG	A	93	20.232	-23.392	-9.035	1.00	0.00	O
ATOM	1432	N	GLY	A	94	18.323	-24.531	-8.668	1.00	0.00	N
ATOM	1433	CA	GLY	A	94	17.840	-23.635	-7.639	1.00	0.00	C
ATOM	1434	C	GLY	A	94	16.872	-24.372	-6.720	1.00	0.00	C

## Appendix D PDB files of the models for the closed state of VDAC

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ATOM	1435	O	GLY	A	94	15.679	-24.492	-6.996	1.00	0.00	O
ATOM	1439	N	LEU	A	95	17.417	-24.835	-5.619	1.00	0.00	N
ATOM	1440	CA	LEU	A	95	16.638	-25.544	-4.605	1.00	0.00	C
ATOM	1441	C	LEU	A	95	16.855	-24.863	-3.247	1.00	0.00	C
ATOM	1442	O	LEU	A	95	17.992	-24.743	-2.781	1.00	0.00	O
ATOM	1458	N	LYS	A	96	15.754	-24.378	-2.655	1.00	0.00	N
ATOM	1459	CA	LYS	A	96	15.801	-23.658	-1.374	1.00	0.00	C
ATOM	1460	C	LYS	A	96	14.879	-24.323	-0.349	1.00	0.00	C
ATOM	1461	O	LYS	A	96	13.700	-24.533	-0.631	1.00	0.00	O
ATOM	1480	N	LEU	A	97	15.405	-24.639	0.845	1.00	0.00	N
ATOM	1481	CA	LEU	A	97	14.588	-25.264	1.888	1.00	0.00	C
ATOM	1482	C	LEU	A	97	14.677	-24.431	3.149	1.00	0.00	C
ATOM	1483	O	LEU	A	97	15.722	-24.435	3.801	1.00	0.00	O
ATOM	1499	N	THR	A	98	13.601	-23.695	3.476	1.00	0.00	N
ATOM	1500	CA	THR	A	98	13.632	-22.841	4.662	1.00	0.00	C
ATOM	1501	C	THR	A	98	12.739	-23.288	5.813	1.00	0.00	C
ATOM	1502	O	THR	A	98	11.542	-22.979	5.834	1.00	0.00	O
ATOM	1513	N	PHE	A	99	13.329	-23.940	6.814	1.00	0.00	N
ATOM	1514	CA	PHE	A	99	12.550	-24.291	7.982	1.00	0.00	C
ATOM	1515	C	PHE	A	99	12.559	-23.046	8.838	1.00	0.00	C
ATOM	1516	O	PHE	A	99	13.623	-22.469	9.076	1.00	0.00	O
ATOM	1533	N	ASP	A	100	11.390	-22.595	9.243	1.00	0.00	N
ATOM	1534	CA	ASP	A	100	11.288	-21.370	10.005	1.00	0.00	C
ATOM	1535	C	ASP	A	100	10.671	-21.636	11.370	1.00	0.00	C
ATOM	1536	O	ASP	A	100	9.842	-22.529	11.514	1.00	0.00	O
ATOM	1545	N	SER	A	101	11.065	-20.822	12.342	1.00	0.00	N
ATOM	1546	CA	SER	A	101	10.536	-20.932	13.701	1.00	0.00	C
ATOM	1547	C	SER	A	101	10.322	-19.554	14.295	1.00	0.00	C
ATOM	1548	O	SER	A	101	11.281	-18.848	14.596	1.00	0.00	O
ATOM	1556	N	SER	A	102	9.059	-19.188	14.467	1.00	0.00	N
ATOM	1557	CA	SER	A	102	8.714	-17.901	15.033	1.00	0.00	C
ATOM	1558	C	SER	A	102	7.775	-18.082	16.215	1.00	0.00	C
ATOM	1559	O	SER	A	102	6.628	-18.500	16.073	1.00	0.00	O
ATOM	1567	N	PHE	A	103	8.295	-17.750	17.376	1.00	0.00	N
ATOM	1568	CA	PHE	A	103	7.544	-17.848	18.626	1.00	0.00	C
ATOM	1569	C	PHE	A	103	7.214	-16.439	19.146	1.00	0.00	C
ATOM	1570	O	PHE	A	103	8.108	-15.752	19.640	1.00	0.00	O
ATOM	1587	N	SER	A	104	5.947	-16.010	19.039	1.00	0.00	N
ATOM	1588	CA	SER	A	104	5.564	-14.670	19.512	1.00	0.00	C
ATOM	1589	C	SER	A	104	4.647	-14.716	20.754	1.00	0.00	C
ATOM	1590	O	SER	A	104	3.996	-15.732	21.036	1.00	0.00	O
ATOM	1598	N	PRO	A	105	4.506	-13.569	21.465	1.00	0.00	N
ATOM	1599	CA	PRO	A	105	3.622	-13.370	22.691	1.00	0.00	C
ATOM	1600	C	PRO	A	105	2.191	-13.886	22.732	1.00	0.00	C
ATOM	1601	O	PRO	A	105	1.775	-14.208	23.861	1.00	0.00	O
ATOM	1612	N	ASN	A	106	1.512	-13.860	21.585	1.00	0.00	N
ATOM	1613	CA	ASN	A	106	0.115	-14.296	21.436	1.00	0.00	C
ATOM	1614	C	ASN	A	106	-0.056	-15.746	21.904	1.00	0.00	C
ATOM	1615	O	ASN	A	106	-0.104	-16.621	21.025	1.00	0.00	O
ATOM	1626	N	THR	A	107	-0.188	-15.958	23.245	1.00	0.00	N
ATOM	1627	CA	THR	A	107	-0.312	-17.259	23.911	1.00	0.00	C
ATOM	1628	C	THR	A	107	0.751	-18.236	23.420	1.00	0.00	C
ATOM	1629	O	THR	A	107	0.445	-19.390	23.122	1.00	0.00	O
ATOM	1640	N	GLY	A	108	1.996	-17.773	23.336	1.00	0.00	N
ATOM	1641	CA	GLY	A	108	3.082	-18.633	22.877	1.00	0.00	C
ATOM	1642	C	GLY	A	108	2.795	-19.157	21.474	1.00	0.00	C
ATOM	1643	O	GLY	A	108	2.913	-20.353	21.213	1.00	0.00	O
ATOM	1647	N	LYS	A	109	2.408	-18.252	20.581	1.00	0.00	N
ATOM	1648	CA	LYS	A	109	2.091	-18.636	19.205	1.00	0.00	C
ATOM	1649	C	LYS	A	109	3.336	-19.152	18.491	1.00	0.00	C
ATOM	1650	O	LYS	A	109	4.289	-18.405	18.265	1.00	0.00	O
ATOM	1669	N	LYS	A	110	3.315	-20.440	18.152	1.00	0.00	N
ATOM	1670	CA	LYS	A	110	4.442	-21.082	17.475	1.00	0.00	C
ATOM	1671	C	LYS	A	110	4.207	-21.161	15.970	1.00	0.00	C
ATOM	1672	O	LYS	A	110	3.224	-21.747	15.514	1.00	0.00	O
ATOM	1691	N	ASN	A	111	5.121	-20.569	15.206	1.00	0.00	N
ATOM	1692	CA	ASN	A	111	5.026	-20.573	13.752	1.00	0.00	C
ATOM	1693	C	ASN	A	111	6.157	-21.407	13.155	1.00	0.00	C
ATOM	1694	O	ASN	A	111	7.305	-20.961	13.112	1.00	0.00	O
ATOM	1705	N	ALA	A	112	5.824	-22.613	12.691	1.00	0.00	N
ATOM	1706	CA	ALA	A	112	6.809	-23.501	12.092	1.00	0.00	C
ATOM	1707	C	ALA	A	112	6.496	-23.709	10.619	1.00	0.00	C
ATOM	1708	O	ALA	A	112	5.501	-24.343	10.276	1.00	0.00	O
ATOM	1715	N	LYS	A	113	7.347	-23.159	9.751	1.00	0.00	N
ATOM	1716	CA	LYS	A	113	7.121	-23.280	8.301	1.00	0.00	C
ATOM	1717	C	LYS	A	113	8.238	-24.033	7.595	1.00	0.00	C
ATOM	1718	O	LYS	A	113	9.403	-23.846	7.902	1.00	0.00	O
ATOM	1737	N	ILE	A	114	7.864	-24.789	6.570	1.00	0.00	N
ATOM	1738	CA	ILE	A	114	8.842	-25.481	5.734	1.00	0.00	C
ATOM	1739	C	ILE	A	114	8.599	-25.078	4.285	1.00	0.00	C
ATOM	1740	O	ILE	A	114	7.656	-25.546	3.655	1.00	0.00	O
ATOM	1756	N	LYS	A	115	9.458	-24.199	3.772	1.00	0.00	N
ATOM	1757	CA	LYS	A	115	9.341	-23.721	2.408	1.00	0.00	C
ATOM	1758	C	LYS	A	115	10.291	-24.497	1.527	1.00	0.00	C
ATOM	1759	O	LYS	A	115	11.488	-24.488	1.765	1.00	0.00	O
ATOM	1778	N	THR	A	116	9.764	-25.106	0.490	1.00	0.00	N
ATOM	1779	CA	THR	A	116	10.603	-25.816	-0.456	1.00	0.00	C
ATOM	1780	C	THR	A	116	10.321	-25.250	-1.828	1.00	0.00	C
ATOM	1781	O	THR	A	116	9.258	-25.487	-2.381	1.00	0.00	O
ATOM	1792	N	GLY	A	117	11.271	-24.474	-2.347	1.00	0.00	N
ATOM	1793	CA	GLY	A	117	11.128	-23.840	-3.631	1.00	0.00	C
ATOM	1794	C	GLY	A	117	12.074	-24.439	-4.640	1.00	0.00	C
ATOM	1795	O	GLY	A	117	13.264	-24.122	-4.658	1.00	0.00	O
ATOM	1799	N	TYR	A	118	11.536	-25.286	-5.505	1.00	0.00	N

ATOM	1800	CA	TYR A	118	12.344	-25.886	-6.547	1.00	0.00	C
ATOM	1801	C	TYR A	118	12.232	-25.017	-7.778	1.00	0.00	C
ATOM	1802	O	TYR A	118	11.147	-24.549	-8.128	1.00	0.00	O
ATOM	1820	N	LYS A	119	13.373	-24.744	-8.376	1.00	0.00	N
ATOM	1821	CA	LYS A	119	13.440	-23.858	-9.511	1.00	0.00	C
ATOM	1822	C	LYS A	119	13.952	-24.594	-10.751	1.00	0.00	C
ATOM	1823	O	LYS A	119	14.703	-25.562	-10.645	1.00	0.00	O
ATOM	1842	N	ARG A	120	13.559	-24.119	-11.923	1.00	0.00	N
ATOM	1843	CA	ARG A	120	14.004	-24.728	-13.176	1.00	0.00	C
ATOM	1844	C	ARG A	120	13.558	-23.834	-14.315	1.00	0.00	C
ATOM	1845	O	ARG A	120	12.483	-24.030	-14.861	1.00	0.00	O
ATOM	1866	N	GLU A	121	14.378	-22.829	-14.637	1.00	0.00	N
ATOM	1867	CA	GLU A	121	14.072	-21.881	-15.701	1.00	0.00	C
ATOM	1868	C	GLU A	121	12.567	-21.582	-15.804	1.00	0.00	C
ATOM	1869	O	GLU A	121	11.749	-22.444	-16.115	1.00	0.00	O
ATOM	1881	N	HIS A	122	12.239	-20.318	-15.606	1.00	0.00	N
ATOM	1882	CA	HIS A	122	10.855	-19.839	-15.729	1.00	0.00	C
ATOM	1883	C	HIS A	122	9.864	-20.573	-14.819	1.00	0.00	C
ATOM	1884	O	HIS A	122	8.734	-20.095	-14.665	1.00	0.00	O
ATOM	1898	N	ILE A	123	10.248	-21.730	-14.248	1.00	0.00	N
ATOM	1899	CA	ILE A	123	9.327	-22.483	-13.390	1.00	0.00	C
ATOM	1900	C	ILE A	123	9.785	-22.537	-11.939	1.00	0.00	C
ATOM	1901	O	ILE A	123	10.774	-23.209	-11.631	1.00	0.00	O
ATOM	1917	N	ASN A	124	9.025	-21.895	-11.045	1.00	0.00	N
ATOM	1918	CA	ASN A	124	9.339	-21.964	-9.614	1.00	0.00	C
ATOM	1919	C	ASN A	124	8.143	-22.530	-8.835	1.00	0.00	C
ATOM	1920	O	ASN A	124	7.019	-22.037	-8.957	1.00	0.00	O
ATOM	1931	N	LEU A	125	8.399	-23.571	-8.038	1.00	0.00	N
ATOM	1932	CA	LEU A	125	7.346	-24.203	-7.240	1.00	0.00	C
ATOM	1933	C	LEU A	125	7.708	-24.169	-5.763	1.00	0.00	C
ATOM	1934	O	LEU A	125	8.608	-24.893	-5.368	1.00	0.00	O
ATOM	1950	N	GLY A	126	7.007	-23.383	-4.934	1.00	0.00	N
ATOM	1951	CA	GLY A	126	7.289	-23.329	-3.511	1.00	0.00	C
ATOM	1952	C	GLY A	126	6.191	-23.996	-2.707	1.00	0.00	C
ATOM	1953	O	GLY A	126	5.016	-23.802	-2.995	1.00	0.00	O
ATOM	1957	N	CYS A	127	6.608	-24.754	-1.681	1.00	0.00	N
ATOM	1958	CA	CYS A	127	5.685	-25.446	-0.781	1.00	0.00	C
ATOM	1959	C	CYS A	127	5.953	-25.063	0.685	1.00	0.00	C
ATOM	1960	O	CYS A	127	6.932	-25.500	1.281	1.00	0.00	O
ATOM	1968	N	ASP A	128	5.107	-24.197	1.223	1.00	0.00	N
ATOM	1969	CA	ASP A	128	5.247	-23.723	2.589	1.00	0.00	C
ATOM	1970	C	ASP A	128	4.271	-24.474	3.497	1.00	0.00	C
ATOM	1971	O	ASP A	128	3.062	-24.242	3.444	1.00	0.00	O
ATOM	1980	N	MET A	129	4.800	-25.364	4.338	1.00	0.00	N
ATOM	1981	CA	MET A	129	3.950	-26.122	5.254	1.00	0.00	C
ATOM	1982	C	MET A	129	4.038	-25.494	6.632	1.00	0.00	C
ATOM	1983	O	MET A	129	5.083	-25.537	7.266	1.00	0.00	O
ATOM	1997	N	ASP A	130	2.938	-24.906	7.082	1.00	0.00	N
ATOM	1998	CA	ASP A	130	2.906	-24.249	8.380	1.00	0.00	C
ATOM	1999	C	ASP A	130	2.332	-25.166	9.449	1.00	0.00	C
ATOM	2000	O	ASP A	130	1.119	-25.327	9.531	1.00	0.00	O
ATOM	2009	N	PHE A	131	3.201	-25.748	10.280	1.00	0.00	N
ATOM	2010	CA	PHE A	131	2.739	-26.634	11.345	1.00	0.00	C
ATOM	2011	C	PHE A	131	2.280	-25.830	12.554	1.00	0.00	C
ATOM	2012	O	PHE A	131	3.032	-25.641	13.514	1.00	0.00	O
ATOM	2029	N	ASP A	132	1.036	-25.372	12.501	1.00	0.00	N
ATOM	2030	CA	ASP A	132	0.456	-24.598	13.592	1.00	0.00	C
ATOM	2031	C	ASP A	132	-0.843	-25.262	14.027	1.00	0.00	C
ATOM	2032	O	ASP A	132	-1.337	-26.160	13.344	1.00	0.00	O
ATOM	2041	N	ILE A	133	-1.394	-24.855	15.159	1.00	0.00	N
ATOM	2042	CA	ILE A	133	-2.618	-25.484	15.617	1.00	0.00	C
ATOM	2043	C	ILE A	133	-3.725	-25.239	14.602	1.00	0.00	C
ATOM	2044	O	ILE A	133	-4.450	-26.170	14.250	1.00	0.00	O
ATOM	2060	N	ALA A	134	-3.804	-24.021	14.060	1.00	0.00	N
ATOM	2061	CA	ALA A	134	-4.782	-23.754	13.013	1.00	0.00	C
ATOM	2062	C	ALA A	134	-4.331	-24.522	11.771	1.00	0.00	C
ATOM	2063	O	ALA A	134	-5.139	-25.062	11.020	1.00	0.00	O
ATOM	2070	N	GLY A	135	-3.008	-24.614	11.614	1.00	0.00	N
ATOM	2071	CA	GLY A	135	-2.410	-25.384	10.528	1.00	0.00	C
ATOM	2072	C	GLY A	135	-2.513	-26.854	10.925	1.00	0.00	C
ATOM	2073	O	GLY A	135	-3.157	-27.163	11.931	1.00	0.00	O
ATOM	2077	N	PRO A	136	-1.908	-27.782	10.228	1.00	0.00	N
ATOM	2078	CA	PRO A	136	-1.083	-27.576	8.981	1.00	0.00	C
ATOM	2079	C	PRO A	136	-1.778	-26.814	7.856	1.00	0.00	C
ATOM	2080	O	PRO A	136	-2.793	-27.281	7.332	1.00	0.00	O
ATOM	2091	N	SER A	137	-1.177	-25.708	7.388	1.00	0.00	N
ATOM	2092	CA	SER A	137	-1.704	-25.006	6.231	1.00	0.00	C
ATOM	2093	C	SER A	137	-0.676	-25.161	5.112	1.00	0.00	C
ATOM	2094	O	SER A	137	0.531	-25.092	5.372	1.00	0.00	O
ATOM	2102	N	ILE A	138	-1.122	-25.384	3.888	1.00	0.00	N
ATOM	2103	CA	ILE A	138	-0.187	-25.547	2.775	1.00	0.00	C
ATOM	2104	C	ILE A	138	-0.232	-24.337	1.848	1.00	0.00	C
ATOM	2105	O	ILE A	138	-1.151	-24.202	1.054	1.00	0.00	O
ATOM	2121	N	ARG A	139	0.774	-23.467	1.973	1.00	0.00	N
ATOM	2122	CA	ARG A	139	0.853	-22.259	1.145	1.00	0.00	C
ATOM	2123	C	ARG A	139	1.988	-22.378	0.129	1.00	0.00	C
ATOM	2124	O	ARG A	139	3.160	-22.298	0.479	1.00	0.00	O
ATOM	2145	N	GLY A	140	1.646	-22.618	-1.132	1.00	0.00	N
ATOM	2146	CA	GLY A	140	2.689	-22.771	-2.168	1.00	0.00	C
ATOM	2147	C	GLY A	140	2.500	-21.741	-3.272	1.00	0.00	C
ATOM	2148	O	GLY A	140	1.424	-21.199	-3.356	1.00	0.00	O
ATOM	2152	N	ALA A	141	3.507	-21.503	-4.147	1.00	0.00	N
ATOM	2153	CA	ALA A	141	3.352	-20.572	-5.236	1.00	0.00	C
ATOM	2154	C	ALA A	141	3.916	-21.219	-6.487	1.00	0.00	C

## Appendix D PDB files of the models for the closed state of VDAC

ATOM	2155	O	ALA A 141	5.066	-21.661	-6.495	1.00	0.00	O
ATOM	2162	N	LEU A 142	3.120	-21.278	-7.525	1.00	0.00	N
ATOM	2163	CA	LEU A 142	3.557	-21.889	-8.781	1.00	0.00	C
ATOM	2164	C	LEU A 142	3.716	-20.806	-9.816	1.00	0.00	C
ATOM	2165	O	LEU A 142	2.713	-20.252	-10.248	1.00	0.00	O
ATOM	2181	N	VAL A 143	4.950	-20.501	-10.218	1.00	0.00	N
ATOM	2182	CA	VAL A 143	5.144	-19.454	-11.212	1.00	0.00	C
ATOM	2183	C	VAL A 143	5.703	-20.019	-12.500	1.00	0.00	C
ATOM	2184	O	VAL A 143	6.575	-20.896	-12.512	1.00	0.00	O
ATOM	2197	N	LEU A 144	5.157	-19.483	-13.578	1.00	0.00	N
ATOM	2198	CA	LEU A 144	5.535	-19.870	-14.927	1.00	0.00	C
ATOM	2199	C	LEU A 144	6.040	-18.642	-15.644	1.00	0.00	C
ATOM	2200	O	LEU A 144	5.447	-17.571	-15.513	1.00	0.00	O
ATOM	2216	N	GLY A 145	7.125	-18.781	-16.403	1.00	0.00	N
ATOM	2217	CA	GLY A 145	7.683	-17.640	-17.121	1.00	0.00	C
ATOM	2218	C	GLY A 145	7.409	-17.724	-18.621	1.00	0.00	C
ATOM	2219	O	GLY A 145	7.359	-18.818	-19.189	1.00	0.00	O
ATOM	2223	N	TYR A 146	7.275	-16.568	-19.276	1.00	0.00	N
ATOM	2224	CA	TYR A 146	7.058	-16.552	-20.723	1.00	0.00	C
ATOM	2225	C	TYR A 146	7.736	-15.325	-21.311	1.00	0.00	C
ATOM	2226	O	TYR A 146	8.612	-14.773	-20.660	1.00	0.00	O
ATOM	2244	N	GLU A 147	7.312	-14.923	-22.532	1.00	0.00	N
ATOM	2245	CA	GLU A 147	7.857	-13.753	-23.261	1.00	0.00	C
ATOM	2246	C	GLU A 147	7.993	-12.497	-22.384	1.00	0.00	C
ATOM	2247	O	GLU A 147	7.494	-11.434	-22.745	1.00	0.00	O
ATOM	2259	N	GLY A 148	8.649	-12.620	-21.247	1.00	0.00	N
ATOM	2260	CA	GLY A 148	8.806	-11.500	-20.336	1.00	0.00	C
ATOM	2261	C	GLY A 148	7.625	-11.425	-19.362	1.00	0.00	C
ATOM	2262	O	GLY A 148	7.202	-10.339	-18.981	1.00	0.00	O
ATOM	2266	N	TRP A 149	7.093	-12.583	-18.968	1.00	0.00	N
ATOM	2267	CA	TRP A 149	5.959	-12.596	-18.032	1.00	0.00	C
ATOM	2268	C	TRP A 149	6.178	-13.594	-16.915	1.00	0.00	C
ATOM	2269	O	TRP A 149	6.669	-14.685	-17.139	1.00	0.00	O
ATOM	2290	N	LEU A 150	5.749	-13.195	-15.728	1.00	0.00	N
ATOM	2291	CA	LEU A 150	5.799	-14.022	-14.525	1.00	0.00	C
ATOM	2292	C	LEU A 150	4.373	-14.166	-13.964	1.00	0.00	C
ATOM	2293	O	LEU A 150	3.834	-13.197	-13.433	1.00	0.00	O
ATOM	2309	N	ALA A 151	3.782	-15.352	-14.022	1.00	0.00	N
ATOM	2310	CA	ALA A 151	2.439	-15.549	-13.446	1.00	0.00	C
ATOM	2311	C	ALA A 151	2.571	-16.485	-12.258	1.00	0.00	C
ATOM	2312	O	ALA A 151	3.356	-17.410	-12.344	1.00	0.00	O
ATOM	2319	N	GLY A 152	1.852	-16.252	-11.143	1.00	0.00	N
ATOM	2320	CA	GLY A 152	1.993	-17.125	-9.978	1.00	0.00	C
ATOM	2321	C	GLY A 152	0.646	-17.561	-9.418	1.00	0.00	C
ATOM	2322	O	GLY A 152	-0.349	-16.833	-9.520	1.00	0.00	O
ATOM	2326	N	TYR A 153	0.625	-18.746	-8.804	1.00	0.00	N
ATOM	2327	CA	TYR A 153	-0.600	-19.276	-8.216	1.00	0.00	C
ATOM	2328	C	TYR A 153	-0.293	-19.675	-6.800	1.00	0.00	C
ATOM	2329	O	TYR A 153	0.464	-20.624	-6.593	1.00	0.00	O
ATOM	2347	N	GLN A 154	-0.859	-18.972	-5.815	1.00	0.00	N
ATOM	2348	CA	GLN A 154	-0.588	-19.292	-4.440	1.00	0.00	C
ATOM	2349	C	GLN A 154	-1.771	-20.024	-3.832	1.00	0.00	C
ATOM	2350	O	GLN A 154	-2.824	-19.430	-3.693	1.00	0.00	O
ATOM	2364	N	MET A 155	-1.588	-21.298	-3.478	1.00	0.00	N
ATOM	2365	CA	MET A 155	-2.679	-22.083	-2.887	1.00	0.00	C
ATOM	2366	C	MET A 155	-2.476	-22.312	-1.389	1.00	0.00	C
ATOM	2367	O	MET A 155	-1.535	-22.984	-0.970	1.00	0.00	O
ATOM	2381	N	ASN A 156	-3.407	-21.783	-0.603	1.00	0.00	N
ATOM	2382	CA	ASN A 156	-3.384	-21.962	0.841	1.00	0.00	C
ATOM	2383	C	ASN A 156	-4.433	-23.004	1.212	1.00	0.00	C
ATOM	2384	O	ASN A 156	-5.636	-22.786	1.041	1.00	0.00	O
ATOM	2395	N	PHE A 157	-3.964	-24.130	1.713	1.00	0.00	N
ATOM	2396	CA	PHE A 157	-4.859	-25.216	2.099	1.00	0.00	C
ATOM	2397	C	PHE A 157	-4.926	-25.366	3.609	1.00	0.00	C
ATOM	2398	O	PHE A 157	-4.081	-26.013	4.215	1.00	0.00	O
ATOM	2415	N	GLU A 158	-5.931	-24.754	4.210	1.00	0.00	N
ATOM	2416	CA	GLU A 158	-6.103	-24.846	5.660	1.00	0.00	C
ATOM	2417	C	GLU A 158	-6.634	-26.241	5.985	1.00	0.00	C
ATOM	2418	O	GLU A 158	-7.815	-26.521	5.821	1.00	0.00	O
ATOM	2430	N	THR A 159	-5.738	-27.141	6.386	1.00	0.00	N
ATOM	2431	CA	THR A 159	-6.130	-28.533	6.651	1.00	0.00	C
ATOM	2432	C	THR A 159	-7.046	-28.694	7.871	1.00	0.00	C
ATOM	2433	O	THR A 159	-7.897	-29.583	7.881	1.00	0.00	O
ATOM	2444	N	ALA A 160	-6.882	-27.864	8.899	1.00	0.00	N
ATOM	2445	CA	ALA A 160	-7.730	-28.005	10.085	1.00	0.00	C
ATOM	2446	C	ALA A 160	-9.191	-27.837	9.693	1.00	0.00	C
ATOM	2447	O	ALA A 160	-10.060	-28.577	10.152	1.00	0.00	O
ATOM	2454	N	LYS A 161	-9.442	-26.882	8.808	1.00	0.00	N
ATOM	2455	CA	LYS A 161	-10.793	-26.641	8.315	1.00	0.00	C
ATOM	2456	C	LYS A 161	-10.938	-27.323	6.959	1.00	0.00	C
ATOM	2457	O	LYS A 161	-12.037	-27.464	6.427	1.00	0.00	O
ATOM	2476	N	SER A 162	-9.798	-27.710	6.404	1.00	0.00	N
ATOM	2477	CA	SER A 162	-9.751	-28.344	5.089	1.00	0.00	C
ATOM	2478	C	SER A 162	-10.431	-27.436	4.076	1.00	0.00	C
ATOM	2479	O	SER A 162	-11.321	-27.858	3.339	1.00	0.00	O
ATOM	2487	N	ARG A 163	-10.004	-26.171	4.063	1.00	0.00	N
ATOM	2488	CA	ARG A 163	-10.569	-25.181	3.156	1.00	0.00	C
ATOM	2489	C	ARG A 163	-9.485	-24.455	2.369	1.00	0.00	C
ATOM	2490	O	ARG A 163	-8.465	-24.043	2.926	1.00	0.00	O
ATOM	2511	N	VAL A 164	-9.759	-24.231	1.090	1.00	0.00	N
ATOM	2512	CA	VAL A 164	-8.868	-23.477	0.250	1.00	0.00	C
ATOM	2513	C	VAL A 164	-9.521	-22.123	0.070	1.00	0.00	C
ATOM	2514	O	VAL A 164	-10.478	-21.978	-0.693	1.00	0.00	O
ATOM	2527	N	THR A 165	-9.027	-21.158	0.801	1.00	0.00	N

ATOM	2528	CA	THR A	165	-9.591	-19.807	0.754	1.00	0.00	C
ATOM	2529	C	THR A	165	-8.547	-18.804	0.315	1.00	0.00	C
ATOM	2530	O	THR A	165	-8.709	-18.067	-0.659	1.00	0.00	O
ATOM	2541	N	GLN A	166	-7.484	-18.792	1.073	1.00	0.00	N
ATOM	2542	CA	GLN A	166	-6.383	-17.885	0.809	1.00	0.00	C
ATOM	2543	C	GLN A	166	-5.644	-18.285	-0.467	1.00	0.00	C
ATOM	2544	O	GLN A	166	-4.927	-19.288	-0.503	1.00	0.00	O
ATOM	2558	N	SER A	167	-5.813	-17.477	-1.508	1.00	0.00	N
ATOM	2559	CA	SER A	167	-5.148	-17.727	-2.782	1.00	0.00	C
ATOM	2560	C	SER A	167	-4.660	-16.407	-3.374	1.00	0.00	C
ATOM	2561	O	SER A	167	-5.341	-15.385	-3.264	1.00	0.00	O
ATOM	2569	N	ASN A	168	-3.463	-16.424	-3.967	1.00	0.00	N
ATOM	2570	CA	ASN A	168	-2.891	-15.211	-4.535	1.00	0.00	C
ATOM	2571	C	ASN A	168	-2.434	-15.414	-5.985	1.00	0.00	C
ATOM	2572	O	ASN A	168	-1.562	-16.239	-6.262	1.00	0.00	O
ATOM	2583	N	PHE A	169	-3.006	-14.627	-6.900	1.00	0.00	N
ATOM	2584	CA	PHE A	169	-2.630	-14.697	-8.318	1.00	0.00	C
ATOM	2585	C	PHE A	169	-1.537	-13.671	-8.598	1.00	0.00	C
ATOM	2586	O	PHE A	169	-1.794	-12.473	-8.530	1.00	0.00	O
ATOM	2603	N	ALA A	170	-0.342	-14.121	-8.944	1.00	0.00	N
ATOM	2604	CA	ALA A	170	0.730	-13.186	-9.257	1.00	0.00	C
ATOM	2605	C	ALA A	170	0.752	-12.912	-10.747	1.00	0.00	C
ATOM	2606	O	ALA A	170	0.745	-13.837	-11.524	1.00	0.00	O
ATOM	2613	N	VAL A	171	0.814	-11.647	-11.137	1.00	0.00	N
ATOM	2614	CA	VAL A	171	0.887	-11.300	-12.555	1.00	0.00	C
ATOM	2615	C	VAL A	171	2.011	-10.287	-12.754	1.00	0.00	C
ATOM	2616	O	VAL A	171	2.053	-9.258	-12.077	1.00	0.00	O
ATOM	2629	N	GLY A	172	2.931	-10.602	-13.664	1.00	0.00	N
ATOM	2630	CA	GLY A	172	4.038	-9.719	-13.909	1.00	0.00	C
ATOM	2631	C	GLY A	172	4.339	-9.548	-15.383	1.00	0.00	C
ATOM	2632	O	GLY A	172	4.208	-10.475	-16.185	1.00	0.00	O
ATOM	2636	N	TYR A	173	4.781	-8.345	-15.703	1.00	0.00	N
ATOM	2637	CA	TYR A	173	5.154	-7.987	-17.060	1.00	0.00	C
ATOM	2638	C	TYR A	173	6.545	-7.382	-17.031	1.00	0.00	C
ATOM	2639	O	TYR A	173	6.824	-6.501	-16.219	1.00	0.00	O
ATOM	2657	N	LYS A	174	7.431	-7.887	-17.864	1.00	0.00	N
ATOM	2658	CA	LYS A	174	8.812	-7.414	-17.869	1.00	0.00	C
ATOM	2659	C	LYS A	174	9.085	-6.264	-18.845	1.00	0.00	C
ATOM	2660	O	LYS A	174	8.547	-6.191	-19.947	1.00	0.00	O
ATOM	2679	N	THR A	175	9.993	-5.415	-18.409	1.00	0.00	N
ATOM	2680	CA	THR A	175	10.467	-4.272	-19.177	1.00	0.00	C
ATOM	2681	C	THR A	175	11.982	-4.173	-18.967	1.00	0.00	C
ATOM	2682	O	THR A	175	12.484	-4.630	-17.940	1.00	0.00	O
ATOM	2693	N	ASP A	176	12.715	-3.632	-19.938	1.00	0.00	N
ATOM	2694	CA	ASP A	176	14.178	-3.558	-19.819	1.00	0.00	C
ATOM	2695	C	ASP A	176	14.615	-3.233	-18.393	1.00	0.00	C
ATOM	2696	O	ASP A	176	14.234	-2.197	-17.846	1.00	0.00	O
ATOM	2705	N	GLU A	177	15.411	-4.136	-17.798	1.00	0.00	N
ATOM	2706	CA	GLU A	177	15.912	-3.957	-16.425	1.00	0.00	C
ATOM	2707	C	GLU A	177	14.785	-3.837	-15.396	1.00	0.00	C
ATOM	2708	O	GLU A	177	15.024	-3.943	-14.186	1.00	0.00	O
ATOM	2720	N	PHE A	178	13.586	-3.510	-15.867	1.00	0.00	N
ATOM	2721	CA	PHE A	178	12.459	-3.277	-14.972	1.00	0.00	C
ATOM	2722	C	PHE A	178	11.470	-4.457	-14.956	1.00	0.00	C
ATOM	2723	O	PHE A	178	11.100	-4.966	-16.007	1.00	0.00	O
ATOM	2740	N	GLN A	179	10.999	-4.848	-13.753	1.00	0.00	N
ATOM	2741	CA	GLN A	179	9.995	-5.923	-13.618	1.00	0.00	C
ATOM	2742	C	GLN A	179	8.699	-5.378	-13.017	1.00	0.00	C
ATOM	2743	O	GLN A	179	8.746	-4.613	-12.048	1.00	0.00	O
ATOM	2757	N	LEU A	180	7.556	-5.852	-13.528	1.00	0.00	N
ATOM	2758	CA	LEU A	180	6.255	-5.457	-12.979	1.00	0.00	C
ATOM	2759	C	LEU A	180	5.676	-6.641	-12.221	1.00	0.00	C
ATOM	2760	O	LEU A	180	5.324	-7.635	-12.824	1.00	0.00	O
ATOM	2776	N	HIS A	181	5.619	-6.528	-10.899	1.00	0.00	N
ATOM	2777	CA	HIS A	181	5.096	-7.604	-10.054	1.00	0.00	C
ATOM	2778	C	HIS A	181	3.863	-7.155	-9.250	1.00	0.00	C
ATOM	2779	O	HIS A	181	3.959	-6.268	-8.399	1.00	0.00	O
ATOM	2793	N	THR A	182	2.723	-7.802	-9.494	1.00	0.00	N
ATOM	2794	CA	THR A	182	1.478	-7.501	-8.760	1.00	0.00	C
ATOM	2795	C	THR A	182	0.825	-8.803	-8.325	1.00	0.00	C
ATOM	2796	O	THR A	182	1.031	-9.822	-8.995	1.00	0.00	O
ATOM	2807	N	ASN A	183	0.016	-8.818	-7.239	1.00	0.00	N
ATOM	2808	CA	ASN A	183	-0.631	-10.049	-6.846	1.00	0.00	C
ATOM	2809	C	ASN A	183	-2.057	-9.757	-6.404	1.00	0.00	C
ATOM	2810	O	ASN A	183	-2.301	-8.808	-5.660	1.00	0.00	O
ATOM	2821	N	VAL A	184	-2.990	-10.589	-6.837	1.00	0.00	N
ATOM	2822	CA	VAL A	184	-4.375	-10.404	-6.444	1.00	0.00	C
ATOM	2823	C	VAL A	184	-4.580	-11.212	-5.199	1.00	0.00	C
ATOM	2824	O	VAL A	184	-4.419	-12.429	-5.224	1.00	0.00	O
ATOM	2837	N	ASN A	185	-4.915	-10.550	-4.105	1.00	0.00	N
ATOM	2838	CA	ASN A	185	-5.104	-11.245	-2.868	1.00	0.00	C
ATOM	2839	C	ASN A	185	-6.568	-11.517	-2.644	1.00	0.00	C
ATOM	2840	O	ASN A	185	-7.344	-10.581	-2.448	1.00	0.00	O
ATOM	2851	N	ASP A	186	-6.901	-12.800	-2.711	1.00	0.00	N
ATOM	2852	CA	ASP A	186	-8.272	-13.283	-2.508	1.00	0.00	C
ATOM	2853	C	ASP A	186	-9.289	-12.582	-3.405	1.00	0.00	C
ATOM	2854	O	ASP A	186	-10.477	-12.546	-3.077	1.00	0.00	O
ATOM	2863	N	GLY A	187	-8.844	-12.040	-4.533	1.00	0.00	N
ATOM	2864	CA	GLY A	187	-9.771	-11.364	-5.433	1.00	0.00	C
ATOM	2865	C	GLY A	187	-10.274	-10.074	-4.802	1.00	0.00	C
ATOM	2866	O	GLY A	187	-11.314	-9.550	-5.200	1.00	0.00	O
ATOM	2870	N	THR A	188	-9.561	-9.604	-3.777	1.00	0.00	N
ATOM	2871	CA	THR A	188	-9.979	-8.412	-3.051	1.00	0.00	C
ATOM	2872	C	THR A	188	-8.919	-7.296	-3.046	1.00	0.00	C



## Appendix D PDB files of the models for the closed state of VDAC

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ATOM	2873	O	THR A 188	-9.142	-6.232	-3.624	1.00	0.00	O
ATOM	2884	N	GLU A 189	-7.788	-7.506	-2.373	1.00	0.00	N
ATOM	2885	CA	GLU A 189	-6.759	-6.442	-2.316	1.00	0.00	C
ATOM	2886	C	GLU A 189	-5.514	-6.776	-3.140	1.00	0.00	C
ATOM	2887	O	GLU A 189	-4.889	-7.813	-2.929	1.00	0.00	O
ATOM	2899	N	PHE A 190	-5.139	-5.884	-4.073	1.00	0.00	N
ATOM	2900	CA	PHE A 190	-3.959	-6.095	-4.903	1.00	0.00	C
ATOM	2901	C	PHE A 190	-2.726	-5.488	-4.239	1.00	0.00	C
ATOM	2902	O	PHE A 190	-2.823	-4.494	-3.516	1.00	0.00	O
ATOM	2919	N	GLY A 191	-1.566	-6.075	-4.509	1.00	0.00	N
ATOM	2920	CA	GLY A 191	-0.319	-5.565	-3.944	1.00	0.00	C
ATOM	2921	C	GLY A 191	0.896	-6.273	-4.518	1.00	0.00	C
ATOM	2922	O	GLY A 191	0.846	-7.472	-4.806	1.00	0.00	O
ATOM	2926	N	GLY A 192	2.008	-5.540	-4.683	1.00	0.00	N
ATOM	2927	CA	GLY A 192	3.216	-6.124	-5.266	1.00	0.00	C
ATOM	2928	C	GLY A 192	4.395	-5.149	-5.219	1.00	0.00	C
ATOM	2929	O	GLY A 192	4.372	-4.244	-4.364	1.00	0.00	O
ATOM	2933	N	SER A 193	5.361	-5.361	-6.097	1.00	0.00	N
ATOM	2934	CA	SER A 193	6.540	-4.491	-6.274	1.00	0.00	C
ATOM	2935	C	SER A 193	6.917	-4.322	-7.736	1.00	0.00	C
ATOM	2936	O	SER A 193	6.329	-5.092	-8.529	1.00	0.00	O
ATOM	2944	N	ILE A 194	7.787	-3.338	-8.017	1.00	0.00	N
ATOM	2945	CA	ILE A 194	8.346	-3.094	-9.329	1.00	0.00	C
ATOM	2946	C	ILE A 194	9.858	-3.072	-9.167	1.00	0.00	C
ATOM	2947	O	ILE A 194	10.388	-2.170	-8.529	1.00	0.00	O
ATOM	2963	N	TYR A 195	10.562	-4.092	-9.644	1.00	0.00	N
ATOM	2964	CA	TYR A 195	12.038	-4.159	-9.565	1.00	0.00	C
ATOM	2965	C	TYR A 195	12.611	-3.216	-10.598	1.00	0.00	C
ATOM	2966	O	TYR A 195	12.321	-3.387	-11.773	1.00	0.00	O
ATOM	2984	N	GLN A 196	13.368	-2.209	-10.289	1.00	0.00	N
ATOM	2985	CA	GLN A 196	13.966	-1.329	-11.279	1.00	0.00	C
ATOM	2986	C	GLN A 196	15.481	-1.223	-11.138	1.00	0.00	C
ATOM	2987	O	GLN A 196	15.970	-0.445	-10.317	1.00	0.00	O
ATOM	3001	N	LYS A 197	16.248	-1.997	-11.926	1.00	0.00	N
ATOM	3002	CA	LYS A 197	17.711	-1.858	-11.931	1.00	0.00	C
ATOM	3003	C	LYS A 197	17.931	-0.572	-12.727	1.00	0.00	C
ATOM	3004	O	LYS A 197	17.925	-0.585	-13.955	1.00	0.00	O
ATOM	3023	N	VAL A 198	18.142	0.596	-12.216	1.00	0.00	N
ATOM	3024	CA	VAL A 198	18.332	1.852	-12.923	1.00	0.00	C
ATOM	3025	C	VAL A 198	19.721	1.974	-13.518	1.00	0.00	C
ATOM	3026	O	VAL A 198	19.888	2.434	-14.648	1.00	0.00	O
ATOM	3039	N	ASN A 199	20.709	1.554	-12.760	1.00	0.00	N
ATOM	3040	CA	ASN A 199	22.090	1.604	-13.222	1.00	0.00	C
ATOM	3041	C	ASN A 199	22.961	0.806	-12.263	1.00	0.00	C
ATOM	3042	O	ASN A 199	22.443	0.082	-11.417	1.00	0.00	O
ATOM	3053	N	LYS A 200	24.270	0.915	-12.404	1.00	0.00	N
ATOM	3054	CA	LYS A 200	25.175	0.161	-11.541	1.00	0.00	C
ATOM	3055	C	LYS A 200	25.020	0.555	-10.069	1.00	0.00	C
ATOM	3056	O	LYS A 200	25.052	-0.303	-9.190	1.00	0.00	O
ATOM	3075	N	LYS A 201	24.889	1.851	-9.798	1.00	0.00	N
ATOM	3076	CA	LYS A 201	24.779	2.316	-8.413	1.00	0.00	C
ATOM	3077	C	LYS A 201	23.336	2.566	-7.949	1.00	0.00	C
ATOM	3078	O	LYS A 201	23.133	2.906	-6.784	1.00	0.00	O
ATOM	3097	N	LEU A 202	22.361	2.638	-8.841	1.00	0.00	N
ATOM	3098	CA	LEU A 202	20.919	2.970	-8.697	1.00	0.00	C
ATOM	3099	C	LEU A 202	20.032	1.783	-9.083	1.00	0.00	C
ATOM	3100	O	LEU A 202	19.504	1.643	-10.211	1.00	0.00	O
ATOM	3116	N	GLU A 203	19.895	0.942	-8.075	1.00	0.00	N
ATOM	3117	CA	GLU A 203	18.837	-0.052	-7.986	1.00	0.00	C
ATOM	3118	C	GLU A 203	17.679	0.615	-7.240	1.00	0.00	C
ATOM	3119	O	GLU A 203	18.052	1.199	-6.182	1.00	0.00	O
ATOM	3131	N	THR A 204	16.446	0.550	-7.649	1.00	0.00	N
ATOM	3132	CA	THR A 204	15.269	1.221	-7.036	1.00	0.00	C
ATOM	3133	C	THR A 204	14.128	0.207	-6.929	1.00	0.00	C
ATOM	3134	O	THR A 204	14.407	-0.892	-7.497	1.00	0.00	O
ATOM	3145	N	ALA A 205	12.999	0.457	-6.241	1.00	0.00	N
ATOM	3146	CA	ALA A 205	11.943	-0.551	-6.150	1.00	0.00	C
ATOM	3147	C	ALA A 205	10.552	0.021	-5.835	1.00	0.00	C
ATOM	3148	O	ALA A 205	10.247	0.248	-4.661	1.00	0.00	O
ATOM	3155	N	VAL A 206	9.650	0.162	-6.828	1.00	0.00	N
ATOM	3156	CA	VAL A 206	8.400	0.929	-6.631	1.00	0.00	C
ATOM	3157	C	VAL A 206	7.383	-0.077	-6.034	1.00	0.00	C
ATOM	3158	O	VAL A 206	7.233	-1.106	-6.746	1.00	0.00	O
ATOM	3171	N	ASN A 207	6.744	0.113	-4.908	1.00	0.00	N
ATOM	3172	CA	ASN A 207	5.632	-0.739	-4.418	1.00	0.00	C
ATOM	3173	C	ASN A 207	4.321	-0.198	-4.963	1.00	0.00	C
ATOM	3174	O	ASN A 207	4.253	0.980	-4.876	1.00	0.00	O
ATOM	3185	N	LEU A 208	3.413	-1.026	-5.468	1.00	0.00	N
ATOM	3186	CA	LEU A 208	2.007	-0.675	-5.829	1.00	0.00	C
ATOM	3187	C	LEU A 208	1.042	-1.405	-4.890	1.00	0.00	C
ATOM	3188	O	LEU A 208	1.396	-2.543	-4.596	1.00	0.00	O
ATOM	3204	N	ALA A 209	-0.044	-0.802	-4.422	1.00	0.00	N
ATOM	3205	CA	ALA A 209	-1.072	-1.509	-3.641	1.00	0.00	C
ATOM	3206	C	ALA A 209	-2.434	-0.833	-3.756	1.00	0.00	C
ATOM	3207	O	ALA A 209	-2.432	0.350	-3.389	1.00	0.00	O
ATOM	3214	N	TRP A 210	-3.483	-1.529	-4.202	1.00	0.00	N
ATOM	3215	CA	TRP A 210	-4.814	-0.936	-4.341	1.00	0.00	C
ATOM	3216	C	TRP A 210	-5.828	-1.858	-3.675	1.00	0.00	C
ATOM	3217	O	TRP A 210	-5.715	-3.082	-3.744	1.00	0.00	O
ATOM	3238	N	THR A 211	-6.906	-1.366	-3.850	1.00	0.00	N
ATOM	3239	CA	THR A 211	-8.006	-2.167	-3.318	1.00	0.00	C
ATOM	3240	C	THR A 211	-9.174	-2.227	-4.277	1.00	0.00	C
ATOM	3241	O	THR A 211	-9.792	-1.203	-4.562	1.00	0.00	O
ATOM	3252	N	ALA A 212	-9.477	-3.429	-4.787	1.00	0.00	N

ATOM	3253	CA	ALA A	212	-10.588	-3.552	-5.726	1.00	0.00	C
ATOM	3254	C	ALA A	212	-11.768	-2.717	-5.271	1.00	0.00	C
ATOM	3255	O	ALA A	212	-12.536	-3.111	-4.390	1.00	0.00	O
ATOM	3262	N	GLY A	213	-11.884	-1.560	-5.900	1.00	0.00	N
ATOM	3263	CA	GLY A	213	-12.937	-0.612	-5.617	1.00	0.00	C
ATOM	3264	C	GLY A	213	-12.674	0.663	-6.412	1.00	0.00	C
ATOM	3265	O	GLY A	213	-11.939	0.645	-7.399	1.00	0.00	O
ATOM	3269	N	ASN A	214	-13.247	1.769	-5.982	1.00	0.00	N
ATOM	3270	CA	ASN A	214	-13.027	3.023	-6.681	1.00	0.00	C
ATOM	3271	C	ASN A	214	-11.962	3.828	-5.941	1.00	0.00	C
ATOM	3272	O	ASN A	214	-11.758	5.005	-6.217	1.00	0.00	O
ATOM	3283	N	SER A	215	-11.309	3.181	-4.970	1.00	0.00	N
ATOM	3284	CA	SER A	215	-10.293	3.847	-4.155	1.00	0.00	C
ATOM	3285	C	SER A	215	-8.939	3.989	-4.875	1.00	0.00	C
ATOM	3286	O	SER A	215	-8.708	3.375	-5.915	1.00	0.00	O
ATOM	3294	N	ASN A	216	-8.095	4.959	-4.372	1.00	0.00	N
ATOM	3295	CA	ASN A	216	-6.775	5.140	-4.995	1.00	0.00	C
ATOM	3296	C	ASN A	216	-5.776	4.040	-4.647	1.00	0.00	C
ATOM	3297	O	ASN A	216	-5.982	3.261	-3.715	1.00	0.00	O
ATOM	3308	N	THR A	217	-4.685	4.022	-5.429	1.00	0.00	N
ATOM	3309	CA	THR A	217	-3.516	3.186	-5.175	1.00	0.00	C
ATOM	3310	C	THR A	217	-2.557	3.937	-4.237	1.00	0.00	C
ATOM	3311	O	THR A	217	-2.000	4.960	-4.639	1.00	0.00	O
ATOM	3322	N	ARG A	218	-2.343	3.443	-3.023	1.00	0.00	N
ATOM	3323	CA	ARG A	218	-1.154	3.820	-2.258	1.00	0.00	C
ATOM	3324	C	ARG A	218	0.032	3.193	-2.975	1.00	0.00	C
ATOM	3325	O	ARG A	218	-0.115	1.981	-3.201	1.00	0.00	O
ATOM	3346	N	PHE A	219	1.083	3.929	-3.308	1.00	0.00	N
ATOM	3347	CA	PHE A	219	2.267	3.394	-4.001	1.00	0.00	C
ATOM	3348	C	PHE A	219	3.529	4.070	-3.505	1.00	0.00	C
ATOM	3349	O	PHE A	219	3.490	5.290	-3.403	1.00	0.00	O
ATOM	3366	N	GLY A	220	4.552	3.300	-3.215	1.00	0.00	N
ATOM	3367	CA	GLY A	220	5.882	3.764	-2.826	1.00	0.00	C
ATOM	3368	C	GLY A	220	6.778	3.821	-4.027	1.00	0.00	C
ATOM	3369	O	GLY A	220	6.937	2.728	-4.442	1.00	0.00	O
ATOM	3373	N	ILE A	221	7.273	4.983	-4.438	1.00	0.00	N
ATOM	3374	CA	ILE A	221	8.528	5.071	-5.201	1.00	0.00	C
ATOM	3375	C	ILE A	221	9.551	5.065	-4.128	1.00	0.00	C
ATOM	3376	O	ILE A	221	9.319	5.771	-3.121	1.00	0.00	O
ATOM	3392	N	ALA A	222	10.599	4.323	-4.343	1.00	0.00	N
ATOM	3393	CA	ALA A	222	11.601	4.170	-3.336	1.00	0.00	C
ATOM	3394	C	ALA A	222	12.947	4.084	-4.029	1.00	0.00	C
ATOM	3395	O	ALA A	222	12.823	3.592	-5.184	1.00	0.00	O
ATOM	3402	N	ALA A	223	14.079	4.504	-3.545	1.00	0.00	N
ATOM	3403	CA	ALA A	223	15.376	4.424	-4.252	1.00	0.00	C
ATOM	3404	C	ALA A	223	16.577	4.324	-3.306	1.00	0.00	C
ATOM	3405	O	ALA A	223	16.590	5.142	-2.404	1.00	0.00	O
ATOM	3412	N	LYS A	224	17.522	3.436	-3.564	1.00	0.00	N
ATOM	3413	CA	LYS A	224	18.758	3.186	-2.804	1.00	0.00	C
ATOM	3414	C	LYS A	224	19.881	3.585	-3.771	1.00	0.00	C
ATOM	3415	O	LYS A	224	20.186	2.760	-4.660	1.00	0.00	O
ATOM	3434	N	TYR A	225	20.451	4.760	-3.583	1.00	0.00	N
ATOM	3435	CA	TYR A	225	21.607	5.268	-4.336	1.00	0.00	C
ATOM	3436	C	TYR A	225	22.910	5.030	-3.593	1.00	0.00	C
ATOM	3437	O	TYR A	225	23.168	5.809	-2.678	1.00	0.00	O
ATOM	3455	N	GLN A	226	23.682	4.054	-4.000	1.00	0.00	N
ATOM	3456	CA	GLN A	226	25.000	3.807	-3.450	1.00	0.00	C
ATOM	3457	C	GLN A	226	25.852	4.916	-4.001	1.00	0.00	C
ATOM	3458	O	GLN A	226	26.089	5.009	-5.087	1.00	0.00	O
ATOM	3472	N	ILE A	227	26.411	5.763	-3.177	1.00	0.00	N
ATOM	3473	CA	ILE A	227	27.132	6.880	-3.597	1.00	0.00	C
ATOM	3474	C	ILE A	227	28.558	6.544	-4.043	1.00	0.00	C
ATOM	3475	O	ILE A	227	28.966	6.747	-5.197	1.00	0.00	O
ATOM	3491	N	ASP A	228	29.257	5.980	-3.060	1.00	0.00	N
ATOM	3492	CA	ASP A	228	30.636	5.510	-3.178	1.00	0.00	C
ATOM	3493	C	ASP A	228	30.658	4.284	-2.291	1.00	0.00	C
ATOM	3494	O	ASP A	228	29.630	3.967	-1.698	1.00	0.00	O
ATOM	3503	N	PRO A	229	31.713	3.556	-2.192	1.00	0.00	N
ATOM	3504	CA	PRO A	229	31.693	2.320	-1.376	1.00	0.00	C
ATOM	3505	C	PRO A	229	31.456	2.595	0.112	1.00	0.00	C
ATOM	3506	O	PRO A	229	31.157	1.671	0.867	1.00	0.00	O
ATOM	3517	N	ASP A	230	31.636	3.852	0.537	1.00	0.00	N
ATOM	3518	CA	ASP A	230	31.489	4.208	1.943	1.00	0.00	C
ATOM	3519	C	ASP A	230	30.219	5.020	2.244	1.00	0.00	C
ATOM	3520	O	ASP A	230	30.035	5.439	3.384	1.00	0.00	O
ATOM	3529	N	ALA A	231	29.294	5.277	1.322	1.00	0.00	N
ATOM	3530	CA	ALA A	231	28.023	5.983	1.637	1.00	0.00	C
ATOM	3531	C	ALA A	231	26.885	5.482	0.733	1.00	0.00	C
ATOM	3532	O	ALA A	231	27.223	4.980	-0.357	1.00	0.00	O
ATOM	3539	N	CYS A	232	25.637	5.538	1.213	1.00	0.00	N
ATOM	3540	CA	CYS A	232	24.417	5.227	0.436	1.00	0.00	C
ATOM	3541	C	CYS A	232	23.279	6.209	0.820	1.00	0.00	C
ATOM	3542	O	CYS A	232	23.135	6.409	2.031	1.00	0.00	O
ATOM	3550	N	PHE A	233	22.553	6.768	-0.146	1.00	0.00	N
ATOM	3551	CA	PHE A	233	21.319	7.575	0.066	1.00	0.00	C
ATOM	3552	C	PHE A	233	20.076	6.748	-0.305	1.00	0.00	C
ATOM	3553	O	PHE A	233	20.126	5.941	-1.234	1.00	0.00	O
ATOM	3570	N	SER A	234	19.024	6.965	0.428	1.00	0.00	N
ATOM	3571	CA	SER A	234	17.831	6.180	0.721	1.00	0.00	C
ATOM	3572	C	SER A	234	16.665	7.211	0.777	1.00	0.00	C
ATOM	3573	O	SER A	234	16.717	7.932	1.771	1.00	0.00	O
ATOM	3581	N	ALA A	235	15.696	7.294	-0.149	1.00	0.00	N
ATOM	3582	CA	ALA A	235	14.668	8.372	-0.139	1.00	0.00	C
ATOM	3583	C	ALA A	235	13.471	8.194	-1.049	1.00	0.00	C

## Appendix D PDB files of the models for the closed state of VDAC

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ATOM	3584	O	ALA A 235	13.714	7.845	-2.214	1.00	0.00	O
ATOM	3591	N	LYS A 236	11.751	8.349	-0.061	1.00	0.00	N
ATOM	3592	CA	LYS A 236	11.059	7.182	-0.597	1.00	0.00	C
ATOM	3593	C	LYS A 236	9.547	7.342	-0.416	1.00	0.00	C
ATOM	3594	O	LYS A 236	9.111	8.051	0.497	1.00	0.00	O
ATOM	3613	N	VAL A 237	8.929	7.748	-1.310	1.00	0.00	N
ATOM	3614	CA	VAL A 237	7.739	8.647	-1.340	1.00	0.00	C
ATOM	3615	C	VAL A 237	6.488	7.804	-1.543	1.00	0.00	C
ATOM	3616	O	VAL A 237	6.289	7.294	-2.625	1.00	0.00	O
ATOM	3629	N	ASN A 238	5.637	7.651	-0.503	1.00	0.00	N
ATOM	3630	CA	ASN A 238	4.424	6.820	-0.618	1.00	0.00	C
ATOM	3631	C	ASN A 238	3.458	7.657	-1.385	1.00	0.00	C
ATOM	3632	O	ASN A 238	3.585	8.768	-1.605	1.00	0.00	O
ATOM	3643	N	ASN A 239	2.370	7.656	-1.940	1.00	0.00	N
ATOM	3644	CA	ASN A 239	1.404	8.476	-2.690	1.00	0.00	C
ATOM	3645	C	ASN A 239	0.425	9.203	-1.749	1.00	0.00	C
ATOM	3646	O	ASN A 239	0.015	10.306	-2.143	1.00	0.00	O
ATOM	3657	N	SER A 240	0.130	8.704	-0.550	1.00	0.00	N
ATOM	3658	CA	SER A 240	-0.500	9.510	0.516	1.00	0.00	C
ATOM	3659	C	SER A 240	0.435	10.615	1.049	1.00	0.00	C
ATOM	3660	O	SER A 240	-0.155	11.321	1.934	1.00	0.00	O
ATOM	3668	N	SER A 241	1.633	10.867	0.637	1.00	0.00	N
ATOM	3669	CA	SER A 241	2.517	12.058	0.566	1.00	0.00	C
ATOM	3670	C	SER A 241	3.598	12.040	1.669	1.00	0.00	C
ATOM	3671	O	SER A 241	3.939	13.096	2.167	1.00	0.00	O
ATOM	3679	N	LEU A 242	4.204	11.016	2.136	1.00	0.00	N
ATOM	3680	CA	LEU A 242	5.268	10.762	3.125	1.00	0.00	C
ATOM	3681	C	LEU A 242	6.582	10.579	2.359	1.00	0.00	C
ATOM	3682	O	LEU A 242	6.969	9.468	1.984	1.00	0.00	O
ATOM	3698	N	ILE A 243	7.279	11.704	2.117	1.00	0.00	N
ATOM	3699	CA	ILE A 243	8.571	11.714	1.389	1.00	0.00	C
ATOM	3700	C	ILE A 243	9.620	11.446	2.465	1.00	0.00	C
ATOM	3701	O	ILE A 243	9.950	12.317	3.262	1.00	0.00	O
ATOM	3717	N	GLY A 244	10.132	10.188	2.525	1.00	0.00	N
ATOM	3718	CA	GLY A 244	11.289	9.850	3.327	1.00	0.00	C
ATOM	3719	C	GLY A 244	12.554	10.456	2.716	1.00	0.00	C
ATOM	3720	O	GLY A 244	12.574	10.798	1.531	1.00	0.00	O
ATOM	3724	N	LEU A 245	13.598	10.527	3.531	1.00	0.00	N
ATOM	3725	CA	LEU A 245	14.996	10.724	3.108	1.00	0.00	C
ATOM	3726	C	LEU A 245	15.912	9.852	3.955	1.00	0.00	C
ATOM	3727	O	LEU A 245	15.668	9.720	5.159	1.00	0.00	O
ATOM	3743	N	GLY A 246	17.032	9.321	4.134	1.00	0.00	N
ATOM	3744	CA	GLY A 246	17.603	8.241	4.999	1.00	0.00	C
ATOM	3745	C	GLY A 246	19.137	8.080	4.954	1.00	0.00	C
ATOM	3746	O	GLY A 246	19.580	6.929	4.914	1.00	0.00	O
ATOM	3750	N	TYR A 247	19.933	9.148	5.047	1.00	0.00	N
ATOM	3751	CA	TYR A 247	21.306	9.176	4.487	1.00	0.00	C
ATOM	3752	C	TYR A 247	22.298	8.307	5.282	1.00	0.00	C
ATOM	3753	O	TYR A 247	22.475	8.635	6.453	1.00	0.00	O
ATOM	3771	N	THR A 248	22.879	7.258	4.694	1.00	0.00	N
ATOM	3772	CA	THR A 248	23.733	6.249	5.375	1.00	0.00	C
ATOM	3773	C	THR A 248	25.201	6.500	4.997	1.00	0.00	C
ATOM	3774	O	THR A 248	25.413	6.717	3.793	1.00	0.00	O
ATOM	3785	N	GLN A 249	26.119	6.464	5.956	1.00	0.00	N
ATOM	3786	CA	GLN A 249	27.560	6.692	5.798	1.00	0.00	C
ATOM	3787	C	GLN A 249	28.373	5.562	6.455	1.00	0.00	C
ATOM	3788	O	GLN A 249	28.690	5.662	7.639	1.00	0.00	O
ATOM	3802	N	THR A 250	28.771	4.553	5.695	1.00	0.00	N
ATOM	3803	CA	THR A 250	29.538	3.398	6.190	1.00	0.00	C
ATOM	3804	C	THR A 250	31.023	3.791	6.129	1.00	0.00	C
ATOM	3805	O	THR A 250	31.618	3.642	5.079	1.00	0.00	O
ATOM	3816	N	LEU A 251	31.662	4.155	7.236	1.00	0.00	N
ATOM	3817	CA	LEU A 251	33.087	4.516	7.175	1.00	0.00	C
ATOM	3818	C	LEU A 251	33.936	3.239	7.092	1.00	0.00	C
ATOM	3819	O	LEU A 251	33.404	2.251	7.653	1.00	0.00	O
ATOM	3835	N	LYS A 252	35.104	3.216	6.418	1.00	0.00	N
ATOM	3836	CA	LYS A 252	36.104	2.123	6.415	1.00	0.00	C
ATOM	3837	C	LYS A 252	36.418	1.782	7.873	1.00	0.00	C
ATOM	3838	O	LYS A 252	36.292	0.628	8.291	1.00	0.00	O
ATOM	3857	N	PRO A 253	36.806	2.763	8.660	1.00	0.00	N
ATOM	3858	CA	PRO A 253	37.097	2.561	10.105	1.00	0.00	C
ATOM	3859	C	PRO A 253	36.116	1.585	10.776	1.00	0.00	C
ATOM	3860	O	PRO A 253	36.391	1.078	11.859	1.00	0.00	O
ATOM	3871	N	GLY A 254	34.984	1.311	10.121	1.00	0.00	N
ATOM	3872	CA	GLY A 254	34.002	0.375	10.669	1.00	0.00	C
ATOM	3873	C	GLY A 254	32.898	1.079	11.454	1.00	0.00	C
ATOM	3874	O	GLY A 254	32.392	0.534	12.436	1.00	0.00	O
ATOM	3878	N	ILE A 255	32.423	2.240	11.038	1.00	0.00	N
ATOM	3879	CA	ILE A 255	31.413	3.064	11.764	1.00	0.00	C
ATOM	3880	C	ILE A 255	30.298	3.332	10.735	1.00	0.00	C
ATOM	3881	O	ILE A 255	30.693	3.814	9.658	1.00	0.00	O
ATOM	3897	N	LYS A 256	29.041	3.064	11.067	1.00	0.00	N
ATOM	3898	CA	LYS A 256	27.858	3.446	10.258	1.00	0.00	C
ATOM	3899	C	LYS A 256	27.187	4.624	10.971	1.00	0.00	C
ATOM	3900	O	LYS A 256	26.937	4.429	12.164	1.00	0.00	O
ATOM	3919	N	LEU A 257	26.966	5.726	10.305	1.00	0.00	N
ATOM	3920	CA	LEU A 257	26.176	6.885	10.762	1.00	0.00	C
ATOM	3921	C	LEU A 257	24.999	7.064	9.803	1.00	0.00	C
ATOM	3922	O	LEU A 257	25.165	7.720	8.768	1.00	0.00	O
ATOM	3938	N	THR A 258	23.856	6.459	10.144	1.00	0.00	N
ATOM	3939	CA	THR A 258	22.586	6.680	9.431	1.00	0.00	C
ATOM	3940	C	THR A 258	21.868	7.887	10.033	1.00	0.00	C
ATOM	3941	O	THR A 258	21.828	7.838	11.242	1.00	0.00	O
ATOM	3952	N	LEU A 259	21.422	8.852	9.267	1.00	0.00	N

ATOM	3953	CA	LEU	A	259	20.477	9.922	9.652	1.00	0.00	C
ATOM	3954	C	LEU	A	259	19.152	9.559	9.018	1.00	0.00	C
ATOM	3955	O	LEU	A	259	19.167	8.636	8.172	1.00	0.00	O
ATOM	3971	N	SER	A	260	18.061	10.401	9.564	1.00	0.00	N
ATOM	3972	CA	SER	A	260	16.930	10.312	8.633	1.00	0.00	C
ATOM	3973	C	SER	A	260	16.077	11.593	8.672	1.00	0.00	C
ATOM	3974	O	SER	A	260	16.285	12.453	9.541	1.00	0.00	O
ATOM	3982	N	ALA	A	261	15.133	11.684	7.739	1.00	0.00	N
ATOM	3983	CA	ALA	A	261	14.038	12.639	7.814	1.00	0.00	C
ATOM	3984	C	ALA	A	261	12.791	12.087	7.120	1.00	0.00	C
ATOM	3985	O	ALA	A	261	12.888	10.999	6.519	1.00	0.00	O
ATOM	3992	N	LEU	A	262	11.658	12.783	7.224	1.00	0.00	N
ATOM	3993	CA	LEU	A	262	10.390	12.256	6.706	1.00	0.00	C
ATOM	3994	C	LEU	A	262	9.407	13.416	6.541	1.00	0.00	C
ATOM	3995	O	LEU	A	262	8.555	13.694	7.375	1.00	0.00	O
ATOM	4011	N	LEU	A	263	9.557	14.137	5.431	1.00	0.00	N
ATOM	4012	CA	LEU	A	263	8.794	15.381	5.155	1.00	0.00	C
ATOM	4013	C	LEU	A	263	7.412	15.000	4.558	1.00	0.00	C
ATOM	4014	O	LEU	A	263	7.367	14.621	3.390	1.00	0.00	O
ATOM	4030	N	ASP	A	264	6.350	15.126	5.372	1.00	0.00	N
ATOM	4031	CA	ASP	A	264	5.032	14.569	5.069	1.00	0.00	C
ATOM	4032	C	ASP	A	264	4.146	15.683	4.518	1.00	0.00	C
ATOM	4033	O	ASP	A	264	3.163	16.057	5.046	1.00	0.00	O
ATOM	4042	N	GLY	A	265	3.832	15.460	3.333	1.00	0.00	N
ATOM	4043	CA	GLY	A	265	3.210	16.583	2.608	1.00	0.00	C
ATOM	4044	C	GLY	A	265	1.726	16.928	2.860	1.00	0.00	C
ATOM	4045	O	GLY	A	265	1.431	18.094	2.567	1.00	0.00	O
ATOM	4049	N	LYS	A	266	0.935	16.039	3.421	1.00	0.00	N
ATOM	4050	CA	LYS	A	266	-0.377	16.286	4.059	1.00	0.00	C
ATOM	4051	C	LYS	A	266	-0.222	16.526	5.557	1.00	0.00	C
ATOM	4052	O	LYS	A	266	-1.279	17.051	6.024	1.00	0.00	O
ATOM	4071	N	ASN	A	267	0.842	16.221	6.269	1.00	0.00	N
ATOM	4072	CA	ASN	A	267	1.108	16.562	7.639	1.00	0.00	C
ATOM	4073	C	ASN	A	267	2.485	17.225	7.859	1.00	0.00	C
ATOM	4074	O	ASN	A	267	2.925	17.112	8.985	1.00	0.00	O
ATOM	4085	N	VAL	A	268	3.147	17.943	6.907	1.00	0.00	N
ATOM	4086	CA	VAL	A	268	4.257	18.908	7.248	1.00	0.00	C
ATOM	4087	C	VAL	A	268	3.760	19.897	8.175	1.00	0.00	C
ATOM	4088	O	VAL	A	268	4.428	20.111	9.128	1.00	0.00	O
ATOM	4101	N	ASN	A	269	2.773	20.422	8.310	1.00	0.00	N
ATOM	4102	CA	ASN	A	269	2.203	21.354	9.253	1.00	0.00	C
ATOM	4103	C	ASN	A	269	1.574	20.600	10.448	1.00	0.00	C
ATOM	4104	O	ASN	A	269	1.499	21.354	11.440	1.00	0.00	O
ATOM	4115	N	ALA	A	270	1.211	19.331	10.429	1.00	0.00	N
ATOM	4116	CA	ALA	A	270	0.925	18.528	11.630	1.00	0.00	C
ATOM	4117	C	ALA	A	270	2.234	17.820	12.032	1.00	0.00	C
ATOM	4118	O	ALA	A	270	2.468	17.923	13.278	1.00	0.00	O
ATOM	4125	N	GLY	A	271	3.081	17.129	11.250	1.00	0.00	N
ATOM	4126	CA	GLY	A	271	4.344	16.676	11.781	1.00	0.00	C
ATOM	4127	C	GLY	A	271	5.303	16.117	10.746	1.00	0.00	C
ATOM	4128	O	GLY	A	271	5.363	16.642	9.669	1.00	0.00	O
ATOM	4132	N	GLY	A	272	6.020	15.100	11.105	1.00	0.00	N
ATOM	4133	CA	GLY	A	272	6.604	14.127	10.187	1.00	0.00	C
ATOM	4134	C	GLY	A	272	8.052	13.832	10.578	1.00	0.00	C
ATOM	4135	O	GLY	A	272	8.366	12.640	10.636	1.00	0.00	O
ATOM	4139	N	HIS	A	273	8.925	14.838	10.715	1.00	0.00	N
ATOM	4140	CA	HIS	A	273	10.349	14.658	10.416	1.00	0.00	C
ATOM	4141	C	HIS	A	273	11.090	13.842	11.466	1.00	0.00	C
ATOM	4142	O	HIS	A	273	11.005	14.122	12.607	1.00	0.00	O
ATOM	4156	N	LYS	A	274	11.861	12.859	11.055	1.00	0.00	N
ATOM	4157	CA	LYS	A	274	12.737	12.135	11.964	1.00	0.00	C
ATOM	4158	C	LYS	A	274	13.904	13.046	12.371	1.00	0.00	C
ATOM	4159	O	LYS	A	274	14.506	13.705	11.539	1.00	0.00	O
ATOM	4178	N	LEU	A	275	13.671	13.348	13.524	1.00	0.00	N
ATOM	4179	CA	LEU	A	275	14.819	13.476	14.407	1.00	0.00	C
ATOM	4180	C	LEU	A	275	15.224	12.068	14.866	1.00	0.00	C
ATOM	4181	O	LEU	A	275	14.565	11.464	15.703	1.00	0.00	O
ATOM	4197	N	GLY	A	276	16.249	11.457	14.457	1.00	0.00	N
ATOM	4198	CA	GLY	A	276	16.791	10.164	14.878	1.00	0.00	C
ATOM	4199	C	GLY	A	276	17.931	9.705	13.980	1.00	0.00	C
ATOM	4200	O	GLY	A	276	18.006	10.210	12.841	1.00	0.00	O
ATOM	4204	N	LEU	A	277	18.734	8.798	14.512	1.00	0.00	N
ATOM	4205	CA	LEU	A	277	19.997	8.306	13.926	1.00	0.00	C
ATOM	4206	C	LEU	A	277	20.042	6.780	14.029	1.00	0.00	C
ATOM	4207	O	LEU	A	277	19.278	6.277	14.866	1.00	0.00	O
ATOM	4223	N	GLY	A	278	20.889	6.129	13.259	1.00	0.00	N
ATOM	4224	CA	GLY	A	278	21.253	4.717	13.401	1.00	0.00	C
ATOM	4225	C	GLY	A	278	22.768	4.595	13.519	1.00	0.00	C
ATOM	4226	O	GLY	A	278	23.374	4.829	12.449	1.00	0.00	O
ATOM	4230	N	LEU	A	279	23.300	4.266	14.676	1.00	0.00	N
ATOM	4231	CA	LEU	A	279	24.733	4.331	14.999	1.00	0.00	C
ATOM	4232	C	LEU	A	279	25.206	2.909	15.258	1.00	0.00	C
ATOM	4233	O	LEU	A	279	24.658	2.381	16.255	1.00	0.00	O
ATOM	4249	N	GLU	A	280	26.076	2.337	14.435	1.00	0.00	N
ATOM	4250	CA	GLU	A	280	26.513	0.932	14.510	1.00	0.00	C
ATOM	4251	C	GLU	A	280	28.038	0.922	14.512	1.00	0.00	C
ATOM	4252	O	GLU	A	280	28.548	1.489	13.510	1.00	0.00	O
ATOM	4264	N	PHE	A	281	28.684	0.372	15.508	1.00	0.00	N
ATOM	4265	CA	PHE	A	281	30.142	0.394	15.701	1.00	0.00	C
ATOM	4266	C	PHE	A	281	30.687	-1.031	15.545	1.00	0.00	C
ATOM	4267	O	PHE	A	281	30.165	-1.913	16.262	1.00	0.00	O
ATOM	4284	N	GLN	A	282	31.669	-1.245	14.686	1.00	0.00	N
ATOM	4285	CA	GLN	A	282	32.364	-2.530	14.528	1.00	0.00	C
ATOM	4286	C	GLN	A	282	33.468	-2.630	15.577	1.00	0.00	C

## Appendix D PDB files of the models for the closed state of VDAC

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ATOM 4287 O GLN A 282      34.465 -1.913 15.520 1.00  0.00      O
ATOM 4301 N ALA A 283      33.032 -3.828 16.429 1.00  0.00      N
ATOM 4302 CA ALA A 283     33.939 -4.204 17.510 1.00  0.00      C
ATOM 4303 C ALA A 283      34.394 -2.978 18.296 1.00  0.00      C
ATOM 4304 O ALA A 283      33.940 -1.862 18.048 1.00  0.00      O
ATOM 4311 N ARG A 284      35.296 -3.200 19.251 1.00  0.00      N
ATOM 4312 CA ARG A 284     35.810 -2.112 20.078 1.00  0.00      C
ATOM 4313 C ARG A 284      36.936 -1.379 19.354 1.00  0.00      C
ATOM 4314 O ARG A 284      37.691 -1.982 18.592 1.00  0.00      O
ATOM 4335 N SER A 285      37.040 -0.074 19.602 1.00  0.00      N
ATOM 4336 CA SER A 285     38.077  0.739 18.969 1.00  0.00      C
ATOM 4337 C SER A 285      39.125  1.169 19.992 1.00  0.00      C
ATOM 4338 O SER A 285      39.658  2.255 19.842 1.00  0.00      O
END
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